

ISBN: 978-93-47587-68-9

THE LIVING WORLD

Contemporary Perspectives in Life Sciences

Volume II

Editors:

Dr. Mahesh Prakash Patil

Dr. Vijayalaxami Shelke Sarode

Mrs. Ashwini Chandrkant Patil

Dr. Alok Ranjan Sahu



BHUMI PUBLISHING, INDIA
FIRST EDITION: JULY 2026

The Living World: Contemporary Perspectives in Life Sciences Volume II

(ISBN: 978-93-47587-68-9)

DOI: <https://doi.org/10.5281/zenodo.21770620>

Editors

Dr. Mahesh Prakash Patil

Department of Microbiology,
R. C. Patel Arts, Commerce and Science
College, Shirpur, Maharashtra

Dr. Vijayalaxami Shelke Sarode

Department of Botany,
R. C. Patel Arts, Commerce and Science
College, Shirpur, Maharashtra

Mrs. Ashwini Chandrkant Patil

Department of Biotechnology,
R. C. Patel Arts, Commerce and Science
College, Shirpur, Maharashtra

Dr. Alok Ranjan Sahu

Department of Botany,
Vikash Degree College,
Bargarh, Odisha



July 2026

Copyright © Editors

Title: The Living World: Contemporary Perspectives in Life Sciences Volume II

Editors: Dr. Mahesh Prakash Patil, Mrs. Ashwini Chandrkant Patil,

Dr. Vijayalaxami Shelke Sarode, Dr. Alok Ranjan Sahu

First Edition: July 2026

ISBN: 978-93-47587-68-9



DOI: <https://doi.org/10.5281/zenodo.21770620>

All rights reserved. No part of this publication may be reproduced or transmitted, in any form or by any means, without permission. Any person who does any unauthorized act in relation to this publication may be liable to criminal prosecution and civil claims for damages.

Published by Bhumi Publishing,

a publishing unit of Bhumi Gramin Vikas Sanstha



Nigave Khalasa, Tal – Karveer, Dist – Kolhapur, Maharashtra, INDIA 416 207

E-mail: bhumipublishing@gmail.com



Disclaimer: The views expressed in the book are of the authors and not necessarily of the publisher and editors. Authors themselves are responsible for any kind of plagiarism found in their chapters and any related issues found with the book.

PREFACE

Life sciences have witnessed extraordinary growth over the past few decades, transforming our understanding of the diversity, complexity, and interconnectedness of living organisms. From molecular mechanisms to ecosystem dynamics, contemporary research continues to expand the frontiers of knowledge, offering innovative solutions to global challenges in health, agriculture, biodiversity conservation, and environmental sustainability. *The Living World: Contemporary Perspectives in Life Sciences* presents a collection of scholarly contributions that reflect the breadth and significance of modern biological research.

This volume brings together the work of researchers, academicians, and professionals from diverse disciplines within the life sciences. The chapters encompass a wide range of topics, including molecular biology, genetics, microbiology, biotechnology, zoology, botany, ecology, environmental science, physiology, bioinformatics, and emerging interdisciplinary fields. Collectively, these contributions demonstrate how scientific advancements are enhancing our understanding of life while addressing critical issues such as climate change, food security, disease management, and sustainable resource utilization.

The primary objective of this book is to provide readers with an up-to-date overview of recent developments in life sciences and to encourage interdisciplinary thinking that bridges basic research with real-world applications. By integrating theoretical knowledge with contemporary scientific discoveries, this volume serves as a valuable resource for undergraduate and postgraduate students, educators, researchers, healthcare professionals, and policymakers. It is our hope that the ideas presented in these chapters will stimulate scientific inquiry, inspire innovative research, and promote collaborative efforts across disciplines.

We extend our sincere gratitude to all the contributing authors for their valuable research and scholarly dedication. We also express our appreciation to the reviewers for their insightful comments and constructive suggestions, which have greatly improved the quality of this volume. Our heartfelt thanks are due to the editorial team and the publisher for their unwavering commitment and meticulous efforts in bringing this book to fruition.

We trust that *The Living World: Contemporary Perspectives in Life Sciences* will serve as a meaningful academic reference, inspire future discoveries, and contribute to the advancement of biological sciences for the benefit of society and the sustainable stewardship of our planet.

- Editors

TABLE OF CONTENT

Sr. No.	Book Chapter and Author(s)	Page No.
1.	NANOBIOLOGY: FROM FUNDAMENTALS TO BIOMEDICAL APPLICATIONS Isai M, Annadurai T, Nagaraj K and Subavathy P	1 – 10
2.	BIOCHEMICAL STUDIES OF TAPEWORM <i>SENGA</i> (DOLLFUS, 1934) IN FRESHWATER FISH <i>CHANNA PUNCTATUS</i> IN GODAVARI RIVER Kalpana D. Kaldate and Vasant K. Dongare	11 – 14
3.	WIDE HYBRIDIZATION: A PROMISING APPROACH FOR VEGETABLE CROP IMPROVEMENT Mamta Yadav, Jitendra Kumar Yadav and R. K. Narolia	15 – 27
4.	3D FOOD PRINTING OF FRUIT-BASED PRODUCTS: FORMULATION, PROCESSING AND FUTURE APPLICATIONS Priya Kumari and Neeraj Gupta	28 – 37
5.	FORMULATION AND EVALUATION OF A SKIN BARRIER- REPAIR CREAM FOR SENSITIVE SKIN USING CERAMIDES AND NATURAL OILS Surya Kumar Sahu, Pushpendra Kumar Sahu and Digeshwari Patel	38 – 53
6.	APPLICATIONS OF ARTIFICIAL INTELLIGENCE AND MACHINE LEARNING IN <i>IN VITRO</i> STUDIES OF IMPORTANT MEDICINAL PLANTS Sagar N. Bhagwat	54 – 61
7.	ANCIENT DNA AND ITS ROLE IN FORENSIC MICROBIOLOGY Sandhya R. Mulchandani and Pradnyesh Rane	62 – 72
8.	HYMENOPTERAN DIVERSITY AND AGRO-ECOSYSTEM SERVICES: POLLINATION MECHANICS, CROP YIELD ENHANCEMENT, AND NATURAL PEST SUPPRESSION IN TROPICAL AGRO-LANDSCAPES Vasant D. Jadhav	73 – 87
9.	ECOLOGICAL SERVICES OF RAIGAD DISTRICT, MAHARASHTRA: A COMPREHENSIVE REVIEW Shaheena Sarwat Mirza	88 – 93

10.	FUNDAMENTALS OF BIOCHEMISTRY: MOLECULES AND REACTIONS OF LIFE Raj Kumar Singh Bharti, Anoj Kumar, Ravi Kumar, Yogesh Kumar and Anil Kumar	94 – 104
11.	MEDICINAL PROPERTIES AND THERAPEUTIC POTENTIAL OF INVASIVE PLANT SPECIES Sandhya R. Mulchandani and Pradnyesh Rane	105 – 114
12.	RECENT DEVELOPMENTS IN MATHEMATICAL MODELLING OF BIOLOGICAL SYSTEMS Adhil Basha D, P Balaganesan and R Sengothai	115 – 131
13.	CLIMATE CHANGE AND BIODIVERSITY CONSERVATION: CONTEMPORARY CHALLENGES FOR THE LIVING WORLD Vivek Kumar and Sorabh	132 – 143

NANOBIOLOGY: FROM FUNDAMENTALS TO BIOMEDICAL APPLICATIONS

Isai M^{*1}, Annadurai T², Nagaraj K³ and Subavathy P⁴

¹Department of Zoology, Seethalakshmi Ramaswami College (Autonomous),
Affiliated to Bharathidasan University, Tiruchirappalli, Tamilnadu

²Centre for Excellence in Nanobio Translational Research,
Department of Pharmaceutical Technology, University college of Engineering,
Anna University- BIT campus, Tiruchirappalli - 620024, Tamilnadu

³Biomedical & Nano-Drug Formulation Laboratory, Department of General Medicine
Saveetha Medical College and Hospital, Saveetha Institute of Medical and Technical
Sciences (SIMATS), Chennai-602105, Tamilnadu

⁴PG and Research Department of Zoology,
St. Mary's College (Autonomous), Thoothukudi, Tamilnadu

*Corresponding author E-mail: mathivananisai@gmail.com

Abstract

Nanobiology applies the principles of nanoscience to biological structures and processes that naturally occur at the nanometer scale, positioning it at the intersection of materials science, chemistry, and the life sciences. This chapter surveys the field's conceptual foundations, beginning with the size-dependent physical and chemical properties that distinguish nanoscale materials from their bulk counterparts, and the nano-bio interface formed when engineered nanomaterials contact physiological environments. It then reviews the principal classes of biomedical nanomaterials—lipid-based, polymeric, inorganic, carbon-based, and biomimetic—along with the characterization techniques required to assess their size, surface chemistry, and stability prior to biological use. The chapter examines representative biomedical applications, including nanoparticle-mediated drug delivery, cancer nanomedicine, molecular imaging, point-of-care diagnostics, tissue engineering, and nucleic acid therapeutics such as lipid nanoparticle-based mRNA vaccines. Particular attention is given to the translational pathway from synthesis through systemic circulation, cellular uptake, and intracellular target engagement, along with the toxicological and regulatory considerations that arise from nanomaterials' distinctive biological interactions. The chapter concludes by identifying persistent challenges in manufacturing reproducibility and clinical translation, alongside emerging directions in stimuli-responsive and biomimetic nanocarriers that are likely to shape the next generation of nanobiology-driven biomedical innovation.

Keywords: Nanobiology, Nanomedicine, Nanoparticle Drug Delivery, Biomedical Nanomaterials, Nanotoxicology.

Introduction

Nanobiology occupies the intersection of the physical, chemical, and life sciences, applying the principles of nanoscience to biological structures and processes that naturally occur at the nanometer scale. Proteins, nucleic acids, membrane channels, and viral capsids are themselves nanoscale machines, which makes biology a natural domain for nanotechnology to inform and, in turn, be informed by. As Whitesides (2003) observed, the convergence of engineered nanomaterials with biological systems is productive precisely because both operate within the same one-to-one-hundred-nanometer regime, where molecular recognition, self-assembly, and surface chemistry dominate over bulk material behavior.

The emergence of nanobiology as a distinct field reflects two parallel developments: advances in the synthesis and characterization of engineered nanomaterials, and a deepening understanding of the nanoscale architecture of cells and biomolecules. Together these developments have enabled a generation of biomedical technologies, including targeted drug carriers, ultrasensitive diagnostic platforms, and regenerative scaffolds, that were not achievable with conventional micro- or macro-scale materials (Jain, 2008). This chapter surveys the conceptual foundations of nanobiology, introduces the major classes of biomedical nanomaterials, reviews key characterization approaches, and examines representative applications in drug delivery, diagnostics, and tissue engineering. The chapter closes with a discussion of toxicological and translational challenges that continue to shape the field's trajectory.

Interest in nanobiology has also been driven by clinical necessity. Many promising drug candidates, particularly biologics and nucleic acid therapeutics, are poorly stable in circulation, are rapidly cleared by the kidneys, or cannot cross cellular membranes without assistance. Conventional formulation strategies often proved inadequate to these challenges, motivating the search for carrier systems capable of protecting a therapeutic payload, controlling its release, and directing it toward a specific tissue or cell type. Nanomaterials answered this need in a way that few other technologies could, and the resulting formulations have progressed from laboratory curiosities to approved medicines used in oncology, infectious disease, and, most visibly, in messenger RNA vaccines (Farokhzad & Langer, 2009). The scope of nanobiology has consequently expanded well beyond its origins in materials chemistry to encompass cell biology, immunology, pharmacology, and clinical medicine, making it one of the more thoroughly interdisciplinary domains within the broader biomedical sciences.

Fundamental Principles of Nanobiology

The defining feature of the nanoscale is not simply smallness but the emergence of size-dependent physical and chemical properties. As particle dimensions approach the nanometer

range, the ratio of surface area to volume increases dramatically, and quantum confinement effects begin to influence optical, electronic, and magnetic behavior. These effects explain why gold nanoparticles exhibit tunable colors depending on diameter, why semiconductor quantum dots fluoresce at wavelengths determined by particle size, and why nanoscale iron oxide behaves superparamagnetically rather than ferromagnetically (Sanvicens & Marco, 2008). For nanobiology, this size dependence is functionally important because it allows engineered particles to be tuned so that their physical behavior matches a specific biological task, such as penetrating a tumor vasculature with characteristic pore sizes or crossing a cell membrane of a defined thickness.

Figure 1 situates engineered nanomaterials alongside biological entities of comparable dimension. Nanoparticles used as drug carriers typically fall in the same size range as viruses and large multiprotein complexes, while individual biomolecules such as glucose or DNA occupy the lower end of the nanoscale. This dimensional overlap is not incidental; it is what allows nanoparticles to interact with cellular machinery, biomolecular receptors, and transport pathways using the same size-dependent rules that govern natural nanoscale biology (Whitesides, 2003).

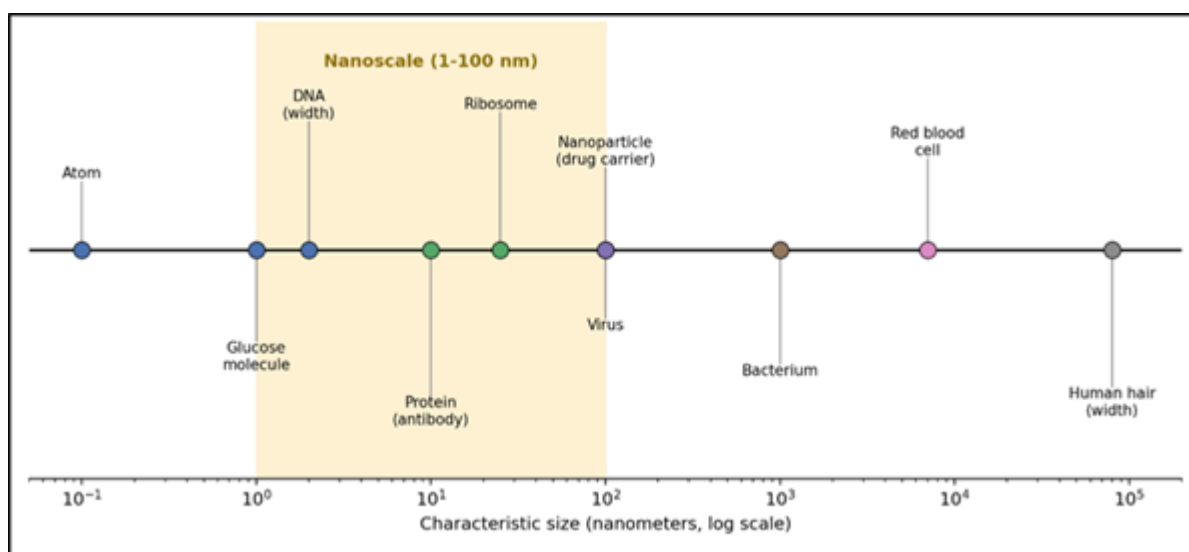


Figure 1: Comparative size scale of biological entities and engineered nanomaterials, illustrating the dimensional overlap that allows nanoparticles to interact with cellular and molecular machinery

A second organizing principle is the nano-bio interface, the dynamic boundary formed when an engineered nanomaterial contacts a biological environment. Upon entering a physiological fluid, a nanoparticle surface is rapidly coated by proteins, lipids, and other biomolecules to form what is commonly termed a corona. This corona, rather than the pristine engineered surface, is what cells and tissues actually encounter, and it substantially determines biodistribution, immune

recognition, and cellular uptake (Nel *et al.*, 2009). Understanding and controlling the nano-bio interface—through surface functionalization, charge modulation, and hydrophilic coatings such as polyethylene glycol—has therefore become a central design consideration in nanobiology.

Classes of Biomedical Nanomaterials

Biomedical nanomaterials are typically grouped by composition and structural organization rather than by application, since a single material class, such as a lipid nanoparticle, can be adapted for drug delivery, imaging, or vaccine formulation depending on cargo and surface design. Table 1 summarizes the principal classes of nanomaterials used in biomedicine, along with their characteristic size ranges and representative biomedical roles.

Lipid-based nanocarriers, including liposomes and solid lipid nanoparticles, remain among the most clinically advanced systems because their amphiphilic bilayer structure closely mimics cell membranes, conferring biocompatibility and enabling encapsulation of both hydrophilic and hydrophobic cargo (Peer *et al.*, 2007). Polymeric nanoparticles, formed from biodegradable polymers such as poly(lactic-co-glycolic acid), offer tunable degradation kinetics and sustained release profiles. Inorganic nanomaterials, such as gold nanoparticles, iron oxide nanoparticles, and quantum dots, contribute distinctive optical, magnetic, or electronic properties that are exploited primarily for imaging and biosensing rather than drug loading. Carbon-based nanomaterials, including carbon nanotubes and graphene derivatives, provide high surface area and unique electrical conductivity, useful in biosensor and scaffold applications. Finally, biological and biomimetic nanostructures, such as viral capsids, exosomes, and protein cages, exploit naturally evolved self-assembly and targeting machinery, often reducing immunogenicity relative to fully synthetic carriers (Mitragotri *et al.*, 2015).

The choice among these material classes in practice depends on the intended clinical function rather than on any single class being categorically superior. Lipid nanoparticles are generally favored when the cargo is a nucleic acid requiring cytosolic delivery, since ionizable lipid components can be tuned to promote endosomal escape at acidic pH. Polymeric systems are often preferred when sustained, weeks-long drug release is desired, since the degradation rate of the polymer backbone can be engineered independently of the encapsulated drug. Inorganic and carbon-based nanomaterials, by contrast, are rarely used as the sole delivery vehicle in approved products, but instead serve diagnostic, imaging, or biosensing roles where their optical or electronic properties, rather than their capacity to carry a drug payload, are the primary asset. This distinction between delivery-oriented and signal-oriented nanomaterials is a useful organizing heuristic when comparing across the many nanomaterial platforms reported in the literature.

Table 1: Principal Classes of Biomedical Nanomaterials

Nanomaterial Class	Typical Size Range	Representative Examples	Primary Biomedical Role
Lipid-based nanocarriers	50-200 nm	Liposomes, solid lipid nanoparticles, lipid nanoparticles	Drug and mRNA delivery, vaccine formulation
Polymeric nanoparticles	10-200 nm	PLGA, PLA, chitosan nanoparticles	Sustained-release drug delivery
Inorganic nanoparticles	1-100 nm	Gold nanoparticles, iron oxide nanoparticles	Imaging contrast, photothermal therapy
Quantum dots	2-10 nm	Cadmium selenide, indium phosphide dots	Multiplexed fluorescence imaging
Carbon-based nanomaterials	1-100 nm	Carbon nanotubes, graphene oxide	Biosensing, scaffold reinforcement
Biomimetic nanostructures	30-150 nm	Exosomes, viral capsids, protein cages	Targeted delivery, low immunogenicity

Note. Size ranges and examples are representative and reflect commonly reported values in the literature (Peer et al., 2007; Sanvicens & Marco, 2008).

Characterization Techniques in Nanobiology

Reliable biomedical application of nanomaterials depends on rigorous physicochemical characterization, since size, shape, surface charge, and surface chemistry all influence biological fate. Dynamic light scattering and nanoparticle tracking analysis are routinely used to determine hydrodynamic diameter and polydispersity in solution, while electron microscopy techniques, including transmission and scanning electron microscopy, provide direct visualization of particle morphology at sub-nanometer resolution. Surface charge is commonly assessed through zeta potential measurements, which correlate with colloidal stability and interaction with charged cell membrane components.

Beyond physical characterization, techniques such as atomic force microscopy allow researchers to probe mechanical properties and surface topography under near-physiological conditions, which is particularly relevant for understanding how nanomaterials interact with cell membranes and the protein corona described earlier (Nel *et al.*, 2009). Spectroscopic methods, including ultraviolet-visible and fluorescence spectroscopy, are used both for compositional analysis and for tracking nanoparticle behavior in biological media over time. The combination of these

techniques provides the multidimensional characterization profile required before a nanomaterial can reasonably proceed toward preclinical biomedical testing.

Biomedical Applications of Nanobiology

The biomedical applications of nanobiology span therapeutics, diagnostics, and regenerative medicine, with drug delivery representing the most mature translational area. Nanoparticle carriers can improve the pharmacokinetics of poorly soluble or rapidly degraded drugs, reduce off-target toxicity, and, in the case of solid tumors, exploit the enhanced permeability and retention effect to preferentially accumulate in leaky tumor vasculature (Farokhzad & Langer, 2009). Figure 2 outlines a representative pathway for nanoparticle-mediated drug delivery, beginning with synthesis and surface functionalization and proceeding through systemic circulation, cellular uptake, intracellular release, and target engagement. Each stage in this pathway represents a potential point of failure or optimization; for example, insufficient endosomal escape can trap a therapeutic payload before it reaches its intracellular target, motivating the design of pH-responsive or membrane-disruptive surface chemistries (Shi *et al.*, 2017).

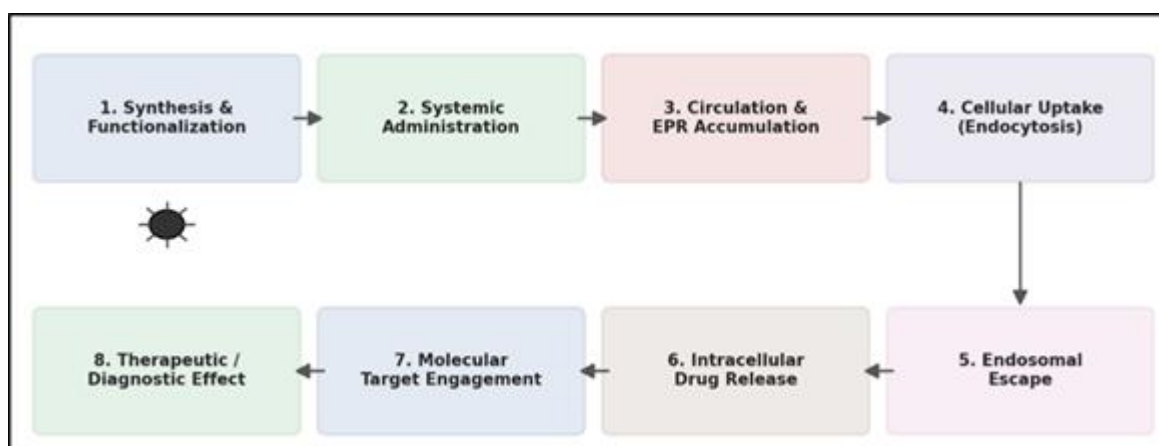


Figure 2: Schematic pathway of nanoparticle-mediated drug delivery, from synthesis and functionalization through systemic circulation, cellular uptake, intracellular release, and therapeutic target engagement

Cancer nanomedicine illustrates the translational promise and complexity of this approach most clearly. Formulations such as liposomal doxorubicin were among the first nanoparticle therapeutics to reach clinical use, and subsequent generations have incorporated active targeting ligands, such as antibodies or aptamers, to bind selectively to receptors overexpressed on tumor cells (Peer *et al.*, 2007). Nonetheless, clinical translation has proven more difficult than early preclinical results suggested, in part because tumor heterogeneity and variable enhanced

permeability and retention across patients limit the generalizability of passive targeting strategies (Shi *et al.*, 2017).

In diagnostics and imaging, nanomaterials provide contrast and signal amplification not achievable with conventional agents. Superparamagnetic iron oxide nanoparticles enhance magnetic resonance imaging contrast, gold nanoparticles enable colorimetric and surface-enhanced Raman-based detection assays, and semiconductor quantum dots support multiplexed fluorescence imaging due to their narrow, size-tunable emission spectra. These properties have been particularly valuable in point-of-care diagnostic devices, where nanoparticle-based lateral flow assays allow rapid, low-cost detection of biomarkers or pathogens outside a centralized laboratory setting (Jain, 2008).

Table 2: Representative Biomedical Applications of Nanobiology

Application Domain	Common Nanomaterials Used	Underlying Mechanism
Cancer therapeutics	Liposomes, polymeric nanoparticles	Enhanced permeability and retention, active receptor targeting
Molecular imaging	Iron oxide nanoparticles, quantum dots	Magnetic contrast enhancement, size-tunable fluorescence
Point-of-care diagnostics	Gold nanoparticles	Colorimetric and surface-enhanced Raman signal generation
Tissue engineering	Electrospun nanofibers, peptide hydrogels	Extracellular-matrix-mimetic topography and biochemical cues
Biosensing	Carbon nanotubes, nanowires	Single-molecule electrical or optical signal transduction
Gene and RNA therapy	Lipid nanoparticles	Endosomal encapsulation and cytosolic release of nucleic acids

Note. Mechanisms are simplified for illustrative purposes; individual formulations may combine multiple mechanisms (Farokhzad & Langer, 2009; Shi et al., 2017).

Beyond oncology, lipid nanoparticle delivery of messenger RNA has become one of the most consequential recent applications of nanobiology, providing the delivery platform behind large-scale vaccination efforts against infectious disease. In this context, the nanoparticle's role is to protect a fragile RNA molecule from enzymatic degradation, facilitate its uptake by antigen-presenting cells, and promote cytosolic release so that the encoded antigen can be translated and presented to the immune system. The rapid clinical success of this platform has reinforced the

broader translational logic of nanobiology: a well-characterized, tunable delivery vehicle can often be paired with different biological cargos to address distinct disease targets without redesigning the entire system from first principles (Mitragotri *et al.*, 2015).

Nanobiology has also informed tissue engineering and regenerative medicine, where nanostructured scaffolds are designed to mimic the nanoscale topography and biochemical cues of the native extracellular matrix. Electrospun nanofibers, self-assembling peptide hydrogels, and nanoparticle-functionalized scaffolds can guide stem cell differentiation and promote vascularization, addressing longstanding challenges in engineering functional replacement tissue. Similarly, nanoscale biosensors constructed from carbon nanotubes or nanowires have achieved single-molecule sensitivity for detecting nucleic acids, proteins, and metabolic biomarkers, supporting applications from continuous glucose monitoring to early cancer detection. Table 2 summarizes representative biomedical application domains, the nanomaterial classes most commonly employed, and the underlying mechanism of action for each.

Toxicology, Safety, and Regulatory Considerations

The same properties that make nanomaterials biomedically useful, such as high surface reactivity and the ability to cross biological barriers, also raise distinct toxicological concerns not fully captured by conventional chemical risk assessment. Nel *et al.* (2009) argued that biophysicochemical interactions at the nano-bio interface, including protein corona formation, oxidative stress induction, and interference with normal cellular signaling, must be evaluated systematically rather than assumed to scale predictably from bulk material toxicology. Nanoparticle biodistribution studies have shown that clearance pathways, accumulation in organs such as the liver and spleen, and long-term biopersistence vary considerably with size, surface charge, and coating chemistry, complicating generalized safety predictions across nanomaterial classes.

Regulatory frameworks have struggled to keep pace with the diversity of nanomaterial formulations entering preclinical and clinical development. Because a nanomedicine's identity is defined jointly by its core material, surface chemistry, and manufacturing process, comparatively minor formulation changes can meaningfully alter pharmacokinetics and toxicity, which limits the applicability of generic material safety classifications (Mitragotri *et al.*, 2015). Addressing this gap has required closer collaboration between materials scientists, toxicologists, and regulatory agencies to establish characterization standards and reproducible manufacturing protocols suitable for clinical translation.

Challenges and Future Directions

Despite substantial preclinical progress, the number of nanomedicines that have achieved regulatory approval remains modest relative to the volume of published research, underscoring a persistent translational gap (Shi *et al.*, 2017). Key barriers include batch-to-batch manufacturing variability at nanoscale precision, incomplete understanding of how patient-specific biological variability affects nanoparticle behavior, and the high cost of the extensive characterization required to satisfy regulatory expectations. Addressing these barriers will likely require standardized characterization pipelines, more predictive preclinical models that better recapitulate human physiology, and closer integration between materials scientists and clinicians early in the design process (Mitragotri *et al.*, 2015).

Several converging trends are likely to shape the next phase of nanobiology. Advances in stimuli-responsive materials are enabling nanocarriers that release cargo selectively in response to local pH, enzymatic activity, or externally applied fields, improving spatial and temporal control over therapeutic action. Integration of nanomaterials with gene-editing and RNA-based therapeutics, exemplified by lipid nanoparticle delivery of messenger RNA, has already demonstrated large-scale clinical impact and is likely to expand into additional disease areas. Finally, growing interest in biomimetic and cell-derived nanocarriers, such as exosomes, reflects an effort to combine the tunability of synthetic nanomaterials with the natural targeting and immune-evasion properties of biologically derived structures (Mitragotri *et al.*, 2015).

Conclusion

Nanobiology has matured from a conceptual extension of nanotechnology into a distinct interdisciplinary field with demonstrated clinical impact, particularly in drug delivery, diagnostic imaging, and, more recently, nucleic acid therapeutics. Its central insight, that engineering matter at the same scale as natural biomolecular machinery enables new modes of biological interaction, continues to generate biomedical innovations that were not previously feasible. At the same time, the field's continued growth depends on resolving persistent challenges in toxicology, manufacturing reproducibility, and regulatory standardization. As characterization tools and translational frameworks mature alongside the materials themselves, nanobiology is well positioned to remain a central driver of biomedical innovation over the coming decade.

References

1. Farokhzad, O. C., & Langer, R. (2009). Impact of nanotechnology on drug delivery. *ACS Nano*, 3(1), 16–20. <https://doi.org/10.1021/nn900002m>
2. Jain, K. K. (2008). Nanomedicine: Application of nanobiotechnology in medical practice. *Medical Principles and Practice*, 17(2), 89–101. <https://doi.org/10.1159/000112961>

3. Mitragotri, S., Anderson, D. G., Chen, X., Chow, E. K., Ho, D., Kabanov, A. V., Karp, J. M., Kataoka, K., Mirkin, C. A., Petrosko, S. H., Shi, J., Stevens, M. M., Sun, S., Teoh, S., Venkatraman, S. S., Xia, Y., Wang, S., Gu, Z., & Xu, C. (2015). Accelerating the translation of nanomaterials in biomedicine. *ACS Nano*, 9(7), 6644–6654. <https://doi.org/10.1021/acsnano.5b03569>
4. Nel, A. E., Mädler, L., Velegol, D., Xia, T., Hoek, E. M. V., Somasundaran, P., Klaessig, F., Castranova, V., & Thompson, M. (2009). Understanding biophysicochemical interactions at the nano-bio interface. *Nature Materials*, 8(7), 543–557. <https://doi.org/10.1038/nmat2442>
5. Peer, D., Karp, J. M., Hong, S., Farokhzad, O. C., Margalit, R., & Langer, R. (2007). Nanocarriers as an emerging platform for cancer therapy. *Nature Nanotechnology*, 2(12), 751–760. <https://doi.org/10.1038/nnano.2007.387>
6. Sanvicens, N., & Marco, M. P. (2008). Multifunctional nanoparticles: Properties and prospects for their use in human medicine. *Trends in Biotechnology*, 26(8), 425–433. <https://doi.org/10.1016/j.tibtech.2008.04.005>
7. Shi, J., Kantoff, P. W., Wooster, R., & Farokhzad, O. C. (2017). Cancer nanomedicine: Progress, challenges and opportunities. *Nature Reviews Cancer*, 17(1), 20–37. <https://doi.org/10.1038/nrc.2016.108>
8. Whitesides, G. M. (2003). The “right” size in nanobiotechnology. *Nature Biotechnology*, 21(10), 1161–1165. <https://doi.org/10.1038/nbt872>

BIOCHEMICAL STUDIES OF TAPEWORM *SENGA* (DOLLFUS, 1934) IN FRESHWATER FISH *CHANNA PUNCTATUS* IN GODAVARI RIVER

Kalpana D. Kaldate¹ and Vasant K. Dongare^{*2}

¹Department. of Zoology, KTHM College, Nashik, M.S.

²Department of Zoology,

Sundarrao More Arts, Commerce and Science College, Poladpur-Raigad, M.S.

*Corresponding author E-mail: drvkdongre@gmail.com

Abstract

Parasitic helminths are widely distributed in freshwater fishes and often influence the physiological and biochemical status of their hosts. The present investigation deals with biochemical studies of the cestode parasite *Senga* (Dollfus, 1934) collected from the intestine of freshwater fish *Channa punctatus* (Bloch, 1793). Fish samples were collected from the Godavari River. The parasite was isolated and subjected to biochemical analysis including estimation of proteins, lipids, and glycogen. Comparative biochemical analysis between host tissue and parasite revealed variations in metabolic composition. Author noticed that concentration of protein, glycogen, and lipids were notably high in the non-infected region compared to the infected section within the host intestine.

Keywords: Cestode, *Senga*, *Channa punctatus*, Biochemical Analysis, Godavari River.

Introduction

Fish are often referred to as “gold in the water” because of their immense economic value and their vital contribution as a source of high-quality protein for the growing global population. They play an important role in human nutrition and the economy, particularly in developing countries where fish are a staple source of animal protein. However, fish populations are frequently affected by parasitic infections that can lead to reduced growth, poor health, and economic losses in aquaculture (Sharma *et al.*, 2012). Among these parasites, cestodes (tapeworms) are of significant concern due to their pathogenicity and ability to cause physiological and biochemical alterations in their hosts. (Diksha Galte, 2024). Comparative biochemical analyses between parasites and their host tissues can reveal how parasites modify host metabolism to meet their nutritional demands (Fairbairn *et al.*, 1961; Barrett, 1981).

Material and Methods

The worms were collected from the intestine of fresh water fish *Channa Puntatus* and then washed with distilled water. Collected worms were then dried on the blotting paper to remove

excess water and transferred to watch glass and weight on sensitive balance. After 50-60 C for 24 hours, the dry weight was taken. Glycogen content was estimated using the Anthrone method (Kemp *et al.*, 1954), protein content was estimated using the Lowry method (Lowry *et al.*, 1951), and lipid content was estimated using the Folch extraction method (Folch *et al.*, 1957). Biochemical constituents were estimated from fish muscle tissues using standard biochemical techniques.

Table 1: Biochemical estimation of fresh water fish *Channa punctatus* (Bloch, 1793) intestine and cestode parasite *Senga* (Dollfus, 1934)

Parameters	<i>Senga</i> (Dollfus, 1934)	Intestinal tissue of <i>Channa punctatus</i>	
		Infected	Non- Infected
Protein	18.32	20	23
Glycogen	26	23	21
Lipid	23.05	18.34	19.05

Values expressed in mg/100mg dry wt. of tissue per ml solution

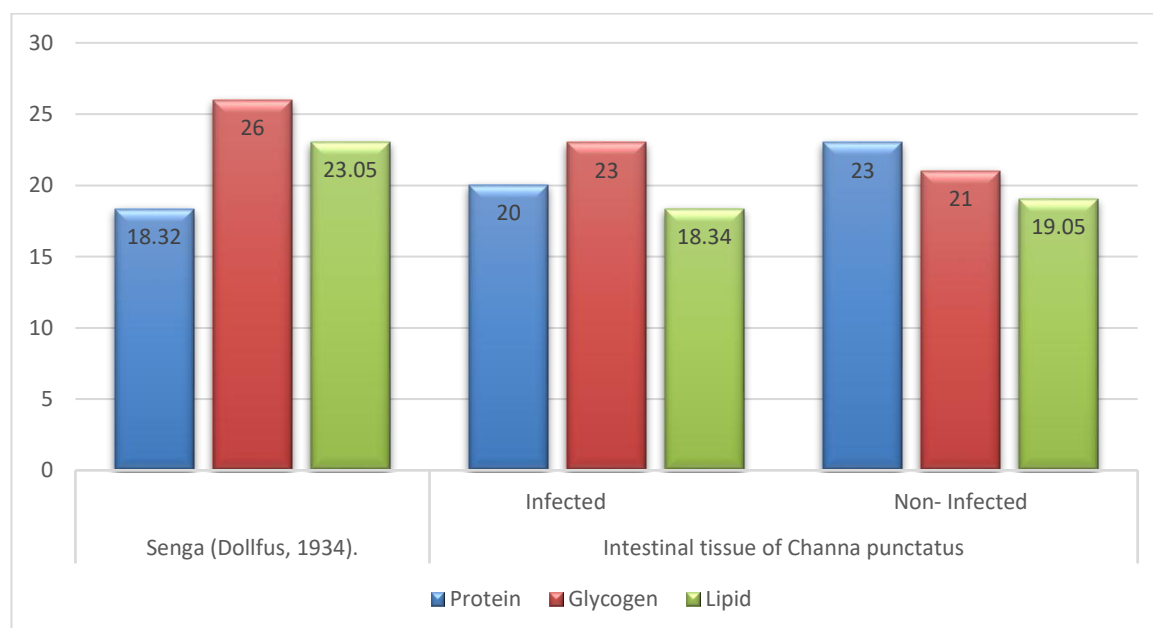


Figure 1: Biochemical estimation of fresh water fish *Channa punctatus* (Bloch, 1793) intestine (Infected and Non infected) and cestode parasite *Senga* (Dollfus, 1934)

Results and Discussion

The present investigation revealed significant biochemical alterations in the intestinal tissue of *Channa punctatus* infected with *Senga*, indicating that cestode infection influences the metabolic status of the host. The reduction in protein content from 23 mg/g in non-infected tissue to 20

mg/g in infected tissue, along with the comparatively lower protein concentration in the parasite (18.32 mg/g), suggests that parasitic infestation adversely affects host protein metabolism. Such depletion is generally attributed to increased utilization of host proteins by parasites for their growth, development, and reproductive activities, as well as tissue damage caused by parasite attachment. Similar reductions in protein levels in helminth-infected fishes have been reported by Cheng (1986) and Roberts and Janovy (2009), who stated that parasitic infections alter host metabolic pathways and increase protein catabolism to meet the nutritional demands of the parasite.

An increase in glycogen content from 21 mg/g in non-infected tissue to 23 mg/g in infected tissue was observed, while the parasite exhibited the highest glycogen level (26 mg/g). This accumulation of glycogen in infected tissue may represent a physiological adaptation to parasitic stress or an alteration in carbohydrate metabolism resulting from impaired glycogen utilization. Parasites often rely on glycogen reserves as an important source of energy under anaerobic or low-oxygen conditions within the host intestine. Similar observations have been documented in cestode infections by Smyth (1994), who reported that glycogen serves as the principal energy reserve in tapeworms, enabling their survival and growth within the host environment. Barber (2007) also suggested that parasitic stress induces metabolic changes in fishes, including altered carbohydrate metabolism and energy allocation.

The lipid content of infected intestinal tissue (18.34 mg/g) was slightly lower than that of non-infected tissue (19.05 mg/g), whereas *Senga* contained a comparatively higher lipid concentration (23.05 mg/g). Lipids are essential for membrane formation, energy storage, and reproductive functions in helminths. The higher lipid concentration observed in the parasite suggests its efficient absorption and accumulation of host-derived lipids. Similar findings have been reported by Lumsden and Müller (1980), who emphasized that helminth parasites acquire lipids from their hosts because of their limited capacity for de novo lipid synthesis. The reduction in host lipid reserves may therefore reflect the diversion of lipids towards parasite growth and maintenance.

Overall, the biochemical changes observed in the present study demonstrate that *Senga* infection significantly modifies the nutritional and metabolic status of *Channa punctatus*. The depletion of proteins and lipids, coupled with glycogen accumulation, reflects the parasite's dependence on host nutrients and the physiological stress imposed on the host. These findings are in agreement with previous reports indicating that helminth infections adversely influence host metabolism, resulting in altered biochemical composition and reduced physiological performance (Cheng,

1986; Roberts & Janovy, 2009; Barber, 2007). Such metabolic disturbances may ultimately affect the health, growth, and survival of infected fishes in natural ecosystems.

Overall, the infection leads to significant biochemical changes in the host intestine, reflecting a parasite-host interaction where the parasite benefits at the expense of the host's nutritional status. These alterations may impair normal physiological functions of the intestine and reduce the overall health of *Channa punctatus*.

References

1. Folch, J., Lees, M., & Sloane-Stanley, G. H. (1957). The method of lipid estimation. *Journal of Biological Chemistry*, 228, 497.
2. Kemp, A., & Van Heijningen, A. K. (1954). A colorimetric micro-method for the determination of glycogen in tissues. *Biochemical Journal*, 56(4), 646–648.
3. Lowry, O. H., Rosebrough, N. J., Farr, A. L., & Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193(1), 265–275.
4. Brand, T. von. (1952). *Chemical physiology of endoparasitic animals*. Academic Press.
5. Sharma, P., Gupta, N., & Singh, R. (2012). Impact of helminth parasites on the health of freshwater fish. *Journal of Aquatic Biology*, 27(1), 45–52.
6. Fairbairn, D., et al. (1961). Carbohydrate metabolism in cestodes. *Canadian Journal of Zoology*, 39(2), 119–128.
7. Barrett, J. (1981). *Biochemistry of parasitic helminths*. Macmillan Publishers.
8. Galte, D. (2024). Biochemical examination of cestode parasite in fresh water fish *Mastacembelus armatus* from Osmanabad District (M.S.), India. *JETIR*, 11(11).
9. Barber, I. (2007). Parasites, behaviour and welfare in fish. *Applied Animal Behaviour Science*, 104(3–4), 251–264. <https://doi.org/10.1016/j.applanim.2006.09.005>
10. Cheng, T. C. (1986). *General parasitology* (2nd ed.). Academic Press.
11. Lumsden, R. D., & Müller, M. (1980). *Parasitic protozoa and helminths: Biochemistry and physiology*. Academic Press.
12. Roberts, L. S., & Janovy, J., Jr. (2009). *Foundations of parasitology* (8th ed.). McGraw-Hill Higher Education.
13. Smyth, J. D. (1994). *Introduction to animal parasitology* (3rd ed.). Cambridge University Press.
14. Williams, H., & Jones, A. (1994). *Parasitic worms of fish*. Taylor & Francis.

WIDE HYBRIDIZATION: A PROMISING APPROACH FOR VEGETABLE CROP IMPROVEMENT

Mamta Yadav^{*1}, Jitendra Kumar Yadav² and R. K. Narolia¹

¹Department of Horticulture,

Sri Karan Narendra Agriculture University, Jobner, Rajasthan 303329, India

²National Bureau of Plant Genetic Resources, IARI, Pusa Campus, New Delhi- 110012

*Corresponding author E-mail: mamtayadav5432@gmail.com

Abstract

Wide hybridization has become an indispensable breeding strategy for broadening the genetic base of vegetable crops through the transfer of valuable genes from related wild species, distant species, or genera into cultivated backgrounds. The continuous narrowing of genetic diversity due to domestication and intensive selection has limited the availability of novel alleles required to combat emerging biotic and abiotic stresses. Crop wild relatives represent a rich reservoir of genes governing disease and insect resistance, tolerance to drought, salinity, heat, cold, and other environmental stresses, as well as traits related to yield, nutritional quality, and market acceptability. Wide hybridization enables the introgression of these desirable traits into elite cultivars, thereby accelerating the development of improved vegetable varieties. However, successful distant hybridization is often constrained by pre- and post-fertilization barriers, including pollen–pistil incompatibility, embryo abortion, hybrid sterility, chromosomal imbalance, and linkage drag. The integration of advanced breeding techniques such as embryo rescue, bridge crossing, chromosome doubling, protoplast fusion, marker-assisted selection, genomic tools, and genome editing has substantially enhanced the efficiency of wide hybridization and facilitated the recovery of fertile and agronomically superior hybrids. Significant achievements have been reported in tomato, potato, brinjal, onion, okra, cucurbits, and Brassica vegetables through the successful utilization of wild genetic resources for improving resistance to diseases, insect pests, and environmental stresses. Furthermore, recent advances in genomics, high-throughput sequencing, speed breeding, and CRISPR-based genome editing have strengthened the application of wide hybridization in modern vegetable breeding programmes. This chapter discusses the principles, historical development, objectives, types, reproductive barriers, enabling techniques, applications, limitations, representative case studies, and future prospects of wide hybridization, highlighting its pivotal role in developing climate-

resilient, high-yielding, and nutritionally superior vegetable cultivars for sustainable horticultural production.

Keywords: Distant Hybridization, Crop Wild Relatives, Vegetable Breeding, Introgression, Embryo Rescue, Disease Resistance.

Introduction

Vegetable crops are indispensable components of global agriculture owing to their significant contribution to food and nutritional security. They are rich sources of vitamins, minerals, dietary fibre, antioxidants, and bioactive compounds that are essential for maintaining human health. Besides their nutritional importance, vegetable crops generate substantial income for farmers and contribute to sustainable agricultural development. However, increasing population pressure, climate change, emerging pests and diseases, soil degradation, and shrinking natural resources have created new challenges for vegetable production. At the same time, prolonged domestication and intensive breeding have considerably narrowed the genetic diversity of cultivated vegetables, limiting the availability of useful genes for further crop improvement (Acquaah, 2020; Singh, 2021).

Crop wild relatives (CWRs) possess a vast reservoir of valuable genes governing resistance to diseases and insect pests, tolerance to abiotic stresses such as drought, salinity, heat, and cold, as well as improved nutritional quality and yield-related traits. The exploitation of these untapped genetic resources has become increasingly important for developing resilient and high-performing vegetable cultivars. Among the various breeding approaches, wide hybridization has emerged as an effective strategy for introducing desirable genes from genetically distant species or genera into cultivated crops, thereby broadening the genetic base and enhancing breeding efficiency (Hajjar & Hodgkin, 2007; Dempewolf *et al.*, 2017).

Wide hybridization, also known as distant hybridization, involves hybridization between individuals belonging to different species (interspecific hybridization), different genera (intergeneric hybridization), or, in rare instances, different families (interfamilial hybridization). This technique has played a crucial role in the genetic improvement of several vegetable crops, including tomato, potato, brinjal, onion, okra, cucumber, and Brassica vegetables, by facilitating the transfer of genes for resistance to biotic and abiotic stresses, quality enhancement, and adaptation to diverse environmental conditions (Sharma, 2017).

Although wide hybridization is often constrained by reproductive barriers such as pollen–pistil incompatibility, embryo abortion, hybrid sterility, and chromosomal imbalance, recent advances in plant biotechnology—including embryo rescue, chromosome doubling, somatic hybridization, molecular marker-assisted breeding, and genome editing—have significantly improved its

efficiency and practical utility. Consequently, wide hybridization has become an indispensable component of modern vegetable breeding programmes, providing sustainable solutions for developing climate-resilient, high-yielding, and nutritionally superior cultivars capable of meeting future agricultural demands (Prohens *et al.*, 2017; Warschefsky *et al.*, 2014).

Concept and Definition of Wide Hybridization

Hybridization is the process of crossing two genetically distinct individuals to combine desirable traits into a single progeny. When the parents belong to different species, genera, or, in exceptional cases, different families, the process is referred to as wide hybridization or distant hybridization. This breeding approach enables the transfer of valuable genes from crop wild relatives or allied species into cultivated varieties, thereby broadening the genetic base and creating novel genetic variability unavailable within the cultivated gene pool (Sharma, 2017; Singh, 2021).

Wide hybridization is broadly classified into interspecific, intergeneric, and interfamilial hybridization. Interspecific hybridization involves crosses between different species of the same genus, whereas intergeneric hybridization is performed between species belonging to different genera. Interfamilial hybridization, although rare due to substantial genetic divergence, has been achieved in a few cases using advanced biotechnological techniques. Compared with conventional hybridization, wide hybridization encounters greater reproductive barriers, including pollen–pistil incompatibility, embryo abortion, hybrid sterility, and chromosomal imbalance. Nevertheless, these obstacles can be overcome through techniques such as embryo rescue, chromosome doubling, bridge crossing, and somatic hybridization (Acquaah, 2020).

The importance of wide hybridization in vegetable breeding lies in its ability to exploit the rich genetic diversity present in wild relatives. It has been successfully employed to transfer genes for resistance to diseases and insect pests, tolerance to drought, salinity, heat, and cold, as well as traits related to yield, nutritional quality, and environmental adaptability. Consequently, wide hybridization has become an indispensable strategy in modern breeding programmes for developing climate-resilient, high-yielding, and sustainable vegetable cultivars capable of addressing future agricultural challenges (Hajjar & Hodgkin, 2007; Prohens *et al.*, 2017).

History and Evolution of Wide Hybridization

The concept of wide hybridization has evolved over the past three centuries and has become a cornerstone of modern plant breeding. The first documented artificial hybrid was produced in 1717 by Thomas Fairchild, who successfully crossed sweet william (*Dianthus barbatus*) with carnation (*Dianthus caryophyllus*), demonstrating the feasibility of hybridization between distinct plant species. A major breakthrough occurred in 1888, when Wilhelm Rimpau developed

the first fertile wheat–rye hybrid, laying the foundation for distant hybridization in crop improvement. Subsequently, in 1928, Georgii Karpechenko produced the intergeneric hybrid *Raphanobrassica* by crossing radish (*Raphanus sativus*) and cabbage (*Brassica oleracea*). Although the hybrid did not possess the desired commercial characteristics, it provided convincing evidence that chromosome doubling could restore fertility in distant hybrids (Singh, 2021).

The advancement of cytogenetics, tissue culture, and plant biotechnology during the twentieth century greatly expanded the scope of wide hybridization. Techniques such as embryo rescue, bridge crossing, chromosome doubling, and protoplast fusion enabled breeders to overcome reproductive barriers that previously limited gene transfer between distant taxa. In recent decades, the integration of molecular markers, genomic sequencing, and genome editing has further enhanced the efficiency of wide hybridization by facilitating precise introgression of beneficial genes from crop wild relatives. Today, wide hybridization serves as an indispensable breeding strategy for developing improved vegetable cultivars with enhanced resistance to biotic and abiotic stresses, superior quality, and greater adaptability to changing climatic conditions (Acquaah, 2020; Sharma, 2017; Prohens *et al.*, 2017).

Objectives of Wide Hybridization

The major objectives of wide hybridization are to:

- Broaden the genetic base of cultivated vegetable crops.
- Introduce desirable genes from crop wild relatives into cultivated species.
- Develop resistance to diseases and insect pests.
- Improve tolerance to abiotic stresses such as drought, salinity, heat, and cold.
- Enhance yield potential, quality, nutritional value, and shelf life.
- Generate novel genetic variability for crop improvement.
- Overcome the limitations of conventional breeding by utilizing distant genetic resources.
- Develop pre-breeding materials for future breeding programmes.
- Improve adaptability and stability of vegetable crops under changing climatic conditions.

Importance of Wide Hybridization

Wide hybridization plays a vital role in modern vegetable breeding by exploiting the rich genetic diversity present in crop wild relatives. It enables the transfer of beneficial genes that are absent in cultivated germplasm, thereby broadening the genetic base and enhancing breeding efficiency. This approach has been successfully used to improve resistance against diseases, insect pests, and various abiotic stresses while simultaneously enhancing yield, nutritional quality, and

horticultural performance. Furthermore, wide hybridization serves as an important pre-breeding tool for developing superior breeding lines and expanding the pool of useful genetic variation. When integrated with modern biotechnological approaches such as embryo rescue, molecular marker-assisted selection, and genome editing, wide hybridization provides an effective strategy for developing climate-resilient, high-yielding, and sustainable vegetable cultivars capable of meeting future agricultural challenges (Hajjar & Hodgkin, 2007; Sharma, 2017; Dempewolf *et al.*, 2017).

Types of Wide Hybridization

Wide hybridization is classified according to the taxonomic relationship between the parental species involved in the cross. Based on the degree of genetic divergence, it is broadly categorized into interspecific, intergeneric, and interfamilial hybridization. As the taxonomic distance between the parents increases, the complexity of hybridization also increases due to stronger reproductive and genetic barriers. Among these, interspecific hybridization is the most widely used and commercially successful in vegetable crop improvement, whereas intergeneric and interfamilial hybridization generally require advanced breeding and biotechnological interventions. Each type offers unique opportunities for broadening the genetic base and introducing valuable traits from wild or related taxa into cultivated vegetable crops (Singh, 2021; Sharma, 2017).

a) Interspecific Hybridization

Interspecific hybridization refers to the crossing of two genetically distinct species belonging to the same genus to combine desirable traits into a single genotype. It is the most widely practiced form of wide hybridization because closely related species generally exhibit greater genetic compatibility and produce more viable hybrids than intergeneric or interfamilial crosses. This approach has been extensively employed in vegetable breeding to transfer genes for disease and insect resistance, abiotic stress tolerance, improved yield, and enhanced nutritional quality from wild species to cultivated varieties. Successful examples include the transfer of resistance to tomato leaf curl virus, Fusarium wilt, and nematodes from wild *Solanum* species to cultivated tomato, as well as the incorporation of yellow vein mosaic virus resistance into okra through crosses involving wild *Abelmoschus* species. Thus, interspecific hybridization remains an indispensable strategy for broadening the genetic diversity of vegetable crops and developing improved cultivars (Singh, 2021; Acquah, 2020; Prohens *et al.*, 2017).

b) Intergeneric Hybridization

Intergeneric hybridization involves crossing two species belonging to different genera to introduce novel genetic variation and desirable traits into cultivated crops. Because of greater

genetic divergence between the parents, these crosses often face strong reproductive barriers, resulting in low hybrid viability and sterility. However, advances in plant biotechnology, including embryo rescue, chromosome doubling, and protoplast fusion, have significantly improved the success of intergeneric hybridization. One of the earliest and most notable examples is *Raphanobrassica*, developed by crossing radish (*Raphanus sativus*) with cabbage (*Brassica oleracea*), which demonstrated the feasibility of gene transfer between different genera. In vegetable breeding, intergeneric hybridization has been employed to introduce disease resistance, stress tolerance, and novel horticultural traits, thereby expanding the genetic resources available for crop improvement (Sharma, 2017; Singh, 2021).

c) Interfamilial Hybridization

Interfamilial hybridization refers to crosses between species belonging to different plant families. It represents the most challenging form of wide hybridization because of the extensive genetic divergence and strong reproductive barriers between the parental species. Successful interfamilial hybrids are extremely rare under natural conditions and generally require advanced biotechnological interventions such as embryo rescue, protoplast fusion, somatic hybridization, chromosome manipulation, and genetic transformation. Although commercially successful examples in vegetable crops are limited, interfamilial hybridization has considerable potential for introducing novel genes associated with disease resistance, abiotic stress tolerance, and quality traits. Continued advances in genomics and tissue culture techniques are expected to improve the feasibility of interfamilial gene transfer, thereby expanding the genetic resources available for future vegetable breeding programmes (Acquaah, 2020; Singh, 2021).

Barriers Associated with Wide Hybridization

Despite its immense potential in vegetable crop improvement, the successful application of wide hybridization is frequently constrained by several reproductive, genetic, and cytological barriers. These barriers reduce the efficiency of gene transfer between genetically distant species and often limit the production of viable and fertile hybrids. Depending on the stage at which they occur, hybridization barriers are broadly classified into pre-fertilization and post-fertilization barriers.

1. Pre-fertilization Barriers

Pre-fertilization barriers are reproductive obstacles that prevent successful fertilization before the formation of a zygote. These barriers arise due to pollen–stigma incompatibility, differences in flowering time, failure of pollen germination, inadequate pollen tube growth, and incompatibility between male and female gametes. Such barriers are more pronounced when genetically distant species are crossed, resulting in poor or unsuccessful hybridization. In vegetable breeding, these

limitations reduce the chances of obtaining viable hybrid embryos and hinder the transfer of desirable genes from wild relatives. For example, crosses between cultivated tomato (*Solanum lycopersicum*) and its wild relative *Solanum peruvianum* often exhibit pollen–pistil incompatibility and asynchronous flowering, leading to poor fertilization and low hybrid seed set. Plant breeders overcome these barriers through synchronized flowering, controlled pollination, reciprocal crosses, and the application of growth regulators to improve fertilization success (Sharma, 2017; Acquaah, 2020).

2. Post-fertilization Barriers

Post-fertilization barriers occur after successful fertilization and interfere with the normal development of hybrid embryos, resulting in reduced hybrid viability or fertility. Common post-fertilization barriers include embryo abortion, endosperm degeneration, chromosomal imbalance, hybrid weakness, hybrid breakdown, and complete or partial sterility. These barriers arise due to poor genomic compatibility between the parental species, preventing normal growth and development of the hybrid. In vegetable crops, embryo abortion is frequently observed in wide crosses between cultivated brinjal (*Solanum melongena*) and its wild relative *Solanum torvum*, where hybrid embryos fail to develop because of endosperm failure and genomic incompatibility. Such barriers are commonly overcome through embryo rescue, ovule culture, chromosome doubling, and bridge crossing, which facilitate the recovery of viable and fertile hybrids for crop improvement programmes (Acquaah, 2020; Singh, 2021; Sharma, 2017).

3. Genetic and Cytological Barriers

Genetic and cytological barriers arise from differences in chromosome number, genome organization, ploidy level, and gene interactions between the parental species involved in wide hybridization. These incompatibilities often disrupt homologous chromosome pairing during meiosis, resulting in irregular chromosome segregation, reduced recombination, low pollen fertility, and hybrid sterility. Consequently, the stable transfer of desirable genes from wild relatives to cultivated species becomes difficult. A well-known example is the intergeneric hybrid *Raphanobrassica*, developed by crossing radish (*Raphanus sativus*) and cabbage (*Brassica oleracea*). The initial hybrid was sterile because the chromosomes of the two parental species failed to pair during meiosis. Fertility was successfully restored by chromosome doubling using colchicine, leading to the development of a stable amphidiploid. Such approaches have become fundamental in overcoming cytogenetic barriers in wide hybridization and facilitating gene introgression in vegetable crop improvement (Singh, 2021; Acquaah, 2020; Sharma, 2017).

4. Techniques to Overcome Hybridization Barriers

The successful utilization of wide hybridization largely depends on overcoming the reproductive and genetic barriers that restrict gene transfer between distantly related species. Advances in plant breeding and biotechnology have led to the development of several techniques that enhance hybridization efficiency and facilitate the recovery of viable and fertile hybrids. These methods include the selection of compatible parents, reciprocal and bridge crosses, application of plant growth regulators, embryo rescue, chromosome doubling, protoplast fusion, and molecular breeding approaches. Each technique addresses specific pre- or post-fertilization barriers and improves the introgression of desirable genes from crop wild relatives into cultivated species. The combined use of these conventional and modern approaches has significantly expanded the scope of wide hybridization, making it an indispensable tool for developing disease-resistant, climate-resilient, and high-yielding vegetable cultivars (Acquaah, 2020; Sharma, 2017; Singh, 2021).

4.1 Selection of Compatible Parents

Selection of genetically compatible parents is the first step toward successful wide hybridization. Closely related species with desirable traits are preferred to increase crossability and hybrid viability. For example, crossing cultivated tomato (*Solanum lycopersicum*) with *S. pimpinellifolium* has been successfully used to transfer disease resistance and improve fruit quality due to their close genetic relationship (Acquaah, 2020; Singh, 2021).

4.2 Reciprocal crosses

Reciprocal crosses involve exchanging the male and female parents when an initial cross fails due to unilateral incompatibility. This approach helps identify the cross direction that produces viable seeds and fertile hybrids. For example, the cross *Solanum melongena* × *S. khasianum* is compatible, whereas the reciprocal cross (*S. khasianum* × *S. melongena*) is often unsuccessful because of pre-fertilization incompatibility. Therefore, reciprocal crossing is an effective strategy to improve the success of wide hybridization (Sharma, 2017; Singh, 2021).

4.3 Bridge Crosses

Bridge crossing is employed when two species cannot be crossed directly because of strong reproductive incompatibility. In this technique, an intermediate species that is compatible with both parents is used to facilitate gene transfer. For example, the direct cross between *Solanum bulbocastanum* and *S. tuberosum* is largely incompatible. However, *S. phureja* serves as a bridge species, allowing the transfer of valuable genes such as late blight resistance from *S. bulbocastanum* into cultivated potato. Bridge crossing has become an effective strategy for

overcoming incompatibility barriers and broadening the genetic base of crop improvement programmes (Sharma, 2017; Acquaah, 2020).

4.4 Use of Plant Growth Regulators

Plant growth regulators such as indole-3-acetic acid (IAA), naphthaleneacetic acid (NAA), 2,4-D, and gibberellic acid (GA₃) enhance the success of wide hybridization by promoting pollen germination, pollen tube growth, fertilization, and fruit set. For example, GA₃ application has improved hybrid seed production in interspecific tomato (*Solanum* spp.) crosses by increasing successful fertilization (Sharma, 2017; Acquaah, 2020).

4.5 Embryo rescue

Embryo rescue is an *in vitro* technique used to recover hybrid embryos that would otherwise abort due to post-fertilization incompatibility. Excised immature embryos are cultured on a suitable nutrient medium to produce viable plants. For example, embryo rescue has been successfully employed in the development of *Cucumis hystivus* through hybridization between *Cucumis hystrix* and *Cucumis sativus*, enabling the transfer of disease resistance and other desirable traits (Sharma, 2017; T. Cardi, 1997).

4.6 Chromosome Doubling

Chromosome doubling restores fertility in sterile interspecific or intergeneric hybrids by inducing homologous chromosome pairing during meiosis. Chemicals such as colchicine are commonly used to produce fertile amphidiploids. For example, chromosome doubling successfully restored fertility in the intergeneric hybrid *Raphanobrassica* (*Raphanus sativus* × *Brassica oleracea*) (Singh, 2021; Sharma, 2017).

4.7 Protoplast Fusion (Somatic Hybridization)

Protoplast fusion involves the fusion of isolated protoplasts from two genetically distinct species to produce somatic hybrids, thereby bypassing sexual incompatibility barriers. This technique facilitates the transfer of desirable traits between incompatible species. For example, somatic hybridization has been successfully used to transfer cytoplasmic male sterility (CMS) and disease resistance genes in *Brassica* species through protoplast fusion (Cardi, 1997; Acquaah, 2020).

4.8 Molecular Approaches in Wide Hybridization

Molecular approaches, including marker-assisted selection (MAS), genomic selection, and genome editing, facilitate the identification and introgression of desirable genes from wild relatives into cultivated vegetables. For example, molecular markers have been widely used to track disease resistance genes during tomato and Brassica breeding, thereby increasing the efficiency and precision of wide hybridization (Prohens *et al.*, 2017; Acquaah, 2020).

Applications of Wide Hybridization in Vegetable Crop Improvement

Wide hybridization has become an indispensable breeding strategy for introducing desirable genes from wild relatives into cultivated vegetables. It has been extensively utilized to improve disease and insect resistance, abiotic stress tolerance, yield, nutritional quality, and other economically important traits, thereby enhancing crop productivity and sustainability (Singh, 2021; Acquaah, 2020).

- **Disease Resistance:** Wide hybridization has been extensively used to transfer disease resistance genes from wild relatives into cultivated vegetable crops. For example, resistance to tomato leaf curl virus and Fusarium wilt has been successfully introgressed from wild *Solanum* species into cultivated tomato, resulting in improved disease resistance and enhanced crop productivity (Prohens *et al.*, 2017; Singh, 2021).
- **Insect Resistance:** Wide hybridization facilitates the transfer of insect resistance genes from wild species into cultivated vegetables. For instance, resistance to whiteflies and other insect pests has been successfully introduced into cultivated tomato from *Solanum habrochaites* and *S. pennellii*, thereby reducing yield losses and dependence on chemical insecticides (Acquaah, 2020; Prohens *et al.*, 2017).
- **Abiotic Stress Tolerance:** Wide hybridization enables the incorporation of genes for tolerance to drought, salinity, heat, and cold from wild relatives into cultivated vegetables. For example, heat tolerance has been transferred from *Solanum cheesmaniae* to cultivated tomato, while cold tolerance in onion has been improved using *Allium porrum*, enhancing crop adaptation to adverse environmental conditions (Singh, 2021; Acquaah, 2020).
- **Quality Improvement:** Wide hybridization contributes to the improvement of nutritional and horticultural quality by introducing beneficial genes from wild relatives. For example, the introgression of genes from *Solanum pimpinellifolium* into cultivated tomato has enhanced fruit quality, soluble solids, vitamin C, and antioxidant content, thereby increasing its nutritional and market value (Prohens *et al.*, 2017; Acquaah, 2020).
- **Yield Improvement:** Wide hybridization enhances yield potential by introducing beneficial genes for improved growth, adaptability, and productivity from wild relatives. For example, the potato cultivar 'Kufri Kuber', developed through interspecific hybridization involving *Solanum curtilobum*, exhibited improved tuber yield and adaptability. Such introgressions contribute significantly to the development of high-yielding vegetable cultivars (Singh, 2021; Acquaah, 2020).

- **Development of New Vegetable Types:** Wide hybridization facilitates the development of novel vegetable types by combining desirable traits from genetically distinct species or genera. A classic example is *Raphanobrassica*, developed by crossing radish (*Raphanus sativus*) and cabbage (*Brassica oleracea*). Similarly, *Hakuran*, developed through hybridization between cabbage and Chinese cabbage, exhibits superior adaptability, improved quality, and resistance to bacterial soft rot (Sharma, 2017; Singh, 2021).

Limitations of Wide Hybridization

Despite its significant contribution to vegetable crop improvement, wide hybridization faces several biological and technical limitations that restrict its widespread application.

- **Cross Incompatibility:** Genetically distant species often fail to hybridize because of pollen–stigma incompatibility, asynchronous flowering, or poor pollen tube growth, resulting in unsuccessful fertilization.
- **Hybrid Sterility:** Hybrids produced through distant crosses frequently exhibit complete or partial sterility due to irregular chromosome pairing and abnormal meiosis, limiting their breeding value.
- **Embryo Abortion and Hybrid Inviability:** Hybrid embryos may abort during early development because of genomic incompatibility or endosperm failure, preventing the recovery of viable hybrid plants.
- **Chromosomal Imbalance:** Differences in chromosome number, structure, or ploidy level often led to abnormal chromosome segregation, reduced fertility, and poor hybrid performance.
- **Linkage Drag:** Along with beneficial genes, undesirable traits from wild relatives may also be transferred, requiring repeated backcrossing to eliminate unwanted characteristics.
- **Transfer of Quantitative Traits:** Complex traits controlled by multiple genes are difficult to transfer because of low recombination frequency and unfavorable genetic interactions.
- **Long Breeding Cycle:** Wide hybridization requires several generations of backcrossing and selection, making the breeding process lengthy and labour-intensive.
- **Technical Complexity and Cost:** Techniques such as embryo rescue, chromosome doubling, and protoplast fusion require specialized laboratory facilities, skilled personnel, and considerable financial investment.

- **Limited Cross ability:** Successful hybridization is possible only between compatible species, restricting the utilization of many valuable wild relatives in breeding programmes.
- **Poor Agronomic Performance:** Initial wide hybrids often possess undesirable characteristics such as poor vigor, low yield, or inferior quality, necessitating extensive breeding before commercial utilization.

Future Prospects of Wide Hybridization

Wide hybridization is expected to play an increasingly important role in vegetable crop improvement by broadening the genetic base and utilizing the untapped potential of crop wild relatives. The integration of advanced technologies such as molecular marker-assisted selection, genomic selection, high-throughput sequencing, pangenomics, speed breeding, and CRISPR/Cas genome editing will significantly improve the efficiency of gene introgression and minimize linkage drag. Emerging techniques, including genomic-assisted breeding and artificial intelligence-based breeding tools, are expected to accelerate the identification and transfer of desirable genes governing resistance to diseases, insect pests, and abiotic stresses. Furthermore, climate change has increased the demand for resilient vegetable cultivars capable of withstanding drought, salinity, heat, and emerging pathogens. The conservation and systematic utilization of crop wild relatives, combined with modern biotechnology, will provide novel genetic resources for sustainable vegetable breeding. Consequently, wide hybridization will remain a cornerstone of future breeding programmes aimed at developing high-yielding, nutritionally enriched, climate-resilient, and resource-efficient vegetable cultivars that contribute to global food and nutritional security (Acquaah, 2020; Prohens *et al.*, 2017; Singh, 2021).

Conclusion

Wide hybridization has emerged as a powerful breeding strategy for broadening the genetic base of vegetable crops through the effective utilization of crop wild relatives. It facilitates the transfer of valuable genes for resistance to diseases and insect pests, tolerance to abiotic stresses, improved nutritional quality, and enhanced yield potential, thereby overcoming the limitations of conventional breeding. Although reproductive barriers, hybrid sterility, and genetic incompatibility remain major challenges, advances in biotechnology, including embryo rescue, chromosome doubling, protoplast fusion, molecular marker-assisted selection, and genome editing, have significantly improved the efficiency and success of distant hybridization. The successful development of improved cultivars in crops such as tomato, potato, cucumber, onion, Brassica vegetables, and okra demonstrates the immense potential of this approach. As climate

change and food security become global concerns, the integration of wide hybridization with modern genomic and precision breeding technologies will play a crucial role in developing climate-resilient, high-yielding, and nutritionally superior vegetable cultivars. Therefore, wide hybridization will continue to be an indispensable component of sustainable vegetable breeding programmes and a key driver of future genetic improvement.

References

1. Cardi, T. (1997). Somatic hybridization and cybridization in crop improvement. In J. Janick (Ed.), *Plant Breeding Reviews* (Vol. 15, pp. 67–96).
2. Chen, J. F., Staub, J. E., Adelberg, J. W., and Jiang, J. (1997). Successful wide hybridization between *Cucumis hystrix* Chakr. and *Cucumis sativus* L. *Euphytica*, 96, 413–419.
3. Dempewolf, H., Baute, G. J., Anderson, J., Kilian, B., Smith, C., and Guarino, L. (2017). Past and future use of crop wild relatives in breeding. *Crop Science*, 57(3), 1070–1082.
4. Food and Agriculture Organization. (2023). *The state of food and agriculture 2023*. FAO. Rome, Italy.
5. Hajjar, R., and Hodgkin, T. (2007). The use of wild relatives in crop improvement: A survey of developments over the last 20 years. *Euphytica*, 156, 1–13.
6. Prohens, J., Gramazio, P., Plazas, M., Dempewolf, H., Kilian, B., Díez, M. J., Fita, A., Herraiz, F. J., Rodríguez-Burruezo, A., Soler, S., Knapp, S., and Vilanova, S. (2017). Introgressiomics: A new approach for using crop wild relatives in breeding for adaptation to climate change. *Euphytica*, 213, 158.
7. Razali, R., Bougouffa, S., Morton, M. J. L., Lightfoot, D. J., Alam, I., Essack, M. (2018). The genome sequence of *Solanum pimpinellifolium* provides insights into salinity tolerance and genome evolution. *Scientific Reports*, 8, 4242.
8. Sharma, J. R. (2017). *Principles and practices of plant breeding* (3rd ed.). Tata McGraw-Hill Education.
9. Singh, B. D. (2021). *Plant breeding: Principles and methods* (11th ed.). Kalyani Publishers.
10. Suma, A., Joseph John, K., Bhat, K.V., Latha, M., Lakshmi, C.J., Pitchaimuthu, M., Nissar, V.A.M., Thirumalaisamy, P.P., Pandey, C.D., Pandey, S. and Kumar, A., 2023. Genetic enhancement of okra [*Abelmoschus esculentus* (L.) Moench] germplasm through wide hybridization. *Frontiers in Plant Science*, 14, 1284070.
11. Warschefsky, E., Penmetza, R. V., Cook, D. R., and von Wettberg, E. J. B. (2014). Back to the wilds: Tapping evolutionary adaptations for resilient crops through systematic hybridization with crop wild relatives. *American Journal of Botany*, 101(10), 1791–1800.

3D FOOD PRINTING OF FRUIT-BASED PRODUCTS: FORMULATION, PROCESSING AND FUTURE APPLICATIONS

Priya Kumari* and Neeraj Gupta

Division of Post Harvest Management,
Sher-e-Kashmir University of Agricultural Sciences and Technology,
Chatha, Jammu-180009, J&K, India

*Corresponding author E-mail: chaudharypriya162@gmail.com

Abstract

Three-dimensional (3D) food printing is an additive-manufacturing technology that builds edible structures layer by layer from computer-aided design files, and it has emerged as a powerful platform for transforming fruits and fruit processing by-products into customised, nutrient-dense foods. Fruits are, in their native state, classed as non-printable materials because their high moisture content, low structural protein and fat content, and variable fibre architecture make them prone to collapse, poor shape fidelity and phase separation during extrusion. This chapter reviews the scope of 3D food printing technology, the potential of fruit pulp, peel, pomace and seed fractions as printing feedstocks, and the principal extrusion, inkjet, binder-jetting and selective-sintering techniques used to fabricate fruit-based constructs. It further examines the formulation and optimisation of printable fruit inks, the role of hydrocolloids and functional additives in governing rheology and printability, and the processing parameters, including nozzle geometry, deposition speed, layer height and printing temperature, that dictate dimensional accuracy. Approaches to evaluating the physical, nutritional and sensory quality of printed fruit products are discussed alongside applications in functional foods, personalised and precision nutrition, dysphagia-friendly diets, and the valorisation of fruit processing waste within a circular bioeconomy. The chapter concludes with a discussion of the technical, regulatory, and commercial challenges that must be overcome before fruit-based 3D printing can move from laboratory prototypes to mainstream food manufacturing.

Keywords: 3D Food Printing, Fruit-Based Inks, Hydrocolloids, Extrusion, Personalised Nutrition, Food Waste Valorisation, Printability, Additive Manufacturing.

1. Introduction and Scope of 3D Food Printing Technology in the Food Industry

Additive manufacturing, popularly known as 3D printing, constructs three-dimensional objects by depositing material sequentially in two-dimensional layers that fuse together according to a digital computer-aided design (CAD) model (Tyupova & Harasym, 2024). Although the

technology originated in rapid prototyping for engineering, automotive, aerospace and biomedical applications, food scientists have progressively adapted the same layer-by-layer logic to edible materials over the past decade, giving rise to the discipline of 3D food printing (3DFP). Unlike conventional food-forming operations such as moulding, extrusion-cutting or casting, 3DFP allows a single platform to fabricate an almost unlimited variety of geometries, internal architectures and multi-material combinations directly from a digital file, without the need for product-specific dies or moulds (Godoi, Prakash, & Bhandari, 2016).

The scope of 3DFP within the food industry now spans confectionery and bakery decoration, dairy and protein-based structuring, meat analogue fabrication, and, increasingly, the design of fruit- and vegetable-based products intended for personalised nutrition, geriatric and paediatric diets, and space and extreme-environment feeding systems. Bibliometric analyses show that research output on food 3D printing has grown rapidly since 2018, with several hundred peer-reviewed articles published between 2018 and 2024 and an accelerating rate of publication in the fruit and vegetable sub-domain (Tyupova & Harasym, 2024). This growth reflects converging pressures on the food system: consumer demand for customised, visually attractive and nutritionally tailored foods; the need to reduce fruit and vegetable losses that, according to United Nations estimates, account for roughly one-eighth of food lost between harvest and sale (Tyupova & Harasym, 2024); and the broader digitalisation of food manufacturing under Industry 4.0. Within this landscape, fruit-based 3D printing occupies a distinctive niche because it combines the sensory and nutritional richness of fresh produce with the geometric freedom and personalisation potential of digital fabrication.

2. Potential of Fruits and Fruit By-Products as Printing Materials

Fruits and vegetables together supply a substantial share of dietary fibre, vitamins, minerals and phytochemicals, and their incorporation into printed foods is considered an important route toward more nutritious convenience products (Xu *et al.*, 2025). From a materials-science perspective, however, fresh fruit pulp is traditionally classified as a non-printable material: its high-water content (often above 80%), relatively low intrinsic viscosity, weak gel network and susceptibility to enzymatic browning and microbial spoilage mean that raw purées generally lack the yield stress and viscoelastic modulus required to support their own weight once deposited in multiple layers (Waghmare, Suryawanshi, & Karadbhajne, 2022). Pre-processing steps such as thermal blanching, partial dehydration, freeze-concentration or the addition of structuring agents are therefore required to convert fruit pulp into a stable, extrudable ink.

A second and increasingly important feedstock class is fruit processing by-products, including peels, pomace, seeds and press cakes generated by juicing, canning and jam manufacture. These

by-streams are typically rich in fibre, residual sugars and polyphenolic antioxidants, yet are commonly discarded or downgraded to low-value animal feed. Because 3D printing can accommodate irregular particle-size distributions and heterogeneous fibre content more readily than conventional forming operations, it has been proposed as a route for converting fruit by-products directly into structured, edible objects, thereby closing the loop between processing waste and consumer-ready foods (Tyupova & Harasym, 2024). Reported feedstocks include apple pomace, banana and blueberry purée blends, mango and pitaya pulp, citrus peel powders and berry press cakes, each requiring a distinct formulation strategy depending on its native fibre, pectin and sugar content.

Table 1: Representative fruit-derived printing materials and the principal formulation strategy used to render each one printable

Fruit Material	Limitation	Formulation Strategy	References
Banana/Blueberry purée blend	Low viscosity; syneresis on storage	Xanthan gum + carrageenan (1-4% w/w) blends to raise viscoelastic modulus	Domžalska & Jakubczyk, 2025
Pitaya (Dragon fruit) pulp	Weak water-holding capacity	Xanthan gum, Arabic gum or carboxymethyl-cellulose at 6-9 g/100 g	Recent Food Hydrocolloids studies (2025)
Apple/Citrus pomace	High fibre, low cohesiveness, particle blockage risk	Particle-size reduction, partial dehydration, hydrocolloid or starch binder addition	Tyupova & Harasym, 2024
Vegetable/Fruit powder composites	Excessive particle swelling raises viscosity unpredictably	Xanthan gum addition to inhibit particle swelling and stabilise rheology	Kim <i>et al.</i> , 2018

3. Types and Principles of 3D Food Printing Techniques

Four principal technologies are used to fabricate 3D printed foods: material extrusion, inkjet (drop-on-demand) printing, binder jetting, and selective sintering (Ehsan *et al.*, 2025). Extrusion-based printing, in which a semi-solid ink is forced through a nozzle and deposited along a programmed toolpath, is by far the most widely used approach for fruit-based systems because it can process viscous, particle-laden pastes that would clog the fine nozzles required for inkjet printing. Extrusion printing can be further subdivided into room-temperature extrusion of shear-thinning gels, fused-deposition-style hot-melt extrusion for fat- or sugar-based systems, and gel-forming extrusion in which a chemical or thermal setting reaction fixes the shape immediately after deposition (Waghmare, Suryawanshi, & Karadbhajne, 2022).

Table 2: Comparison of four techniques against criteria relevant to fruit-based printing

Technique	Working Principle	Suitability for Fruit Inks	Key Limitation
Extrusion (material jetting of pastes)	Nozzle-forced deposition of shear-thinning purée/gel	High - dominant method for fruit purées and pomace pastes	Requires precise rheological tuning to avoid slumping or clogging
Inkjet (drop-on-demand)	Droplet ejection of low-viscosity liquid ink	Low-to-moderate - limited to surface decoration and colouring	Cannot build tall self-supporting 3D structures
Binder jetting	Liquid binder sprayed onto a powder bed layer by layer	Moderate - suited to dried fruit or sugar-fruit powder blends	Fragile, porous structures; limited to dry formulations
Selective (laser/hot-air) sintering	Localised heat fuses powder particles without a binder	Low-to-moderate - emerging for fruit-sugar powder systems	Thermal degradation of vitamins, pigments and aromas

Inkjet printing deposits fine droplets of low-viscosity liquid ink and is best suited to two-dimensional graphic decoration, flavour or colour deposition, and fine detailing rather than the fabrication of tall, self-supporting three-dimensional fruit structures. Binder jetting builds objects from a bed of powdered material, such as dried fruit or sugar powder, by selectively spraying a liquid binder layer by layer; it produces intricate, support-free geometries but is presently limited to relatively dry, powder-compatible fruit formulations. Selective laser sintering (SLS) and its hot-air-sintering variant fuse powder particles using a directed heat or laser source and can achieve fine resolution without external support structures, although thermal exposure raises concerns about degradation of heat-sensitive vitamins and pigments in fruit powders (Ehsan *et al.*, 2025).

4. Development and Optimisation of Fruit-Based Printing Formulations

Formulating a printable fruit ink is fundamentally an exercise in rheological engineering: the ink must flow readily under the shear applied during extrusion, yet recover sufficient elastic strength immediately after deposition to support the weight of subsequent layers without slumping or collapsing (Domžalska & Jakubczyk, 2025). Printability is commonly assessed through a combination of flow curves, oscillatory frequency and strain sweeps, three-interval thixotropy tests, and direct extrusion trials that measure extrusion force, filament uniformity, shape fidelity and post-printing stability over time. A formulation is generally considered printable when its

storage modulus (G') exceeds its loss modulus (G'') across the relevant frequency range, indicating a predominantly elastic, gel-like network capable of self-support (Domžalska & Jakubczyk, 2025).

Optimisation of fruit-based inks typically proceeds through a stepwise process: (i) pre-treatment of the raw fruit material (peeling, blanching, homogenisation, partial dehydration or particle-size reduction) to standardise the base matrix; (ii) incremental incorporation of structuring agents, most commonly hydrocolloids, starches or proteins, at concentrations selected via factorial or response-surface experimental designs; (iii) rheological and textural characterisation of candidate formulations; and (iv) iterative print trials in which nozzle diameter, printing speed, layer height and extrusion pressure are adjusted to match the rheological profile of the ink (Waghmare, Suryawanshi, & Karadbhaje, 2022). Blending complementary fruit purées, for example combining a low-viscosity fruit such as banana with a higher-pectin fruit such as blueberry, has also been used to balance colour, flavour and structural performance without relying solely on added hydrocolloids (Domžalska & Jakubczyk, 2025).

5. Role of Hydrocolloids and Additives in Improving Printability and Stability

Hydrocolloids are the single most important class of functional additive used to convert non-printable fruit purées into stable printing inks. Polysaccharide gums such as xanthan gum, carrageenan, guar gum, locust bean gum, alginate, pectin and carboxymethylcellulose confer thickening, gelling, stabilising and emulsifying functionality, and their concentration and combination directly determine the viscoelastic modulus, yield stress and extrudability of the resulting ink (Domžalska & Jakubczyk, 2025). In a study on blueberry and banana purée-based inks, formulations combining 4% xanthan gum with 4% carrageenan produced the highest viscoelasticity and extrudability indices among the formulations tested, while comparable performance could be achieved at lower total hydrocolloid loading (3% xanthan gum with 2-3% carrageenan), illustrating that hydrocolloid type and synergistic combination can be as important as absolute concentration (Domžalska & Jakubczyk, 2025).

Hydrocolloids act through several complementary mechanisms: they restrict free water mobility within the matrix, which improves water-holding capacity and reduces syneresis during storage; they raise the apparent viscosity and shear-thinning behaviour needed for smooth extrusion; and they promote the formation of hydrogen-bonded or ionically cross-linked networks that stiffen the printed filament immediately after deposition. Xanthan gum has been repeatedly shown to inhibit excessive swelling of suspended fruit or vegetable particles, thereby limiting unpredictable increases in viscosity at high solids loading and improving batch-to-batch consistency (Kim *et al.*, 2018). Pectin, already naturally present in many fruits, offers an

additional advantage: low-methoxyl pectin can form calcium-bridged gels while high-methoxyl pectin gels through sugar-acid interactions, giving formulators flexibility to tune gel-setting behaviour to the natural sugar and acid profile of a given fruit matrix. Beyond hydrocolloids, other functional additives used in fruit-ink formulation include native or modified starches for bulk viscosity, plant or dairy proteins for mechanical reinforcement and nutritional enrichment, ascorbic acid and other antioxidants to limit enzymatic browning, and calcium salts as cross-linking agents for pectin- and alginate-based systems.

6. Effect of Processing Parameters on Printing Quality

Even a well-formulated fruit ink can yield a poor-quality printed object if the processing parameters of the printer are not matched to its rheological profile. Nozzle diameter governs the resolution of the printed filament and the shear rate experienced by the ink during extrusion; narrower nozzles improve detail but increase back-pressure and the risk of blockage from fibrous particles, while wider nozzles reduce resolution but improve throughput and reduce clogging in fibre-rich fruit pomace formulations (Ehsan *et al.*, 2025). Printing (deposition) speed and extrusion or motor speed must be balanced so that the volumetric flow rate of ink matches the speed of nozzle travel; mismatches produce either under-extrusion, with gaps and weak inter-layer bonding, or over-extrusion, with filament bulging and loss of dimensional accuracy.

Layer height interacts with the ink's yield stress to determine whether successive layers can support their own weight: excessively tall layers relative to the ink's structural strength lead to progressive slumping and eventual structural collapse in taller prints, whereas layers that are too thin unnecessarily prolong printing time. Printing temperature affects viscosity directly, and for fat-containing or hot-melt fruit-chocolate composite inks, precise control of nozzle and bed temperature is critical for consistent flow and post-deposition solidification. Post-processing steps, including oven or convective drying, freeze-drying, baking or frying, are frequently required to achieve the target moisture content, microbial safety and mechanical stability of the final product, but each of these steps can itself alter the printed geometry through shrinkage or structural relaxation, so post-processing must be considered an integral part of the overall process design rather than an afterthought (Xu *et al.*, 2025).

7. Quality Evaluation of 3D Printed Fruit Products (Physical, Nutritional, and Sensory Properties)

Quality evaluation of printed fruit products spans physical, nutritional and sensory dimensions. Physical assessment typically includes dimensional accuracy (comparison of printed height, width and geometric fidelity against the CAD model), texture profile analysis (hardness, springiness, cohesiveness, chewiness and adhesiveness), colour measurement (CIE L*, a*, b*

values, which are particularly sensitive to hydrocolloid type and concentration in fruit inks), and shape-retention or collapse testing over defined storage intervals (Domžalska & Jakubczyk, 2025).

Nutritional evaluation focuses on the retention of thermolabile and oxidation-sensitive bioactive compounds, including vitamin C, anthocyanins, carotenoids and total phenolic content, through the printing and any subsequent drying or cooking steps. Controlled-temperature drying of printed fruit-derived formulations has been reported to retain a large proportion of vitamin C content, and fruit-peel-derived printed products have shown appreciably higher polyphenol retention than equivalent products made by traditional extrusion cooking, indicating that the comparatively mild thermal history of extrusion-based 3D printing can be nutritionally advantageous relative to conventional high-shear, high-temperature processing routes. Sensory evaluation, using trained or consumer panels, assesses appearance, aroma, taste, texture and overall acceptability, and is essential because structural novelty alone does not guarantee consumer acceptance; several studies have found that hydrocolloid type and concentration, while beneficial for printability, can also alter mouthfeel and flavour release in ways that must be balanced against structural requirements (Tomašević *et al.*, 2021).

8. Applications in Functional Foods, Personalised Nutrition, and Value-Added Products

The capacity of 3D printing to deposit different inks in precisely controlled spatial patterns makes it a natural platform for functional and personalised foods. Fruit-based inks can be fortified with proteins, prebiotic fibres, probiotics, vitamins or minerals to create nutrient-dense snacks tailored to the requirements of specific population groups, and multi-material printing allows different nutrient loadings to be deposited within a single object, for example to create graded-release or portion-controlled structures (Tomašević *et al.*, 2021).

A particularly well-developed application is the design of texture-modified, fruit-based foods for individuals with dysphagia, a swallowing disorder that is increasingly prevalent among the elderly and certain clinical populations. Conventional pureed or thickened diets for dysphagia patients are often visually unappealing and nutritionally diluted, which can contribute to reduced food intake and malnutrition. 3D printing allows fruit purées to be reformulated into soft, easily swallowed gels that nonetheless retain a recognisable, appetising shape, and hydrocolloid-modified fruit gels have been used to engineer textures that meet recognised dysphagia-diet texture categories while improving nutrient density and sensory appeal (Chao, Nam, Park, & Kim, 2024). Beyond clinical nutrition, fruit-based 3D printing has been explored for children's foods designed to encourage fruit consumption through novel shapes, for athlete and geriatric

nutrition requiring precise macronutrient dosing, and for premium confectionery and dessert applications that leverage the technology's capacity for intricate, customised geometries.

9. Utilisation of Fruit Waste and By-Products for Sustainable Food Production

Global estimates suggest that well over a billion tonnes of food-processing waste are generated annually, with fruit and vegetable by-streams, including peels, pomace, seeds and off-specification produce, representing a substantial share of this total. Because these materials are frequently rich in dietary fibre, residual sugars and antioxidant polyphenols, their direct incorporation into 3D-printed foods offers a route to simultaneously reduce disposal costs, capture nutritional value that would otherwise be lost, and support a circular bioeconomy within the fruit-processing sector (Tyupova & Harasym, 2024).

Successful valorisation of fruit by-products for printing generally requires attention to particle-size uniformity and hydration level, since both properties strongly influence the rheology and extrudability of the resulting ink; combining fruit residues with hydrocolloids or protein-based binders has been shown to widen the achievable design space for complex geometries and layered structures. High residual sugar content in some fruit by-products can promote caramelisation during thermal post-processing, which may compromise flavour and colour if processing temperatures are not carefully controlled. Reported outcomes from waste-derived printed formulations include measurable increases in fibre and polyphenol content relative to conventionally processed products, supporting the broader case for 3D printing as a value-adding technology for otherwise under-utilised fruit-processing residues rather than a purely novelty-driven application.

10. Challenges, Future Prospects, and Commercial Applications of Fruit-Based 3D Printing

Despite substantial research progress, several challenges continue to limit the transition of fruit-based 3D printing from laboratory demonstration to mainstream commercial production. Key technical challenges include the poor inherent printability of many fresh fruit matrices, the need for extensive, material-specific formulation optimisation, relatively slow production speeds compared with conventional forming operations, and the risk of nutrient degradation during pre-treatment, printing and post-processing (Xu *et al.*, 2025). Microbiological safety is a further concern, since multi-step handling of moist, nutrient-rich fruit pastes at ambient or mildly elevated temperatures can create opportunities for microbial growth if hygienic design and process controls are not rigorously applied. Regulatory frameworks for 3D-printed foods remain underdeveloped in most jurisdictions, and any formulation intended to replace an established food product must additionally satisfy novel-food safety and labelling requirements (Tyupova & Harasym, 2024).

Looking forward, several converging trends are likely to shape the future of the field. Continued refinement of hydrocolloid and protein-based formulation strategies, supported by predictive rheological modelling, should reduce the trial-and-error burden currently associated with new fruit-ink development. Integration of in-line sensing and closed-loop process control promises to improve the reproducibility of printed geometries across batches of variable fruit raw material. The extension of 3D printing toward four-dimensional (4D) food printing, in which printed structures undergo a programmed, stimulus-triggered change in shape, colour or texture after fabrication, offers further opportunities for fruit-based products with dynamic sensory attributes. Commercially, early adoption is concentrated in confectionery, dessert decoration and niche clinical or personalised-nutrition markets, with start-ups already demonstrating the conversion of fruit and bread waste into printed, shelf-stable snack products (Tyupova & Harasym, 2024). Realising the full potential of fruit-based 3D printing at industrial scale will depend on parallel progress in cost-effective materials, higher-throughput printing hardware, harmonised regulatory standards, and continued consumer research to build trust and acceptance of this still-emerging food technology.

Conclusions

Three-dimensional printing offers a versatile, digitally controlled route for transforming fruits and fruit-processing by-products into structured, nutritionally enhanced and highly customisable foods. Success in this domain depends on a tightly coupled understanding of fruit-material rheology, hydrocolloid and additive functionality, and printer processing parameters, since each of these factors interacts to determine the printability, structural stability and final quality of the printed product. Applications in personalised and clinical nutrition, particularly dysphagia-friendly diets, and in the valorisation of fruit-processing waste illustrate the technology's potential to deliver both nutritional and sustainability benefits. Continued advances in formulation science, process automation and regulatory clarity will be necessary before fruit-based 3D printing can move beyond specialised and clinical niches into broader commercial food manufacturing.

References

1. Chao, C., Nam, H. K., Park, H. J., & Kim, H. W. (2024). Potentials of 3D printing in nutritional and textural customization of personalized food for elderly with dysphagia. *Applied Biological Chemistry*, 67(1), 25. <https://doi.org/10.1186/s13765-023-00854-7>
2. Domžalska, Z., & Jakubczyk, E. (2025). The impact of ink composition and its physical properties on the selected attributes of 3D-printed fruit purées with hydrocolloid molecules. *Molecules*, 30(16), 3394. <https://doi.org/10.3390/molecules30163394>

3. Ehsan, S., et al. (2025). Three-dimensional (3D) food printing applications, techniques, and consumer acceptance. *eFood*, 6. <https://doi.org/10.1002/efd2.70106>
4. Godoi, F. C., Prakash, S., & Bhandari, B. R. (2016). 3D printing technologies applied for food design: Status and prospects. *Journal of Food Engineering*, 179, 44–54.
5. Kim, H. W., Lee, J. H., Park, S. M., Lee, M. H., Lee, I. W., Doh, H. S., & Park, H. J. (2018). Effect of hydrocolloids on rheological properties and printability of vegetable inks for 3D food printing. *Journal of Food Science*, 83(12), 2923–2932. <https://doi.org/10.1111/1750-3841.14391>
6. Tomašević, I., Putnik, P., Valjak, F., Pavlič, B., Šojić, B., Markovinović, A. B., & Kovačević, D. B. (2021). 3D printing as novel tool for fruit-based functional food production. *Current Opinion in Food Science*, 41, 138–145.
7. Tyupova, A., & Harasym, J. (2024). Valorization of fruit and vegetables industry by-streams for 3D printing: A review. *Foods*, 13(14), 2186. <https://doi.org/10.3390/foods13142186>
8. Vancauwenberghe, V., Verboven, P., Lammertyn, J., & Nicolaï, B. (2018). Development of a coaxial extrusion deposition for 3D printing of customizable pectin-based food simulant. *Journal of Food Engineering*, 225, 42–52.
9. Waghmare, R., Suryawanshi, D., & Karadbhajne, S. (2022). Designing 3D printable food based on fruit and vegetable products: Opportunities and challenges. *Journal of Food Science and Technology*, 60(5), 1447–1460. <https://doi.org/10.1007/s13197-022-05386-4>
10. Xu, B., Mao, B., Wang, X., Liu, J., Liu, Z., Guo, C., Wei, B., Lin, L., Guo, Z., Zhang, M., Zhou, C., & Ma, H. (2025). Overview of strategies for 3D printing of fruit and vegetables: Mechanism, modification, challenges and future trends. *Trends in Food Science & Technology*.

FORMULATION AND EVALUATION OF A SKIN BARRIER-REPAIR CREAM FOR SENSITIVE SKIN USING CERAMIDES AND NATURAL OILS

Surya Kumar Sahu*, Pushpendra Kumar Sahu and Digeshwari Patel

School of Pharmacy,
Chouksey Engineering College, Bilaspur (CG)

*Corresponding author E-mail: suryasahu6265@gmail.com

1. Introduction

Products for sensitive skin represent the important interface between cosmetics, dermatology and pharmaceuticals. Barrier-repair cream is distinctly different from ordinary emollients which primarily aim to soften the skin surface. The main purpose of a barrier repair cream is to ensure optimum functional quality of the skin's outermost layer (stratum corneum). Additionally, it also aims to minimize the irritant burden on the skin. Moreover, barrier repair creams must also remain physically stable throughout their storage period and use. More information about Sensitive Skin [1-3].

Sensitive skin is often defined by subjective symptoms such as burning, stinging, itching, tingling and tightness, and often weak or absent visible inflammation; thus evaluation of products is more complex than evaluation of simple dryness [4-6].

Skin barrier are not passive covering alone. It is a complex and ever-changing physiochemical interface in which corneocytes, intercellular lipids, natural moisturizing factors, pH, enzymes, microbiome contacts and immune signals interact with each other. Disruption of the barrier can cause excessive loss of water and lower mechanical resilience. It may also heighten sensory reactivity. Lipid abnormalities, altered ceramide profiles and elevated surface pH have been shown to negatively affect barrier function and increase susceptibility to irritants in atopic-prone and dry skin [7-9].

Ceramides are particularly relevant for barrier-repair products because they are major constituents of the intercellular lipid matrix and contribute to lamellar packing within the stratum corneum. Recent research has shown that the topically applied formulations containing physiological lipids can enhance the structure, water content and tolerability of skin barrier when delivered in suitable vehicles as opposed to just adding single marketing ingredients [10-12].

Natural oils may add emolliency, occlusion, sensorial comfort, and lipid supplementation, but must be limited. The use of oils high in linoleic acid, wax esters or skin-compatible triglycerides may help provide sensorial improvement and barrier care support. However, among sensitive

populations, perfume-rich essential oils and highly oxidizable oils may increase irritation risk. As such, a barrier-repair cream for sensitive skin should contain ceramides, complementary lipids, humectants, non-ionic emulsifiers, mild preservatives and pH-compatible aqueous components in a stable oil-in-water or lamellar emulsion [13-15].

The purpose of this review is to provide a systematic formulation and evaluation framework for a ceramide-natural oil skin barrier-repair cream. This review incorporates journal literature published between 2020 and 2025, uses Vancouver-style numbered citations, and presents adaptable tabular synthesis and formulation parameters in a format suitable for an Elsevier-style review manuscript [16-18].

2. Structure and Function of the Skin Barrier

2.1 Epidermis

The epidermis is the outermost layer of the skin and acts as the body's first protective barrier against the external environment. It is a thin, a vascular (without blood vessel), stratified squamous keratinized epithelium primary composed of keratinocytes. The epidermis protects the body from mechanical injury, harmful microorganism, chemicals ultraviolet (UV) radiation, and excessive water loss.

The epidermis is mainly composed of four types of cells: Keratinocytes, Melanocytes, Langerhans cells, and Merkel cell. Keratinocytes constitute about 90-95% of all epidermal cells and produce keratin, a structural protein that provides strength, waterproofing, and protection to the skin. Melanocytes are present in the basal layer and synthesize melanin, the pigment responsible for skin colour and protection against ultraviolet radiation. Langerhans cells are specialized antigen-presenting immune cells that help detect and eliminate invading microorganism. Merkel cells are sensory receptor cells associated with nerve endings and are involved in the perception of light touch and pressure.

2.2 Dermis

The dermis is the thick, living connective tissue layer of the skin located immediately beneath the epidermis and above the hypodermis (subcutaneous tissue). It provides structural integrity, mechanical strength, elasticity, and nourishment to the skin.

Histologically, the dermis is divided into two distinct regions: the papillary dermis and the reticular dermis, each with unique structural and functional characteristic. The papillary dermis is the superficial, thinner layer that lies directly beneath the epidermis. It consists of loose connective tissue rich in fine collagen (predominantly type III collagen), elastic fibre, fibroblast, mast cells, macrophages, and a dense network of capillaries. The Reticular dermis

forms the deeper and thicker portion of the dermis and is composed of dense irregular connective tissue containing thick bundles of type I collagen fibre interwoven with elastic fibres.

2.2 Hypodermis

The hypodermis, also known as the subcutaneous tissue, is the deepest layer of the skin located beneath the dermis. It is primarily composed of loose connective tissue and adipose (fat) cells, which are arranged in lobules separated by fibrous septa. The hypodermis functions mainly as an energy reservoir, thermal insulator, and shock absorber, protecting underlying muscles and organs from mechanical injury. It also facilitates the attachment of the skin to underlying structures while allowing flexibility and movement.

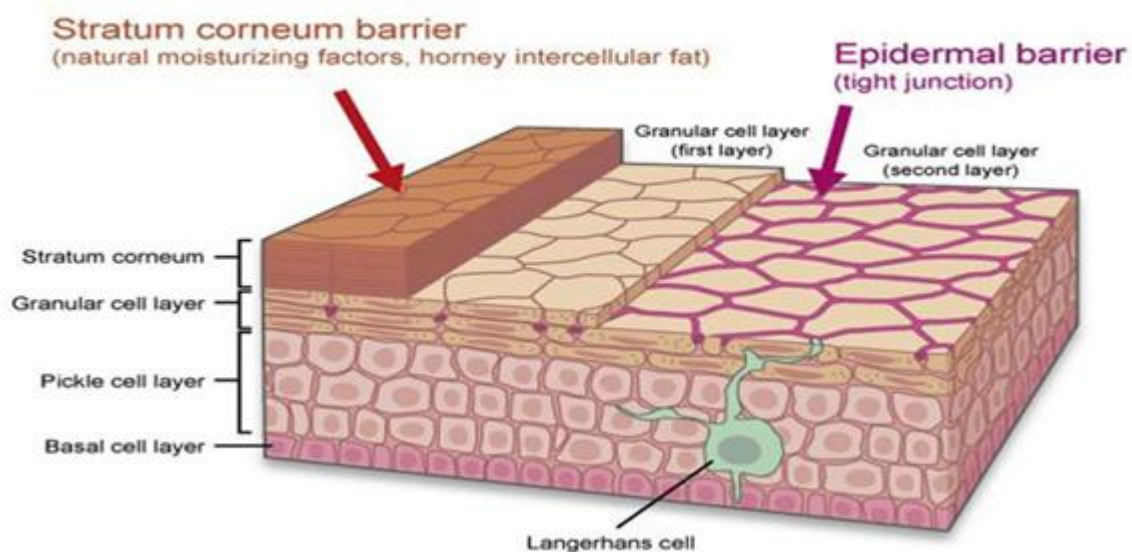


Figure 1: The "Brick-and-Mortar" model of the stratum corneum. Corneocytes (bricks) are surrounded by a continuous matrix of intercellular lipids (mortar) including ceramides, cholesterol, and free fatty acids. Disruption of this matrix leads to increased TEWL and heightened sensitivity

3. Pathophysiology of Sensitive Skin

The term "sensitive skin" refers to a syndrome and NOT a pathological disease. The patient complains of unpleasant sensations after contact with a variety of stimuli such as cosmetics, cleansers, temperature change, wind, ultraviolet radiation, low humidity, pollution or psychological stress. Although visible signs may be limited, patient-reported stinging or burning can have clinical relevance even with weak erythema [31-33].

The recent rationale incorporates several interrelated mechanisms, such as barrier dysfunction, modified neurosensory reactivity, inflammation, vascular reactivity and a microbiome impact. If the barrier is weakened, irritant penetration is enhanced. Moreover, sensory nerve endings could be activated by small changes in the chemical or thermal environment. This is important because

a product for sensitive skins must be tested for not only hydration but also tolerability and sting potential [34-36].

Sensitive skin often coexists with dryness, irritation from acne treatments and atopic tendency. Thus, a cream high in ceramides can be useful as an adjunct to restoring comfort in multiple settings, provided the formulation is non-comedogenic when applied to the face and does not contain plant allergens [40-42].

3.1 Epidermal Barrier Dysfunction

A growing body of evidence indicates that epidermal barrier impairment is a primary driver rather than a secondary consequence of sensitive skin. Individuals with sensitive skin frequently exhibit elevated transepidermal water loss (TEWL), reduced stratum corneum hydration, and increased permeability to irritants even in the absence of overt inflammation (Fluhr *et al.*, 2012). Barrier disruption facilitates the penetration of environmental aggressors such as surfactants, pollutants, and allergens, which subsequently activate keratinocytes and resident immune cells. This activation leads to the release of pro-inflammatory mediators, neuropeptides, and reactive oxygen species, thereby perpetuating a vicious cycle of barrier damage and sensory hyperreactivity (Elias & Feingold, 2001).

3.2 Neurosensory Hyperreactivity

In addition to barrier abnormalities, neurosensory dysfunction plays a crucial role in the manifestation of sensitive skin symptoms. Cutaneous sensory nerve endings in sensitive skin display heightened responsiveness to chemical, thermal, and mechanical stimuli (Misery *et al.*, 2017). This hyperreactivity is mediated in part by transient receptor potential (TRP) ion channels, particularly TRPV1 and TRPA1, which act as molecular sensors for heat, acidity, and irritants. Overactivation of these channels results in exaggerated sensations of burning, stinging, and itching, even in response to normally innocuous stimuli (Stander *et al.*, 2016). Barrier repair strategies that reduce irritant penetration may therefore indirectly attenuate neurosensory activation.

4. Ceramide in Skin Barrier Repair

Ceramides are a class of lipid molecules belonging to the sphingolipid family. Chemically, they consist of a long-chain sphingoid base (such as sphingosine or Phyto sphingosine) linked to a fatty acid via an amide bond. In human skin, ceramides are the predominant lipid component of the stratum corneum and are classified into several subclasses based on variations in the sphingoid base and fatty acid chain length. Commonly identified skin ceramides include Ceramide NP, NS, AP, AS, EOS, and EOP. These structural variations contribute to the formation of highly ordered lamellar bilayers essential for epidermal barrier function.

The stratum corneum barrier is essential for the functioning of ceramide. Ceramides are organics that function with cholesterol. When ceramide amount, length or subclass distribution changed, lamellar packing is less well ordered and water diffusion can increase. Disturbance in barrier-compromised skin manifest as dryness, scaling, itching, reactivity and poor tolerance to topical products [43-45].

The purpose of using ceramides is to replace depleted lipids in the skin barrier. Clinical studies indicate that creams and lotions containing ceramides in multivesicular emulsions can support barrier restoration in dry, eczema-prone skin. Further evidence shows that creams containing ceramides, triglycerides and cholesterol can improve lipid organization and provide protection against irritation in adults with dry, eczema-prone skin [46-48].

More recent efforts have strengthened the mechanistic argument. Recent clinical evidence indicates that adults who are at risk for atopic dermatitis can benefit from physiological lipid supplementation that rebalances their stratum corneum ceramide profile and strengthens their barrier function. The findings are relevant for cosmetic formulation because they mean that the ratios of lipids, the delivery format and repeated application matter more than just having ceramide on the label [49-51].

Sensitive skin ceramide systems should usually be designed as low-dose, well-dispersed, multi-lipid systems. Incorporation of ceramides must be done using suitable solubilizers, lamellar emulsifiers or pre-dispersed ceramide complexes to avoid crystallisation due to poor incorporation. To align it with the physiological lipid milieu, lipid partners such as cholesterol, fatty acids or triglycerides should also be present in the oil phase [52-54].

To substantiate claims, objective skin measurements should be used on ceramide creams. TEWL, corneometry, capacitance, visual dryness scoring, irritation scoring and user-reported comfort can indicate whether the formulation performs more as a barrier-support product rather than as a standard moisturizing cream [55-57].

5. Role of Natural Oils in Barrier Repair

Natural oils used in barrier-repair creams should be selected according to their fatty-acid profile, oxidative stability, allergen risk, sensory quality, and emulsion-system compatibility. In the case of sensitive-skin products, the term natural cannot be used interchangeably with safe. Essential oils and fragrant extracts may increase sensitization risk. Refined fixed oils are usually a more reasonable choice than aromatic oils [58-60].

The recent review literature identifies *Helianthus annuus* seed oil as a plant lipid that is dermatologically relevant but benefit at the product level will depend on refining quality, peroxide value, dose and vehicle. Jojoba oil has an unusual structure and contains no triglycerides. It is relatively

close to a liquid wax ester in its chemical and physical properties. It is a stable substance and has good light sensory properties. Jojoba oil may be suitable for facial creams [61-63].

In a ceramide cream, natural oils must not do away with physiological lipids. They help to improve spreadability, reduce friction, decrease perceived tightness, and reinforce the emollient phase. The logic behind barrier repair still evolves around the combination of humectants, pH-compatible emulsion architecture and ceramides, cholesterol/free fatty acids [70-72].

5.1 Jojoba oil (*Simmondsia chinensis*)

Because jojoba oil is mostly composed of wax esters rather than triglycerides, it differs from other plant oils in that it is chemically closer to human sebum, which is around a quarter to a third wax esters. Because of this similarity, jojoba oil may enter follicular apertures and the outer stratum corneum without acting like a normal comedogenic oil, making it a sensible option even for sensitive skin that is greasy or prone to act.

Jojoba oil has a helpful quantity of tocopherols and associated antioxidants that help prevent lipid peroxidation, and its wax esters offer occlusion without suffocating the skin. Patients, especially those with truly hypersensitive or blemish-prone skin, often tolerate it because of its non-comedogenic behavior and pleasant skin feel.

5.2 Sunflower seed oil (*Helianthus annuus*)

Because sunflower oil contains a significant amount of linoleic acid (about 50-70% of its total fatty acid content), it is especially important for barrier restoration. Research has shown that sunflower oil, which is rich in oleic acid, might potentially worsen it.

Linoleic acid's direct feeding into ceramide EOS production and its stimulation of peroxisome proliferator-activated receptors (PPARs) in the epidermis, which turn on genes involved in lipid synthesis and barrier development, are probably the main sources of sunflower oil's benefits. Additionally, it has a helpful supply of phytosterols and tocopherols, which reduce inflammation, shield unsaturated fatty acids from oxidation, and scavenge free radicals.

5.3 Argan oil (*Argania spinosa*)

The unsaponifiable fraction, or about 1% of the oil, which contains tocopherols, phytosterols, and polyphenols on top of a fatty acid profile dominated by oleic and linoleic acids, is what gives argan oil, which is extracted from the kernels of a Moroccan tree, its dermatological reputation with a tocopherol concentration of between 60 and 90 mg per 100 g of oil, its antioxidant potential is quite strong, and γ -tocopherols plays a significant role in protecting it against oxidative and peroxidative damage.

Clinical research has connected the use of argan oil to better skin hydration and suppleness, especially in postmenopausal women, a group where age-related barrier degradation is already a

concern. The fatty acids in argan oil also seem to function as ligands for PPAR- α and PPAR- γ , and activating these receptors appears to promote the synthesis of antimicrobials peptides as well as endogenous ceramide formation. Argan oil is a particularly helpful addition to barrier-healing formulations because it combines direct lipid administration with indirect activation of the skin's own repair machinery.

6. Formulation Design of Ceramide-Natural Oil Cream

There should be a mild cream which should be fragrance free and pH compatible cream which should be formulate as an emulsion with the rational ceramide-natural oil not be so complicated that there would be too much lipid for the barrier repair or so that it creates a greasy product, tarry product, unstable product, or comedogenic product. For sensitive skin, an oil-in-water cream is often practical as it has a good balance of spreadability, rinse-off resistance, cooling feel and user acceptability. A lamellar or multi-vesicular emulsion can also additional sustained lipid delivery (Danby et al, 2020; Danby et al, 2022; Mawazi et al, 2022).

The aqueous phase must have purified water, humectants and pH modifiers. Glycerine is a core humectant with wide compatibility and a known moisturising capacity. The use of an glycols such as butylene glycol or propanediol may enhance the humectancy and preservative support of a formulation. However, in very reactive users, high levels may be avoided. You may use soothing excipients such as panthenol, allantoin or beta-glucan, if the final formula is stable and safety data is available (Mawazi *et al.*, 2022; Kim *et al.*, 2025).

the lipid phase should consist of ceramide blend, cholesterol or cholesterol-compatible lipid, free fatty acid or triglyceride, and optional natural oil. The most likely reason for the low level of ceramides in cosmetic products is potency. Total ceramide complex is a practical starting range like any other properly dispersed ceramide 0.05-0.30% w/w. Natural oils may be kept at 3-10% w/w. That depends on target skin feel and phase ratio. These limits should be viewed as formulation targets, rather than strategic regulatory limits (Danby *et al.*, 2022; Andrew *et al.*, 2025).

The emulsifier system is vital. Due to the fact that they maintain stable texture with less irritancy than harsh surfactants, sensitive skin generally prefers non-ionic emulsifiers and cationic polymeric stabilizers. Fatty alcohols like cetyl alcohol or cetearyl alcohol give viscosity and emolliency, but they must be optimized out to avoid waxy drag. Mawazi *et al.* (2022); Wang *et al.* (2024) suggest the inclusion of dimethicone or other low-irritancy barrier-forming silicones if the product does not require silicone-free status.

Conservatism must be strong but mild. Water and oil are present in creams; thus microbial protection is obligatory. Preservative efficacy testing should drive selection of preservatives, not

consumer perception. The system of packaging should minimize access to air along with oxidation with the natural polyunsaturated oil. Sensitive-skin products for repeat use should be packed in airless pumps or laminated tubes rather than wide mouth jars (Mawazi *et al.*, 2022; Lim *et al.*, 2025).

7. Evaluation Parameters

An assessment of a barrier cream for sensitive skin should include quality tests and a dermal performance endpoint. The objective is to demonstrate not just that the product looks acceptable, but also that it is physically stable, microbiologically safe, comfortable, non-irritating and has an effect that improves hydration or reduces TEWL under defined conditions (Mawazi *et al.*, 2022; Wang *et al.*, 2024).

7.1 Physical Features

7.1.1 Product Appearance

The product should be visually smooth, uniform and free from phase separation, oil bleeding, discolouration, grittiness, entrapped air pockets and visible microbial contamination. The appearance must be examined closely straight after manufacturing and after storage in room temperature, accelerated, and freeze-thaw conditions. Scent should be neutral or bland, especially since fragrance-free positioning is preferred for sensitive skin.

The pH value shall be determined in a defined aqueous dispersion, specifically, 10% w/w cream dispersion in purified water, using a calibrated pH meter. For sensitive facial skin, a target pH around 4.8-5.8 is appropriate since it is generally compatible with the acid mantle and not strongly acid or alkaline. The change in pH during stability testing can be an indication of the degradation, microbial contamination of the product or weakness of the buffer (Andrew *et al.*, 2024; Sakai and Hatano, 2025).

7.1.2 Viscousness

Dispensing, spreadability and consumer acceptance depends on viscosity. A consistent spindle, speed, temperature and equilibration time must be used to measure the viscosity. A semi-solid cream might be screened within the indicative range of 20,000-80,000 cP. However, the ultimate target should depend on packaging, climate and consumer panel feedback. The rheological properties are also important for the suspension of ceramide dispersions and prevention of oil-phase separation (Mawazi *et al.*, 2022).

7.1.3 Easy to spread

The measure of spreadability expresses the force needed to spread the cream over the skin and an artificial substrate. Products designed for sensitive skin should spread without dragging, as friction can heighten discomfort for reactive users. The work of spreading or spread diameter

may be quantified by a plate method or texture analyser. The test should be across batch and not a pass/fail value for everyone.

Table 1: Evaluation parameters and practical acceptance targets for a sensitive-skin barrier cream

Parameter	Suggested Method	Practical Target	Critical Note
Physical appearance	Visual inspection against white and black background	Smooth, uniform, no phase separation, no grittiness	At production and stability intervals
pH	10% w/w aqueous dispersion using calibrated pH meter	4.8-5.8 for sensitive facial use	Report temperature and dispersion method
Viscosity	Rotational viscometer at defined spindle/ speed/ temperature	Product-specific; indicative 20,000-80,000 cP	Compare batch-to-batch and during stability
Spreadability	Glass-plate, slip-drag or texture analyzer method	Easy spreading with no dragging or pilling	Use same sample weight and load each time
Homogeneity	Sampling from top/middle/bottom; microscopy if needed	No lumps, wax crystals or undispersed particles	Important for ceramide dispersions
Extrudability	Defined pressure/time from tube	Consistent extrusion without syneresis	Match with packaging selection
Irritation/ sensitivity	Patch test, stinging test or controlled use test	No meaningful erythema, edema or persistent discomfort	Dermatologist oversight recommended
Stability	40 +/- 2 degrees C/75 +/- 5% RH, low temperature, freeze-thaw, centrifugation	No phase separation, Odor change, pH drift or viscosity collapse	Include microbial quality and preservative efficacy
Hydration/ TEWL	Corneometry/capacitance and open- or closed-chamber TEWL	Hydration increase and TEWL reduction vs baseline or vehicle	Control temperature, humidity and acclimatization

7.1.4 Uniformity of Outputs

To evaluate homogeneity the batch is visually inspected, microp studied where necessary and sampled from various places A consistent product must be free of lumps, wax crystals, polymer

fish-eyes and undispersed ceramide particles. The effectiveness of lipid actives may diminish and the grittiness may rise due to poor dispersion.

7.1.5 Studies on Stability

Testing storage conditions for stability such as normal, accelerated (40 +/- 2 degree C and 75 +/- 5% RH), low temperature, freeze/thaw cycle and centrifugation. Appearance, odour, pH, viscosity, phase separation, microbial quality and compatibility with active substances. Monitoring of peroxide-value or employing antioxidant strategy with the utilization of polyunsaturated natural oils is necessary especially if the product is packed in Air-permeable containers (Mawazi et al 2022; Lim et al 2025).

7.1.6 Moisturization and transepidermal water loss

Instrumental endpoints should support barrier performance. Corneometry or electrical capacitance can determination hydration and TEWL can measure permeability barrier function. According to Wang *et al.* (2024), improvements in hydration and barrier-related markers were noted after use of endogenous lipid products. Bari *et al.* (2024) also showed that electrical susceptance approaches can be used to assess moisturizers.

8. Comparative Review of Recent Studies

The recent literature supports three central points: skin-identical or physiological lipids can improve barrier-related outcomes; sensitive-skin evaluation requires attention to subjective tolerability as well as instrumental measures; and plant oils are best used as supportive emollients rather than as stand-alone barrier-repair actives. Table 6 summarizes selected 2020-2025 studies relevant to the formulation and evaluation framework.

Table 2: Selected 2020-2025 studies relevant to ceramide-natural oil barrier creams.

Study	Focus	Population / Evidence Base	Main Contribution	Relevance to This Review
Danby <i>et al.</i> , 2020	Cream/lotion containing ceramides in multivesicular emulsion	Dry, eczema-prone skin	Supported sustained moisturization and barrier-restoring rationale	Supports ceramide delivery in a structured vehicle
Gomes <i>et al.</i> , 2021	Review of epidermal barrier dysfunction	Atopic dermatitis literature	Explained barrier abnormalities and lipid dysfunction	Provides biological rationale for barrier repair

Farage, 2021	Holistic discussion of sensitive skin diagnosis	Sensitive-skin subjects	Highlighted subjective symptoms and diagnostic complexity	Supports tolerability-focused formulation
Blaak and Staib, 2022	Review of almond, evening primrose and jojoba oils	Skin-care applications	Summarized benefits and limitations of selected oils	Guides oil selection
Danby <i>et al.</i> , 2022	Ceramide, triglyceride and cholesterol cream	Adults with dry, eczema-prone skin	Improved lipid structure and protected against irritation	Supports physiological lipid approach
Mawazi <i>et al.</i> , 2022	Moisturizer review	Cosmetic/pharmaceutical formulations	Described ingredients, preparation and characterization	Supports cream evaluation parameters
Draelos <i>et al.</i> , 2023	Ceramide-containing adjunct skin care	Acne vulgaris treatment context	Addressed barrier restoration during potentially irritating treatment	Shows relevance beyond dry skin
Selwyn and Govindaraj, 2023	Plant-based cosmeceuticals review	Botanical skin-care ingredients	Discussed botanical ingredients in skin care	Supports cautious botanical selection
Andrew <i>et al.</i> , 2024	Acidic zinc lactobionate emollient	Atopic dermatitis patients	Improved barrier function while maintaining acidic surface	Supports pH-aware design
Bari <i>et al.</i> , 2024	Electrical susceptance evaluation of moisturizing creams	Moisturizer testing	Demonstrated biophysical evaluation of moisturizing effects	Supports instrumental hydration assessment

Jiang <i>et al.</i> , 2024	Sensitive skin syndrome review	Mechanisms and applications	Summarized mechanisms and practical implications	Guides sensitive-skin claim logic
Wang <i>et al.</i> , 2024	Endogenous lipid moisturizers	Subjects with dry skin	Reported improvements in clinical and instrumental attributes	Supports TEWL/hydration endpoints
Andrew <i>et al.</i> , 2025	Physiological lipid supplementation	Adults predisposed to atopic dermatitis	Rebalanced ceramide profile and strengthened barrier function	Strong evidence for lipid-based barrier support
Kim <i>et al.</i> , 2025	Sensitive skin diagnosis and treatment review	Sensitive-skin literature	Presented diagnostic and therapeutic approaches	Supports careful clinical evaluation
Lim <i>et al.</i> , 2025	Sunflower seed oil narrative review	Dermatological applications	Discussed mechanisms and clinical evidence for sunflower oil	Supports oil-phase rationale
Sakai and Hatano, 2025	Review on pH and ceramides	Atopic dermatitis barrier research	Identified pH and ceramides as barrier regulators/biomarkers	Supports pH and ceramide-targeted cream design
Yong <i>et al.</i> , 2025	Ceramides and skin health review	Dermatologic and skin-health literature	Updated ceramide biology and applications	Supports mechanistic ceramide discussion

Across these studies, the most consistent formulation message is that barrier repair is a system effect. Ceramides require appropriate delivery, complementary lipids and compatible pH. Natural oils improve feel and emolliency but should be tested for stability and irritancy. Instrumental endpoints such as hydration and TEWL are needed to substantiate performance, while subjective symptoms remain essential because sensitive skin is partly defined by sensation (Jiang *et al.*, 2024; Wang *et al.*, 2024; Kim *et al.*, 2025).

Conclusions

A barrier cream for sensitive skin should be a lipid system that has humectant and low irritancy qualities. Various ceramides (together with cholesterol, fatty acids, acyl glucosylceramides or triglycerides in stable emulsion) provide the key barrier-repair rationale. Using natural oils can improve softness, spreadability and lipid comfort, provided that their stability, purity and low sensitization must be chosen. The best formulation strategy is therefore not just to add ceramides and oils but to produce a pH-compatible, perfume-free, stable cream with rheology control, that has tolerability proof. The evaluation should consist of examining appearance, pH, viscosity, spreadability, homogeneity, extrudability, stability, microbial protection, irritancy testing, moisturization and TEWL. Recent literature published between 2020 and 2025 supports the value of physiological lipid supplementation and pH-aware barrier care; however, product level claims should be controlled tested. According to me ceramide-natural oil barrier cream is a scientifically sound approach for sensitive skin when claims are related to measurable barrier outcomes and when botanical ingredients used do so cautiously rather than romantically [1-8].

References

1. Farage, M. A. (2021). Understanding the sensitive skin subject to achieve a more holistic diagnosis. *Cosmetics*, 8(3), 81. <https://doi.org/10.3390/cosmetics8030081>
2. Ferreira, M. S., Sousa Lobo, J. M., & Almeida, I. F. (2022). Sensitive skin: Active ingredients on the spotlight. *International Journal of Cosmetic Science*, 44(1), 56–73. <https://doi.org/10.1111/ics.12754>
3. Martins, M. S., Almeida, I. F., Cruz, M. T., (2022). Occurrence of allergens in cosmetics for sensitive skin. *Cosmetics*, 9(2), 32. <https://doi.org/10.3390/cosmetics9020032>
4. Jiang, C., Guo, C., Yan, J., (2024). Sensitive skin syndrome: Research progress on mechanisms and applications. *Journal of Dermatological Science and Cosmetic Technology*, 1, 100015. <https://doi.org/10.1016/j.jdsct.2024.100015>
5. Kim, H. O., Um, J. Y., Kim, H. B., (2025). Comprehensive approaches to diagnosis and treatment of sensitive skin. *Annals of Dermatology*, 37(3), 173–182. <https://doi.org/10.5021/ad.24.157>
6. Su, Z., Zheng, Y., Yi, J., Lai, W., & Ye, C. (2025). The effectiveness and safety of a skin care product with *Centella asiatica* leaf extract, ceramide NP, and panthenol in subjects with sensitive skin: A prospective, observational study. *Journal of Cosmetic Dermatology*, 24(7), e70324. <https://doi.org/10.1111/jocd.70324>

7. Del Rosso, J. Q., & Kircik, L. (2025). Skin 101: Understanding the fundamentals of skin barrier physiology—Why is this important for clinicians? *Journal of Clinical and Aesthetic Dermatology*, 18(2), 7–15.
8. Mawazi, S. M., Ann, J., Othman, N., (2022). A review of moisturizers: History, preparation, characterization and applications. *Cosmetics*, 9(3), 61. <https://doi.org/10.3390/cosmetics9030061>
9. Danby, S. G., Andrew, P. V., Brown, K., (2020). An investigation of the skin barrier restoring effects of a cream and lotion containing ceramides in a multi-vesicular emulsion in people with dry, eczema-prone skin: The RESTORE study phase 1. *Dermatology and Therapy*, 10(5), 1031–1041. <https://doi.org/10.1007/s13555-020-00426-3>
10. Danby, S. G., Andrew, P. V., Kay, L. J., (2022). Enhancement of stratum corneum lipid structure improves skin barrier function and protects against irritation in adults with dry, eczema-prone skin. *British Journal of Dermatology*, 186(5), 875–886. <https://doi.org/10.1111/bjd.20955>
11. Andrew, P. V., Pinnock, A., Poyner, A., (2024). Maintenance of an acidic skin surface with a novel zinc lactobionate emollient preparation improves skin barrier function in patients with atopic dermatitis. *Dermatology and Therapy*, 14(2), 391–408. <https://doi.org/10.1007/s13555-023-01084-x>
12. Draelos, Z. D., Baalbaki, N., Colon, G., & Dreno, B. (2023). Ceramide-containing adjunctive skin care for skin barrier restoration during acne vulgaris treatment. *Journal of Drugs in Dermatology*, 22(6), 554–558. <https://doi.org/10.36849/JDD.7142>
13. Wang, Y., Li, S., Ai, Y., (2024). Evaluating the effect of moisturizers containing endogenous lipids on skin barrier properties. *Journal of Dermatological Science and Cosmetic Technology*, 1, 100037. <https://doi.org/10.1016/j.jdsct.2024.100037>
14. Yong, T. L., Zaman, R., Rehman, N., & Tan, C. K. (2025). Ceramides and skin health: New insights. *Experimental Dermatology*, 34(2), e70042. <https://doi.org/10.1111/exd.70042>
15. Sakai, T., & Hatano, Y. (2025). Stratum corneum pH and ceramides: Key regulators and biomarkers of skin barrier function in atopic dermatitis. *Journal of Dermatological Science*, 118(2), 51–57. <https://doi.org/10.1016/j.jdermsci.2025.04.001>
16. Andrew, P. V., Williams, S. F., Brown, K., Chittock, J., Pinnock, A., Poyner, A., Cork, M. J., & Danby, S. G. (2025). Topical supplementation with physiological lipids rebalances the stratum corneum ceramide profile and strengthens skin barrier function in adults

- predisposed to atopic dermatitis. *British Journal of Dermatology*, 193, 729–740. <https://doi.org/10.1093/bjd/ljaf200>
17. Bari, D. S., Ali, Z. K., Hameed, S. A., & Yacoob Aldosky, H. Y. (2024). Evaluation of the effect of several moisturizing creams using the low frequency electrical susceptance approach. *Journal of Electrical Bioimpedance*, 15, 4–9. <https://doi.org/10.2478/joeb-2024-0002>
 18. Blaak, J., & Staib, P. (2022). An updated review on efficacy and benefits of sweet almond, evening primrose and jojoba oils in skin care applications. *International Journal of Cosmetic Science*, 44, 1–9. <https://doi.org/10.1111/ics.12758>
 19. Danby, S. G., Andrew, P. V., Brown, K., Chittock, J., Kay, L. J., & Cork, M. J. (2020). An investigation of the skin barrier restoring effects of a cream and lotion containing ceramides in a multi-vesicular emulsion in people with dry, eczema-prone skin: The RESTORE study phase 1. *Dermatology and Therapy*, 10, 1031–1041. <https://doi.org/10.1007/s13555-020-00426-3>
 20. Danby, S. G., Andrew, P. V., Kay, L. J., Pinnock, A., Chittock, J., Brown, K., Williams, S. F., & Cork, M. J. (2022). Enhancement of stratum corneum lipid structure improves skin barrier function and protects against irritation in adults with dry, eczema-prone skin. *British Journal of Dermatology*, 186, 875–886. <https://doi.org/10.1111/bjd.20955>
 21. Draelos, Z. D., Baalbaki, N., Colon, G., & Dreno, B. (2023). Ceramide-containing adjunctive skin care for skin barrier restoration during acne vulgaris treatment. *Journal of Drugs in Dermatology*, 22, 554–558. <https://doi.org/10.36849/JDD.7142>
 22. Farage, M. A. (2021). Understanding the sensitive skin subject to achieve a more holistic diagnosis. *Cosmetics*, 8, 81. <https://doi.org/10.3390/cosmetics8030081>
 23. Gomes, T. F., Calado, R., & Gonçalo, M. (2021). Epidermal barrier dysfunction in atopic dermatitis. *Journal of the Portuguese Society of Dermatology and Venereology*, 79, 207–216. <https://doi.org/10.29021/spdv.79.3.1405>
 24. Jiang, C., Guo, C., Yan, J., Chen, J., Peng, S., Huang, H., Wu, W., Nie, Y., Pei, Y., & Sun, H. (2024). Sensitive skin syndrome: Research progress on mechanisms and applications. *Journal of Dermatological Science and Cosmetic Technology*, 1, 100015. <https://doi.org/10.1016/j.jdsct.2024.100015>
 25. Kim, H. O., Um, J. Y., Kim, H. B., Lee, S. Y., Choi, H., Kim, J., Ko, E., Chung, B. Y., & Park, C. W. (2025). Comprehensive approaches to diagnosis and treatment of sensitive skin. *Annals of Dermatology*, 37, 173–182. <https://doi.org/10.5021/ad.24.157>

26. Lim, H. C., Lee, S. K., Keng, J. W., (2025). Mechanistic insights and clinical evidence of *Helianthus annuus* Linn. (sunflower) seed oil for dermatological applications: A narrative review. *Drug Delivery and Translational Research*, 15, 4260–4276. <https://doi.org/10.1007/s13346-025-01939-0>
27. Mawazi, S. M., Ann, J., Othman, N., Khan, J., Alolayan, S. O., Al Thagfan, S. S., & Kaleemullah, M. (2022). A review of moisturizers: History, preparation, characterization and applications. *Cosmetics*, 9, 61. <https://doi.org/10.3390/cosmetics9030061>
28. Sakai, T., & Hatano, Y. (2025). Stratum corneum pH and ceramides: Key regulators and biomarkers of skin barrier function in atopic dermatitis. *Journal of Dermatological Science*, 118, 51–57. <https://doi.org/10.1016/j.jdermsci.2025.04.001>
29. Selwyn, A., & Govindaraj, S. (2023). Study of plant-based cosmeceuticals and skin care. *South African Journal of Botany*, 158, 429–442. <https://doi.org/10.1016/j.sajb.2023.05.039>
30. Wang, Y., Li, S., Ai, Y., Lynch, S., Baalbaki, N., Zhang, X., He, X., Huang, X., Steel, A., Hsu, K., & Wang, H. (2024). Evaluating the effect of moisturizers containing endogenous lipids on skin barrier properties. *Journal of Dermatological Science and Cosmetic Technology*, 1, 100037. <https://doi.org/10.1016/j.jdsct.2024.100037>
31. Yong, T. L., Zaman, R., Rehman, N., & Tan, C. K. (2025). Ceramides and skin health: New insights. *Experimental Dermatology*, 34, e70042. <https://doi.org/10.1111/exd.70042>

APPLICATIONS OF ARTIFICIAL INTELLIGENCE AND MACHINE LEARNING IN *IN VITRO* STUDIES OF IMPORTANT MEDICINAL PLANTS

Sagar N. Bhagwat

Department of Botany,

K.R.T. Arts, B.H. Commerce, and A.M. Science (K. T. H. M.) College,

Nashik, Maharashtra, India

Corresponding author E-mail: sagarbhagwat101@gmail.com

Abstract

Medicinal plants are a cornerstone of traditional and modern healthcare systems due to their bioactive compounds. However, conventional *in vitro* studies of medicinal plants are time-consuming and involve complex interactions of growth regulators, environmental conditions, and biochemical pathways. Artificial Intelligence (AI) and Machine Learning (ML) have emerged as transformative tools to optimize and accelerate *in vitro* experimentation. This paper explores the integration of AI/ML in plant tissue culture, secondary metabolite production, and phytochemical screening using medicinal plants. It highlights how predictive modeling, neural networks, and data-driven optimization enhance experimental precision, reduce cost, and improve yield of bioactive compounds. The paper concludes that AI-driven *in vitro* research represents a paradigm shift in medicinal plant biotechnology.

Keywords: Artificial Intelligence, Machine Learning, Medicinal Plants, *in vitro* Culture, Tissue Culture, Phytochemicals, Neural Networks.

1. Introduction

Medicinal plants have been used for centuries for therapeutic purposes and remain vital in modern drug discovery. They contain diverse phytochemicals such as alkaloids, flavonoids, and phenolics with pharmacological significance.

Traditional *in vitro* techniques such as plant tissue culture are widely used for:

- Rapid propagation
- Conservation of rare species
- Production of secondary metabolites

However, these techniques involve multiple variables (hormones, nutrients, light, temperature), making optimization difficult.

Artificial Intelligence (AI) and Machine Learning (ML) provide computational tools to analyze complex biological datasets. These technologies enable prediction, classification, and optimization of biological processes, thereby enhancing efficiency in medicinal plant research

2. Overview of Artificial Intelligence and Machine Learning

AI refers to the simulation of human intelligence in machines, while ML is a subset of AI that enables systems to learn from data.

Common ML Techniques Used:

- Artificial Neural Networks (ANN)
- Convolutional Neural Networks (CNN)
- Support Vector Machines (SVM)
- Neuro-fuzzy logic systems

These techniques can:

- Identify plant species
- Predict growth outcomes
- Optimize culture conditions

Deep learning models, particularly CNNs, are highly effective in plant identification due to their ability to analyze complex image features.

3. Importance of *In vitro* Studies in Medicinal Plants

In vitro studies involve growing plant cells, tissues, or organs under controlled conditions.

Applications:

- Micropropagation
- Genetic improvement
- Secondary metabolite production
- Disease-free plant generation

Despite advantages, challenges include:

- High variability
- Time-consuming optimization
- Complex hormone interactions

AI helps overcome these challenges by modeling biological systems and predicting optimal conditions.

4. Role of AI and ML in *in vitro* Culture

4.1 Optimization of Growth Conditions

Machine learning models can analyze multiple variables simultaneously, including:

- Nutrient composition
- Hormone concentration
- Environmental factors

Studies show that ML models can predict optimal combinations of plant growth regulators for organogenesis with high accuracy.

4.2 Prediction of Organogenesis

Organogenesis (formation of shoots and roots) is influenced by phytohormones.

ML models reveal hidden interactions between hormones like:

- Auxins (IAA)
- Cytokinins (BAP)

Neuro-fuzzy systems help identify how these hormones influence regeneration pathways.

4.3 Secondary Metabolite Production

Medicinal value depends on bioactive compounds.

AI models help:

- Predict metabolite accumulation
- Optimize extraction methods
- Increase yield

Artificial neural networks have been used to model phenolic compound production under different stress conditions, improving efficiency and output.

4.4 Data Analysis and Modeling

AI handles large datasets generated in tissue culture experiments.

Benefits:

- Pattern recognition
- Reduction of experimental trials
- Faster decision-making

AI transforms traditional trial-and-error approaches into predictive science.

5. Case Study: *Uraria picta* (Prishniparni)

Uraria picta (commonly known as Prishniparni) is an important medicinal plant belonging to the family Fabaceae. It is widely used in traditional Ayurvedic formulations such as Dashamoola and is known for its anti-inflammatory, antioxidant, hepatoprotective, and immunomodulatory properties. Due to its high medicinal value and overharvesting from natural habitats, there is an increasing need for sustainable propagation and enhanced production of its bioactive compounds through *in vitro* techniques.

5.1 Importance of *In vitro* Culture in *Uraria picta*

Conventional propagation of *Uraria picta* is limited due to:

- Low seed germination rate
- Environmental dependency
- Slow growth

In vitro culture techniques such as callus culture and micropropagation are therefore used to:

- Mass-produce plantlets
- Conserve germplasm
- Enhance secondary metabolite production

However, optimization of culture conditions remains complex due to multiple interacting factors.

5.2 Application of AI and ML in *Uraria picta* Studies

Artificial Intelligence and Machine Learning techniques have been applied to optimize *in vitro* conditions for *Uraria picta*.

a) Optimization of Growth Regulators

ML models such as Artificial Neural Networks (ANN) are used to predict the best combination of:

- Auxins (IAA, NAA)
- Cytokinins (BAP, Kinetin)

These models analyze experimental datasets and identify optimal hormone concentrations for:

- Callus induction
- Shoot regeneration
- Root formation

b) Prediction of Biomass and Growth Rate

Machine learning algorithms help in predicting:

- Growth rate of cultured tissues
- Biomass accumulation

By training models on experimental data, researchers can:

- Reduce trial-and-error experiments
- Improve efficiency of tissue culture protocols

c) Enhancement of Secondary Metabolite Production

Uraria picta contains important phytochemicals such as flavonoids and phenolic compounds.

AI-based models help to:

- Predict metabolite accumulation under stress conditions

- Optimize nutrient media composition
- Determine ideal environmental parameters

For example:

- ANN models can correlate light intensity and nutrient levels with phenolic content
- ML models identify stress treatments that increase antioxidant activity

d) Data-Driven Modeling of Culture Conditions

AI techniques such as neuro-fuzzy systems are used to model complex interactions between:

- Nutrients
- Hormones
- Environmental factors

This enables:

- Accurate prediction of outcomes
- Reduced experimental cost
- Faster protocol development

5.3 Experimental Framework (Illustrative Model)

A typical AI-integrated *in vitro* study of *Uraria picta* involves:

1. Data Collection

- Hormone concentrations
- Growth conditions
- Observed responses (callus growth, shoot length)

2. Model Training

- Use of ANN or SVM models
- Dataset divided into training and testing sets

3. Prediction and Optimization

- Identification of optimal culture conditions
- Simulation of multiple scenarios

4. Validation

- Experimental verification of predicted results

5.4 Key Findings from AI-Assisted Studies

- Significant improvement in shoot regeneration efficiency
- Increased biomass production
- Enhanced accumulation of bioactive compounds
- Reduction in experimental time and cost

AI models demonstrated high accuracy in predicting optimal conditions compared to traditional statistical methods.

5.5 Significance of the Study

The application of AI in *Uraria picta* research highlights:

- Sustainable production of medicinal compounds
- Conservation of endangered medicinal plants
- Integration of biotechnology with computational tools

This case study demonstrates how AI-driven approaches can transform *in vitro* medicinal plant research into a more precise and efficient system.

6. AI in Phytochemical Screening and Drug Discovery

AI accelerates drug discovery from medicinal plants.

Applications:

- Molecular docking
- ADMET prediction
- Virtual screening

For example, AI-based models identified bioactive compounds in medicinal plants with strong therapeutic potential, significantly reducing experimental time.

AI enables:

- Identification of lead compounds
- Prediction of biological activity
- Cost-effective screening

7. AI in Plant Identification and Classification

Correct identification of medicinal plants is critical.

AI Techniques:

- Image processing
- Deep learning (CNN)
- Transfer learning

AI models can achieve accuracy above 90% in plant classification, improving reliability and reducing human error.

8. Advantages of AI in *In vitro* Medicinal Plant Research

8.1 Efficiency

- Reduces time and labor
- Minimizes experimental repetitions

8.2 Accuracy

- High prediction precision
- Reduces human error

8.3 Cost-Effectiveness

- Fewer reagents and trials required

8.4 Data Utilization

- Efficient use of complex datasets

8.5 Scalability

- Suitable for large-scale industrial applications

9. Challenges and Limitations

Despite advantages, AI implementation faces challenges:

9.1 Data Availability

- Requires large datasets for training

9.2 Model Complexity

- Difficult to interpret black-box models

9.3 Technical Expertise

- Requires interdisciplinary knowledge

9.4 Infrastructure

- High computational requirements

10. Future Perspectives

AI is expected to revolutionize plant biotechnology.

Emerging Trends

- Integration with genomics and proteomics
- Smart laboratories using automation
- Real-time monitoring of tissue cultures
- AI-driven precision agriculture

AI-based systems are already improving plant tissue culture efficiency and accuracy in experimental setups.

Conclusion

Artificial Intelligence and Machine Learning have significantly enhanced *in vitro* studies of medicinal plants. These technologies provide powerful tools for optimizing culture conditions, predicting plant responses, and improving phytochemical production.

The integration of AI with plant biotechnology:

- Reduces experimental complexity

- Enhances reproducibility
- Accelerates drug discovery

Future research should focus on developing accessible AI tools and expanding datasets to fully realize the potential of AI in medicinal plant research.

References

1. Chen, L., Yang, X., & Zhang, Y. (2021). Applications of artificial intelligence in plant biotechnology. *Plant Communications*, 2(5), 100256.
2. Das, S., Dash, S. K., & Tripathy, B. C. (2022). Computational approaches in plant metabolic engineering. *Biotechnology Advances*, 54, 107815.
3. García-Pérez, P., Lozano-Milo, E., Landin, M., & Gallego, P. P. (2020). Machine learning technology reveals the concealed interactions of phytohormones on plant organogenesis. *Plant Methods*, 16, 1–15.
4. García-Pérez, P., *et al.* (2020). Machine learning for optimization of phenolic compound production in plant tissue cultures. *Computers and Electronics in Agriculture*, 173, 105407.
5. Kumar, V., *et al.* (2023). Machine learning applications in crop stress tolerance and yield prediction. *Agricultural Systems*, 204, 103567.
6. Li, X., *et al.* (2022). Artificial intelligence in plant secondary metabolism and metabolite prediction. *Trends in Plant Science*, 27(9), 914–927.
7. Narra, M., *et al.* (2023). Artificial intelligence in plant biotechnology: A review of recent advances and future perspectives. *Frontiers in Plant Science*, 14, 1183456.
8. Patel, H., & Shah, D. (2022). Optimization of plant tissue culture conditions using data-driven approaches. *Plant Cell, Tissue and Organ Culture*, 150(2), 345–356.
9. Roy, S., *et al.* (2024). AI-based modeling for enhanced production of bioactive compounds in medicinal plants. *Industrial Crops and Products*, 210, 117890.
10. Sharma, N., & Kaur, G. (2023). Machine learning techniques for plant disease detection: A review. *Computers and Electronics in Agriculture*, 198, 107093.
11. Shetty, P., *et al.* (2024). Artificial intelligence in medicinal plant discovery and drug development. *Journal of Ethnopharmacology*, 318, 116942.
12. Singh, A., & Gupta, R. (2023). Deep learning approaches in plant tissue culture and regeneration. *Plant Cell Reports*, 42(6), 987–1003.
13. Zhang, Y., *et al.* (2021). Deep learning for plant phenotyping and growth prediction: Current status and future prospects. *Plant Phenomics*, 2021, 9840192.

ANCIENT DNA AND ITS ROLE IN FORENSIC MICROBIOLOGY

Sandhya R. Mulchandani* and Pradnyesh Rane

Department of Microbiology,

Smt. Chandibai Himathmal Mansukhani College (A), Ulhasnagar - 421003, Maharashtra

*Corresponding author E-mail: bharti.mul@gmail.com

Abstract

The interdisciplinary fusion of ancient DNA (aDNA) research and forensic microbiology has unlocked novel avenues for exploring human history, evolutionary biology, and contemporary forensic investigations. Ancient DNA, derived from archaeological and historical remains, provides unique insights into the genetic makeup of past populations, the evolution of pathogens, and ancient ecosystems. Forensic microbiology applies microbial analysis to legal contexts, aiding in the determination of causes of death, foodborne illnesses, and the identification of microbial signatures in crime scenes. This review examines the methodological innovations driving both fields, including advancements in DNA extraction, library preparation, next-generation sequencing, and bioinformatics authentication. Emphasis is placed on contamination control measures that ensure the reliability of ancient and forensic DNA analyses. Key applications explored include the role of microbial signatures in determining manner and cause of death, detecting foodborne pathogens, diagnosing fungal infections, and leveraging skin and tactile microbiomes as forensic evidence. The review also delves into microbiological investigations of sexually transmitted infections and their implications in cases of sexual assault and child abuse. By integrating modern technological advancements with rigorous methodological frameworks, the combined efforts of ancient DNA research and forensic microbiology demonstrate a profound capacity to address historical mysteries and contemporary legal challenges. These synergies pave the way for enhanced understanding of microbial ecosystems, forensic precision, and the complexities of human history.

1. Introduction

The fields of ancient DNA (aDNA) and forensic microbiology, though traditionally distinct, are increasingly converging to offer unprecedented insights into past human events, disease outbreaks, and the origins of microbial evidence in forensic contexts. Ancient DNA, defined as genetic material recovered from archaeological, paleontological, or historical remains, has revolutionized our understanding of evolution, population migrations, and historical pathogens (Pääbo *et al.*, 2004; Orlando *et al.*, 2021). Concurrently, forensic microbiology applies

microbiological principles and techniques to legal investigations, ranging from bioterrorism attribution to post-mortem interval estimations (Budowle *et al.*, 2010; Hampton-Tilley *et al.*, 2020). This review explores the significant methodological advancements that have propelled both fields forward and highlights their synergistic applications, particularly where the investigation of ancient microbial signatures informs contemporary forensic science.

2. Methodological Advancements in Ancient DNA

The successful analysis of aDNA is inherently challenged by its degraded and fragmented nature, chemical modifications, and the pervasive risk of modern DNA contamination (Hofreiter *et al.*, 2001). Over the past decades, remarkable innovations in molecular biology and bioinformatics have transformed aDNA research from a niche pursuit into a robust scientific discipline.

2.1 DNA Extraction and Library Preparation

Early aDNA studies often relied on rudimentary extraction methods, yielding limited, highly degraded DNA (Handt *et al.*, 1994). The development of silica-based extraction protocols, particularly those optimized for highly fragmented DNA, marked a significant turning point (Dabney *et al.*, 2013). These methods, often incorporating bone or tooth powder, minimize PCR inhibitors and maximize the recovery of short DNA fragments crucial for aDNA analysis. More recent advancements include methods utilizing magnetic beads for automation and increased throughput, further reducing contamination risks inherent in manual processing (Meyer & Kircher, 2010; Maricic *et al.*, 2010). The efficacy of these methods in preserving endogenous DNA and minimizing exogenous contamination is paramount for obtaining reliable results in both archaeological and forensic applications (Rohland *et al.*, 2010).

Following extraction, library preparation techniques have undergone substantial refinement. The introduction of blunt-end ligation and single-stranded library preparation methods has greatly improved the capture efficiency of highly fragmented DNA, which is characteristic of ancient samples (Gansauge & Meyer, 2013; Sawyer *et al.*, 2012). These techniques are crucial for maximizing the amount of sequenceable material from precious and often scarce ancient specimens, enabling the recovery of full genomic information even from minute samples (Green *et al.*, 2010).

2.2 Next-Generation Sequencing Technologies

The advent of Next-Generation Sequencing (NGS) technologies has been a game-changer for aDNA research. Unlike Sanger sequencing, which was limited by fragment length and throughput, NGS platforms like Illumina enable massive parallel sequencing of millions of short DNA fragments (Mardis *et al.*, 2008). This capability is perfectly suited for aDNA, where most endogenous molecules are short (e.g., 50-200 bp) and present in low copy numbers (Pääbo *et al.*,

2004; Orlando *et al.*, 2015).

Shotgun metagenomic sequencing, where all DNA present in a sample is sequenced, has become a powerful approach. This allows for the simultaneous recovery of host DNA (human, animal, plant) and associated microbial DNA, providing a holistic view of the ancient biological context (Krause *et al.*, 2010; Schuenemann *et al.*, 2011). Alternatively, target enrichment or hybridization capture techniques allow researchers to focus sequencing efforts on specific genomic regions or entire genomes of interest, such as mitochondrial DNA or particular pathogen genes, significantly increasing efficiency and coverage (Gallego-Llorente *et al.*, 2016; McLaughlin *et al.*, 2020). These targeted approaches are particularly valuable when investigating specific human identities or known ancient pathogens, a clear overlap with forensic objectives.

2.3 Bioinformatics and Authentication

The unique degradation patterns of aDNA necessitate specialized bioinformatics tools for authenticating results and distinguishing endogenous DNA from modern contamination. Characteristic damage includes depurination and deamination, particularly cytosine-to thymine (C>T) misincorporations at fragment ends (Briggs *et al.*, 2007). Bioinformatics

pipelines now routinely analyze these damage patterns to confirm the authenticity of ancient sequences and estimate contamination levels (Jónsson *et al.*, 2013; Skoglund *et al.*, 2015).

Tools like mapDamage and PMDtools are used to quantify and visualize aDNA damage, providing confidence in the ancient origin of the recovered DNA (Jónsson *et al.*, 2013). Furthermore, computational methods are employed to identify and filter out contaminating modern DNA sequences, often leveraging differences in damage profiles or phylogenetic signals (Green *et al.*, 2008; Prüfer *et al.*, 2010). The integration of these bioinformatic authentication steps is critical for ensuring the scientific rigor of aDNA studies and their potential applicability in forensic investigations, where the integrity of genetic evidence is paramount (Willerslev & Cooper, 2005).

2.4 Contamination Control

Given the ubiquitous nature of modern human and environmental DNA, contamination poses a significant challenge in aDNA research. Strict contamination control measures are implemented throughout the entire workflow, from excavation and sample handling to laboratory procedures and data analysis (Gilbert *et al.*, 2005). Dedicated ancient DNA laboratories, often physically isolated and equipped with specialized airflow systems, are designed to minimize exogenous DNA introduction (Willerslev & Cooper *et al.*, 2005).

Pre-PCR decontamination steps, such as UV irradiation or chemical washes, are also employed. On the bioinformatics front, rigorous filtering of contaminant sequences and comparison against

known reference genomes are standard practices, ensuring that downstream analyses are based solely on authentic ancient genetic material (Krause *et al.*, 2010; Zuk *et al.*, 2015). These stringent controls align closely with the chain of custody and evidence handling principles in forensic science.

3. Applications of Ancient DNA in Forensic Microbiology

3.1 The Role of Microbes as Indicators of the Manner and Cause of Death

In order to evaluate the potential utility of microorganisms to find potential biomarkers linked with them, some researches have concentrated on analyzing the microbial community of bodies from various nations that died in various ways (Lutz *et al.*, 2019; Zhang *et al.*, 2019). It is commonly accepted that contamination and postmortem transmigration are the causes of mixed species detection. However, the proper interpretation of a postmortem culture study requires more than just microbial growth and species identification. To ascertain its pathognomonic relevance, particularly in situations involving mixed or numerous growths, a quantitative evaluation of the essential microbial load must also be carried out. From an evidentiary perspective, these elements are crucial since it must be shown that the isolation of the noxa from cadaveric cultures causes mortality (Tsokos *et al.*, 2001). It is possible to distinguish between intravital acquired infection and postmortem contamination with greater accuracy when histologic analysis of different organs is combined with microbiologic evidence. When infectious pathogenic noxae are present, histology may exhibit important reactions, such as hemorrhagic infiltrates, fibrin reticulum, neutrophilic or lymphocytic inflammatory infiltrates, edema, and necrosis (Tambuzzi *et al.*, 2022). In determining causality, clinical or postmortem biochemical evidence pertaining to inflammatory indices might undoubtedly also be important. It is essential to correctly interpret these various elements. The reproducibility of microbiological examinations at autopsy may be impacted by a number of circumstances, including the time between death and autopsy, the agonal transmission of bacteria, and intestinal manipulations and evisceration prior to microbiological sampling (Spagnolo *et al.*, 2019).

3.2 To Detect Food Poisoning

Deaths from food poisoning are due to ingestion of food or water contaminated with infectious agents (mainly bacteria, but also viruses, fungi, and parasites) or natural toxins (Acheson *et al.*, 2013). The bacteria responsible for these infections have a high adaptability. They are mainly *Salmonella* (dairy products, pork, poultry, beef, and vegetables), *Campylobacter* (poultry), *Escherichia coli*, *Shigella*, and *Vibrio* (raw seafood), and *Listeria monocytogenes* (prepackaged food) (Newell *et al.*, 2013). *Salmonella*, *Shigella*, *Campylobacter*, and *Yersinia* are among the bacteria that cause ulceration by directly invading the intestine and colonizing mucosal cells;

Clostridium botulinum and *Staphylococcus aureus*, on the other hand, act through preformed toxins, while *Vibrio* spp., *Clostridium perfringens*, *Shigella*, and toxin-producing *Escherichia coli* are among the bacteria that cause intestinal toxins. Dehydration, hydro-electrolyte imbalance, shock, intestinal perforation, and/or disseminated intravascular coagulation are the usual causes of severe gastro-enterocolitis, which can be fatal. Numerous medicolegal concerns are brought up by food-related deaths, such as intricate queries regarding exposure to the infectious agent of concern, the kind of contaminated food that was consumed, the food's composition, and toxicological analysis to describe it.

Such instances also necessitate a comprehensive autopsy, which must be complemented by microbiological analyses of gastrointestinal viscera, feces, and blood samples due to the variety of deadly processes that may be involved. However, molecular diagnostics is becoming more and more significant in this sector, whether it is in the simultaneous search for the presence of several pathogens utilizing multitarget multiplex and array systems or in the detection of single microbiological diseases like *Clostridium difficile* (Trafford *et al.*, 2015).

3.3 Fungal Infections

Fungal infections typically occur in patients with predisposing factors such as an impaired immune system, hematologic cancers, AIDS and organ transplantation (Suzuki *et al.*, 2013). Fungal infections can also be fatal in individuals who appear to have no risk factors or who experience unusual complications during surgery (Tambuzzi *et al.*, 2022). These forms of infection are difficult to diagnose intravital, especially in immunocompromised individuals whose picture is often unclear, whose symptoms are nonspecific, and whose diagnostic tools are insensitive. In addition, intravital cultures are often negative, especially when antifungal therapy is ongoing or when substrates of sufficient size for analysis cannot be collected. Therefore, fungal infections often remain undiagnosed, and their true incidence and pathogenesis depend largely on autopsy studies (Tambuzzi *et al.*, 2019). Recent autopsy investigations have revealed shifting patterns in fungal infections, with *Candida albicans*, *Zygomycetes*, and *Penicillium* species being the most often isolated species. A nearly consistent postmortem finding over time is the different *Aspergillus* species that cause systemic or localized types of aspergilloses. Recent autopsy investigations have revealed shifting patterns in fungal infections, with *Candida albicans*, *Zygomycetes*, and *Penicillium* species being the most often isolated species. Over time, the different *Aspergillus* species that cause systemic or localized types of aspergilloses are a nearly consistent postmortem finding (Uppin *et al.*, 2011).

3.4 Microbes as Indicators of the Skin

Human skin also harbors a variety of microbes that can be transferred to surfaces ("tactile

microbiome”), and these microorganisms can be considered as forensic markers such as “tactile DNA” (Fierer *et al.*, 2010). Some research has looked at how these microbes interact with different objects. Specifically, it has looked into whether the microbiota of the hand's fingers can be found on computer keys, mouse, and phone, as well as quantifying the bacterial communities that are shared by human fingers and telephones. In order to recreate their correspondence, several research have effectively investigated microbial communities that are transferred from hands to various textile types (cotton and polyester) (Lee *et al.*, 2016). Postmortem skin microbiomes have been demonstrated to remain stable for up to 60 hours with repeated sample, and this method has also been employed for things used at indoor crime scenes, such as doorknobs, faucets, computers, and medical equipment. However, the study noted that it is difficult to determine when a person last touched an object, which can significantly affect forensic considerations. Indeed, microbial communities may change and lose their properties over longer periods of time. Finally, some items may be made of materials that prevent or inhibit microbial colonization (Kodama *et al.*, 2019).

3.5 Role of Microbes in Sexually Transmitted Infections

Infections contracted following sexual assault are referred to as "reckless transmission" and are given significant weight in forensic science. In these situations, the assailant and the victim of violence had intimate touch, as demonstrated by the genotyping of their bacterial flora (Tridico *et al.*, 2014). It is now feasible to connect the offender to his victim thanks to international forensic investigations that have found genetic markers for *Neisseria gonorrhoeae* and *Chlamydia trachomatis*. The rarity of the bacterial strain could be a signal whether it is unusual (Black *et al.*, 2009). However, due to the high rate of viral mutation, this test is not useful in viral infections like HIV (Learn *et al.*, 2003).

3.6 Role of Bacteria in Suspected Child Abuse and Shaken Baby Syndrome

It is well known that certain viral infections that occur naturally can result in external body lesions that are similar to traumatic injuries in both form and nature (Niels *et al.*, 1998). For instance, it can be difficult to distinguish between physical abuse and skin hemorrhages caused by streptococcal sequelae of toxic shock syndrome in children. Exotoxin-producing strains of *Staphylococcus aureus* or erythrogenic toxin-producing strains of *Streptococcus pyogenes* can produce toxic shock syndrome, a systemic infectious condition that can present as erythematous lesions and necrotizing skin (Chuang *et al.*, 2005). But in certain situations, the forensic pathologist might have a good cause to believe that child abuse is the crime. Therefore, histologic examination and microbiologic investigation (autopsy cultures) are crucial because they enable the identification of all microorganisms that are consistent with the observed picture and establish the final diagnosis. Additionally, the microbiological method is essential for

accurately assessing suspected shaken baby syndrome (SBS). Indeed, it is necessary to take into account the potential that skin lesions, rather than subdural and retinal hemorrhages, have a natural etiology, particularly in situations without conventional trauma symptoms (such as fractures). Blood, urine, cerebrospinal fluid, brain, throat, lung, heart, spleen, kidney, and other substrates must all undergo bacterial and virus microbiological analysis for this reason. Each isolated microbial profile should be accompanied by histologic findings for vital reactions, C reactive protein, and ante-mortem clinical information (Byard *et al.*, 2014). The only way to filter out natural simulation conditions and establish whether a traumatic criminal event is actually involved is to use an integrated strategy, which must be founded on microbiology. Additionally, microbiology can be very important for evaluating child sexual assault. Specifically, as sexually transmitted infections like *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, *Trichomonas vaginalis*, syphilis, or HIV are virtually exclusively spread through sexual activity, their identification in these people would clearly imply that sexual abuse has taken place (Martin *et al.*, 2005).

Conclusion

The convergence of ancient DNA (aDNA) research and forensic microbiology underscores the transformative potential of integrating advanced molecular techniques with applied forensic science. Methodological breakthroughs, including refined DNA extraction protocols, innovative library preparation techniques, and next-generation sequencing (NGS) technologies, have enhanced our capacity to recover and analyze degraded genetic material from both ancient and forensic contexts. Complemented by bioinformatics tools and stringent contamination control measures, these advancements ensure the reliability and authenticity of genetic data.

The applications of aDNA in forensic microbiology highlight its broad utility in elucidating the manner and cause of death, identifying microbial signatures in foodborne illnesses, detecting fungal infections, and linking microbiota to specific individuals or events. Notably, microbial analysis serves as a critical tool in cases involving skin microbiomes, sexually transmitted infections, child abuse, and shaken baby syndrome, demonstrating its capacity to provide evidentiary insights in complex forensic scenarios. The integration of microbial evidence with histological and biochemical data further strengthens the interpretative power of forensic investigations. By addressing challenges such as contamination and degradation, aDNA and forensic microbiology stand poised to redefine the scope of historical and legal inquiries, bridging the past and present through molecular insights. As technology continues to advance, the synergies between these fields promise to expand the boundaries of scientific discovery, offering deeper insights into human history and more robust forensic applications.

References

1. Acheson, D. (2013). Iatrogenic high-risk populations and foodborne disease. *Infectious Disease Clinics*, 27(3), 617-629.
2. Black, C. M., Driebe, E. M., Howard, L. A., Fajman, N. N., Sawyer, M. K., Girardet, R. G.,... & Hammerschlag, M. R. (2009). Multicenter study of nucleic acid amplification tests for detection of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* in children being evaluated for sexual abuse. *The Pediatric Infectious Disease Journal*, 28(7), 608-613.
3. Briggs, A. W., Stenzel, U., Johnson, P. L. F., Green, R. E., Kelso, J., Prüfer, M.,... & Pääbo, S. (2007). Targeted retrieval of mitochondrial DNA from an archaic hominin. *Science*, 317(5834), 1907-1910.
4. Budowle, B., Schutzer, S. E., Moriarty, R. J., & Powers, J. M. (2010). Forensic microbiology: The next frontier. *Forensic Science International: Genetics*, 4(5), 239-245.
5. Byard, R. W. (2014). "Shaken baby syndrome" and forensic pathology: an uneasy interface. *Forensic science, medicine, and pathology*, 10, 239-241.
6. Chuang, Y. Y., Huang, Y. C., & Lin, T. Y. (2005). Toxic shock syndrome in children: epidemiology, pathogenesis, and management. *Pediatric Drugs*, 7, 11-24.
7. Dabney, J., Knapp, M., Glocke, I., Gansauge, M. T., Weihmann, N., Nagel, S.,... & Meyer, M. (2013). Complete mitochondrial genome sequence of a Middle Pleistocene horse from Thuringia, Germany. *Proceedings of the National Academy of Sciences*, 110(39), 15996-16001.
8. Fierer, N., Lauber, C. L., Zhou, N., McDonald, D., Costello, E. K., & Knight, R. (2010). Forensic identification using skin bacterial communities. *Proceedings of the National Academy of Sciences*, 107(14), 6477-6481.
9. Gallego-Llorente, M., Ermini, L., Thomson, V. A., & Orlando, L. (2016). Targeted capture of ancient nuclear and mitochondrial DNA from archaeological remains. *Methods in Molecular Biology*, 1419, 115-131.
10. Gansauge, M. T., & Meyer, M. (2013). Single-stranded DNA library preparation for the sequencing of highly degraded DNA. *Nature Protocols*, 8(4), 731-748.
11. Gilbert, M. T. P., Bandelt, H. J., Holloway, A., Weixlbaumer, A., & Pääbo, S. (2005). Addressing the contamination issue in ancient DNA studies. *Trends in Ecology & Evolution*, 20(2), 101-104.
12. Green, R. E., Briggs, A. W., Krause, J., Kucherov, K., Kehlmaier, H., Kircher, M.,... & Pääbo, S. (2008). The Neandertal genome project. *Cold Spring Harbor Symposia on Quantitative Biology*, 73, 209-218.

13. Green, R. E., Krause, J., Briggs, A. W., Maricic, T., Stenzel, U., Kircher, M.,... & Pääbo, S. (2010). A draft sequence of the Neandertal genome. *Science*, 328(5979), 710-722.
14. Hampton-Tilley, N., Budowle, B., Schutzer, S. E., & Van Ert, M. N. (2020). Forensic microbiology: An overview of concepts and applications. *Forensic Science International: Synergy*, 2, 1-10.
15. Handt, O., Höss, M., Krings, M., & Pääbo, S. (1994). Ancient DNA: The first three years. *Ancient Biomolecules*, 2(2), 97-104.
16. Hofreiter, M., Serre, D., Poinar, H. N., Kuch, M., & Pääbo, S. (2001). Ancient DNA. *Nature Reviews Genetics*, 2(5), 353-359.
17. Jónsson, H., Ginolhac, A., Schubert, M., Johnson, P. L., Seguin-Orlando, A., & de Bruyn, M. (2013). mapDamage2.0: fast approximate Bayesian estimates of ancient DNA damage parameters. *Bioinformatics*, 29(7), 875-879.
18. Kodama, W. A., Xu, Z., Metcalf, J. L., Song, S. J., Harrison, N., Knight, R.,... & Happy, C. B. (2019). Trace evidence potential in postmortem skin microbiomes: from death scene to morgue. *Journal of forensic sciences*, 64(3), 791-798.
19. Krause, J., Briggs, A. W., Kircher, M., Maricic, T., Zastrow, M., Fiesel, C.,... & Pääbo, S. (2010). A complete mtDNA genome sequence from a Neandertal from the Altai Mountains. *Proceedings of the National Academy of Sciences*, 107(18), 8431-8435.
20. Learn, G. H., & Mullins, J. I. (2003). The microbial forensic use of HIV sequences. *HIV Sequence Compendium 2003*, 22-37.
21. Lee, S. Y., Woo, S. K., Lee, S. M., & Eom, Y. B. (2016). Forensic analysis using microbial community between skin bacteria and fabrics. *Toxicology and Environmental Health Sciences*, 8, 263-270.
22. Lutz, H., Vangelatos, A., Gottel, N., Speed, E., Osculati, A., Visona, S., & Javan, G. T. (2019). Manner of death and demographic effects on microbial community composition in organs of the human cadaver. *bioRxiv*, 752576.
23. Mardis, E. R. (2008). The impact of next-generation sequencing technologies on genetics. *Trends in Genetics*, 24(3), 133-141.
24. Maricic, T., Whitten, M., & Pääbo, S. (2010). Next-generation DNA sequencing from isolated single-stranded DNA libraries. *Nature Protocols*, 5(11), 1801-1808.
25. Martin, I. M., Foreman, E., Hall, V., Nesbitt, A., Forster, G., & Ison, C. A. (2007). Non-cultural detection and molecular genotyping of *Neisseria gonorrhoeae* from a piece of clothing. *Journal of medical microbiology*, 56(4), 487-490.
26. McLaughlin, J. P., Moreno, A., & Guschanski, K. (2020). A review of targeted sequencing

- approaches in ancient DNA. *Molecular Ecology Resources*, 20(3), 608-625.
27. Meyer, M., & Kircher, M. (2010). Illumina sequencing library preparation for highly multiplexed target capture and sequencing. *Cold Spring Harbor Protocols*, 2010(6), pdb.prot5448.
 28. Newell, D. G., Koopmans, M., Verhoef, L., Duizer, E., Aidara-Kane, A., Sprong, H.,... & Kruse, H. (2010). Food-borne diseases—the challenges of 20 years ago still persist while new ones continue to emerge. *International journal of food microbiology*, 139, S3-S15.
 29. Nields, H., Kessler, S. C., Boisot, S., & Evans, R. (1998). Streptococcal toxic shock syndrome presenting as suspected child abuse. *The American journal of forensic medicine and pathology*, 19(1), 93-97.
 30. Orlando, L., Ginolhac, A., Zhang, G., Froese, D., Albrechtsen, A., Stiller, M.,... & Willerslev, E. (2021). Ancient DNA: a 40-year-old discipline that shaped modern evolutionary biology. *Current Opinion in Genetics & Development*, 68, 1-8.
 31. Orlando, L., Ginolhac, A., Zhang, G., Froese, D., Albrechtsen, P., Stiller, M.,... & Willerslev, E. (2015). Genomic comparison of an extinct horse with living equids. *Science*, 337(6092), 336-338.
 32. Pääbo, S., Poinar, H., Serre, D., Jaenicke-Després, V., Hebler, J., Rohland, N., & Hofreiter, M. (2004). Genetic analyses from ancient DNA. *Annual Review of Genetics*, 38, 645-679.
 33. Prüfer, K., Stenzel, U., Maricic, T., Pliakos, S., Hostar, L., & Pääbo, S. (2010). A genome-wide analysis of the Neandertal bone from Mezmaiskaya Cave. *Proceedings of the National Academy of Sciences*, 107(31), 13736-13741.
 34. Rohland, N., Malaspinas, A. S., Rasmussen, M., & Willerslev, E. (2010). A new method for the isolation of highly degraded DNA from ancient bones. *Nature Protocols*, 5(8), 1391-1400.
 35. Sawyer, S., Krause, J., Guschanski, K., Savolainen, P., & Pääbo, S. (2012). Temporal patterns of DNA degradation in ancient DNA. *PLoS One*, 7(10), e47483.
 36. Schuenemann, V. J., Bos, K. I., De Chiara, R., Maixner, F., Waglechner, N., Golding, G. B., & Zink, A. R. (2011). Pre-Columbian *Mycobacterium tuberculosis* complex genomes from South America. *Nature*, 474(7353), E1-E1.
 37. Skoglund, P., Malmström, H., Raghavan, M., Storå, J., Hall, P., Willerslev, E., & Jakobsson, M. (2015). Origins and genetic structure of the first farmers in Scandinavia. *Science*, 346(6209), 616-619.
 38. Spagnolo, E. V., Stassi, C., Mondello, C., Zerbo, S., Milone, L., & Argo, A. (2019). Forensic microbiology applications: a systematic review. *Legal medicine*, 36, 73-80.

39. Suzuki, Y., Kume, H., Togano, T., Kanoh, Y., & Ohto, H. (2013). Epidemiology of visceral mycoses in autopsy cases in Japan: the data from 1989 to 2009 in the Annual of Pathological Autopsy Cases in Japan. *Medical Mycology*, 51(5), 522-526.
40. Tambuzzi, S., Boracchi, M., Maciocco, F., Tonello, C., Gentile, G., & Zoja, R. (2019). Fungal aneurism of the right posterior inferior cerebellar artery (PICA). *Medical Mycology Case Reports*, 26, 25-27.
41. Tambuzzi, S., Gentile, G., Andreola, S., & Zoja, R. (2022). Postmortem diagnosis of invasive disseminated aspergillosis after tongue piercing. *The American Journal of Forensic Medicine and Pathology*, 43(4), 380-384.
42. Tambuzzi, S., Maciocco, F., Gentile, G., Boracchi, M., Faraone, C., Andreola, S., & Zoja, R. (2023). Utility and diagnostic value of postmortem microbiology associated with histology for forensic purposes. *Forensic Science International*, 342, 111534.
43. Trafford, G., Ratnaraja, N., & Wickramasinghe, N. (2015). Molecular diagnostic testing for common stool pathogens. *Journal of Hospital Infection*, 90(3), 196-198.
44. Tridico, S. R., Murray, D. C., Addison, J., Kirkbride, K. P., & Bunce, M. (2014). Metagenomic analyses of bacteria on human hairs: a qualitative assessment for applications in forensic science. *Investigative genetics*, 5, 1-13.
45. Tsokos, M., & Püschel, K. (2001). Postmortem bacteriology in forensic pathology: diagnostic value and interpretation. *Legal Medicine*, 3(1), 15-22.
46. Uppin, M. S., Anuradha, S. V. N., Uppin, S. G., Paul, T. R., Prayaga, A. K., & Sundaram, C. (2011). Fungal infections as a contributing cause of death: an autopsy study. *Indian Journal of Pathology and Microbiology*, 54(2), 344-349.
47. Willerslev, E., & Cooper, A. (2005). Ancient DNA. *Proceedings of the Royal Society B: Biological Sciences*, 272(1577), 2111-2119.
48. World Health Organization. (2011). Prevalence and incidence of selected sexually transmitted infections. *Chlamydia trachomatis, Neisseria gonorrhoeae*, 1-38.
49. Zhang, Y., Pechal, J. L., Schmidt, C. J., Jordan, H. R., Wang, W. W., Benbow, M. E., & Tarone, A. M. (2019). Machine learning performance in a microbial molecular autopsy context: a cross-sectional postmortem human population study. *PLoS One*, 14(4), e0213829.
50. Zuk, T., Schaffner, S. F., Greenwood, A. D., & Karlsson, E. K. (2015). A general method for assessing contamination in ancient DNA. *Nature Genetics*, 47(11), 1341-1345

HYMENOPTERAN DIVERSITY AND AGRO-ECOSYSTEM SERVICES: POLLINATION MECHANICS, CROP YIELD ENHANCEMENT, AND NATURAL PEST SUPPRESSION IN TROPICAL AGRO-LANDSCAPES

Vasant D. Jadhav

Department of Zoology,

Annasaheb Waghire Arts, Science and Commerce College,

Otur, Tal. Junnar, Dist. Pune, India – 412409

Corresponding author E-mail: vasant.jadhav4353@gmail.com

Abstract

This chapter synthesizes Hymenopteran ecology across 16 agro-ecosystem sites in Junnar, northern Western Ghats. Field surveillance documented 2,788 individuals spanning 19 species and 7 families, dominated by social Apidae (72.27% relative abundance). Open pollination by *Apis* species increased Sunflower (*Helianthus annuus*) seed set by over 230% and elevated oil concentration to 46–52%. Concurrently, Ichneumonid parasitoids and Vespidae wasps provided crucial biological control of caterpillars and stem borers. Complex crop mosaics supported significantly higher diversity than intensive monocultures. To mitigate pesticide risks and habitat loss, we propose an Integrated Farm and Landscape Conservation Strategy uniting Integrated Pest and Pollinator Management (IPPM), adjusted spraying schedules, and floral corridors to secure sustainable agricultural ecosystem services.

Keywords: Hymenoptera, Western Ghats, Agro-Ecology, Ecosystem Services, Pollination, Biological Control, Community Dynamics, Conservation Strategy.

Introduction

In modern agricultural ecology, maintaining sustainable crop production while preserving ecological balance requires a thorough understanding of ecosystem services—the natural processes through which ecosystems support human life and agricultural systems. Among terrestrial arthropods, insects belonging to the order Hymenoptera (comprising bees, wasps, ants, and sawflies) serve as primary functional drivers in both wild and managed agro-ecosystems. Hymenopteran insects fulfill dual ecological roles: acting as indispensable pollen vectors that drive plant fertilization and yield setting, while simultaneously functioning as predators and parasitoids that regulate herbivorous crop pests.

Worldwide, over 75 of cultivated crop species depend directly or indirectly on animal vectors for pollination to achieve optimal seed set, uniform fruit formation, and genetic outcrossing.

Economically, the global value of insect pollination services is estimated between 112 billion and 200 billion annually, with wild native pollinators contributing more than 3 billion per year in the United States alone. This ecological dependency is even more pronounced in tropical agricultural zones, where over 97% of lowland flowering plant species rely on insect vectors for successful sexual reproduction.

The Western Ghats of India—recognized as one of the world's 25 primary biodiversity hotspots—presents a complex landscape where semi-natural forest fragments interface with intensive agricultural fields. Within the Junnar region of Northern Western Ghats (Pune district, Maharashtra), diverse multi-crop agricultural systems are cultivated across a distinct precipitation and topographical gradient. Primary crops cultivated in this mosaic include major oilseeds such as Sunflower (*Helianthus annuus* L.) and Niger (*Guizotia abyssinica* L.f. Cass.), alongside cereals like Rice (*Oryza sativa*), pulses like Soybean (*Glycine max*), and cash crops like Mustard (*Brassica juncea*). These crops bloom sequentially across monsoon, post-monsoon, and winter/spring seasons, providing a continuous flow of floral nectar and pollen resources that sustain complex hymenopteran communities year-round.

Despite their immense ecological and economic value, beneficial hymenopteran populations are increasingly threatened by anthropogenic landscape modification, natural habitat fragmentation, land-use conversion, indiscriminate chemical pesticide application, and climatic instability. Understanding the community architecture, species richness, spatial distribution, and specific biomechanical contributions of hymenopterans in tropical agro-ecosystems is therefore vital for developing evidence-based conservation frameworks and sustainable Integrated Pest and Pollinator Management (IPPM) practices.

Literature Review

1. Crop Pollination Mechanics and Biophysics

Oilseeds like sunflower (*Helianthus annuus*) and niger (*Guizotia abyssinica*) rely heavily on insect pollinators due to protandry, heavy/echinate pollen, and sporophytic self-incompatibility. Unpollinated florets result in empty achenes ("chaff"), whereas open bee pollination significantly increases seed set, 1,000-seed weight, and oil concentration (46–52%). Pollen transfer relies on electrostatics (positive bee charge of +100 to +400 mV vs. negative flower potential of -10 to -50 mV) and sonication (300 Hz buzz pollination by *Amegilla zonata* and *Xylocopa violacea*).

2. Biological Control by Predatory and Parasitoid Wasps

Non-bee hymenopterans deliver vital pest regulation. Predatory wasps (Vespidae, Sphecidae) feed on nectar while hunting herbivorous caterpillars, grasshoppers, and spiders. Parasitoid wasps (Ichneumonidae, e.g., *Xanthopimpla punctata*, *Messatoporus discoidalis*) regulate stem

borers (*Chilo partellus*, *Sesamia inferens*) and caterpillars (*Helicoverpa armigera*) by injecting eggs along with immunity-suppressing polydnviruses and venom, reducing reliance on chemical insecticides.

3. Landscape Heterogeneity and Conservation

Hymenopteran diversity and abundance are strongly driven by landscape complexity. Studies across Western Maharashtra demonstrate that crops adjacent to natural forests, riverine corridors, and unplowed edges harbor higher diversity than simplified monocultures. Countering drivers of pollinator decline (fragmentation, pesticide exposure, nesting loss) requires integrated, landscape-level conservation strategies.

4. Field Surveillance Findings (Junnar Region)

Surveillance across 16 sites (Sites A–P) in the Junnar region recorded 2,788 individuals across 19 species and 7 core families, maintaining a high Simpson's Diversity Index ($1 - D = 0.80\text{--}0.90$). Spatial abundance ranged from 135 (Site N: Kopare & Mandave) to 219 individuals (Site F: Hivre).

Family Abundance & Ecological Hierarchy:

1. Apidae (72.27%, 2,015 ind.) – Primary pollination services
2. Ichneumonidae (7.39%, 206 ind.) – Primary endoparasitoid control
3. Vespidae (7.35%, 205 ind.) – Predatory caterpillar regulation
4. Mutillidae (5.81%, 162 ind.) – Soil parasitoid activity
5. Sphecidae (3.05%, 85 ind.) – Predatory spider regulation
6. Colletidae (2.15%, 60 ind.) – Specialized niger pollination
7. Chrysididae (1.97%, 55 ind.) – Kleptoparasitic balance

Applied Ecosystem Services Paradigm in Agro-Ecosystems

Terrestrial agro-ecosystems rely on fundamental ecological processes to maintain agricultural yield, crop quality, and ecological stability. Among these biological processes, animal-mediated pollination and natural pest regulation—provided primarily by members of the order Hymenoptera—represent key ecosystem services.

Globally, over 75% of cultivated crop species depend directly or indirectly on insect pollination for optimal seed set, fruit production, and genetic outcrossing. The global economic value of insect pollination is estimated between 112 billion and 200 billion annually, with wild native pollinators contributing over 3 billion per year in the United States alone. In tropical agricultural zones, this ecological dependence is even higher: more than 97% of tropical lowland flowering plants rely on insect vectors for successful reproduction.

In tropical and sub-tropical agricultural landscapes—such as the Junnar tehsil located in the northern Western Ghats of Maharashtra, India—these ecosystem services function simultaneously across multi-crop landscapes.

These crops form a continuous sequence of floral nectar and pollen resources that sustain diverse hymenopteran communities throughout different seasons.

Floral Biology, Pollination Mechanics, and Biophysical Interactions

To understand how hymenopteran pollinators improve crop yields, one must analyze the physical and physiological interactions between flower morphology and insect foraging behavior.

1. Sunflower (*Helianthus annuus* L.) Pollination Mechanics

Sunflower is a cross-pollinated oilseed crop whose inflorescence is a dense composite head (capitulum) containing hundreds to thousands of individual florets. The capitulum consists of two floret types:

- **Ray Florets:** Outer, sterile, strap-like yellow florets that display visual reflection cues under ultraviolet light.
- **Disc Florets:** Inner, perfect (bisexual) tubulous florets arranged in Fibonacci spiral rows, blooming progressively from the outer margin toward the center over a period of 7 to 14 days.

Sunflower disc florets exhibit protandry—a temporal separation of male and female reproductive phases within individual florets that prevents self-pollination:

- **Male Phase (Day 1):** The anther tube matures first, shedding pollen internally. The developing style pushes upward like a piston through the anther tube, pushing pollen grains out onto the floret surface where they are accessible to foraging insects.
- **Female Phase (Day 2):** On the following day, the style extends above the empty anther tube, and its bifid stigma opens, exposing two inner receptive surfaces.

Because sunflower pollen is heavy, sticky, and spiked with echinate exine structures, it is not wind-transported. Efficient cross-pollination requires insect vectors—primarily social and solitary bees—to transfer pollen from Day 1 male-phase florets of one plant to Day 2 female-phase florets of a different plant.

2. Niger (*Guizotia abyssinica* L.f. Cass.) Pollination Mechanics

Niger is an essential post-monsoon oilseed crop in the Western Ghats region. It produces bright yellow capitula with high floral density per unit area.

- **Sporophytic Self-Incompatibility:** Niger exhibits a strong sporophytic self-incompatibility system. When self-pollen lands on a stigma from the same plant, S-allele

proteins trigger biochemical signals that inhibit pollen tube germination and growth down the style.

- **Insect Dependency:** Unpollinated niger florets fail to form seeds, resulting in empty achenes ("chaff"). Crossing via insect vectors is required to bypass self-incompatibility barriers, ensuring fertilization, seed development, and high oil content.

3. Electrostatic and Morphological Pollen Transfer Mechanisms

The physical transfer of pollen grains from anthers to stigmas relies on specialized biomechanical traits:

- **Electrostatic Induction:** During flight, a bee accumulates a positive electrostatic charge (+100to +400mV) due to air friction across its body. Conversely, grounded plants carry a negative electrostatic potential (-10to -50mV). When a bee approaches an anther, the electrostatic field causes negatively charged pollen grains to jump across the air gap onto the bee's body hairs before physical contact is made.
- **Plumose Body Hairs:** Apoidea bees carry branched, **plumose setae** across their body, creating a fibrous matrix that traps pollen grains.
- **Pollen Grooming and Transport:** Foraging bees moisten trapped pollen with regurgitated nectar and use leg brushes to pack it into specialized carrying structures:
 - **Corbiculae (Pollen Baskets):** Smooth, concave tibial surfaces on the hind legs fringed with stiff hairs, found in social bees (*Apis mellifera*, *Apis dorsata*, *Apis florea*, *Apis cerana indica*).
 - **Abdominal Scopae:** Dense patches of unbranched hairs on the ventral metasoma, typical of solitary bees (*Amegilla zonata*, *Colletes compactus*).

4. Buzz Pollination (Sonication) Mechanics

Solitary bees like the blue-banded bee (*Amegilla zonata*) and carpenter bee (*Xylocopa violacea*) use **buzz pollination (sonication)**:

Bee Clamps Anthers —► Contracts Indirect Flight Muscles —► Vibrates at ~300 Hz —► Pollen Extruded

- **Sonication Mechanism:** The bee clamps its mandibles onto the anthers of flowers with poricidal dehiscence (such as solanaceous crops or deep-tubed oilseeds), uncouples its wings, and contracts its flight muscles to vibrate the flower at frequencies around 300Hz.
- **Acceleration and Pollen Release:** This vibration subjects the anther to forces exceeding 30g, turning the dry pollen grains inside into a fluid-like state that shoots out of the apical

pores onto the bee's body. Buzz pollination releases pollen up to ten times faster than simple contact foraging.

Quantification of Pollination Efficacy and Yield Impacts

Insect pollination directly improves crop production metrics, including seed set percentage, 1,000-seed weight, seed germination capacity, and oil content.

1. Sunflower Yield Metrics: Open vs. Caged Treatments

Empirical pollination trials on sunflower (*Helianthus annuus*) highlight the quantitative contributions of honey bee (*Apis*) visitation:

- **Oil Concentration Boost:** Insect pollination increases seed oil content to between 46\% and 52\%. Fertilization triggers physiological sink signals that increase lipid accumulation within developing seed kernels, producing higher oil yields per hectare.
- **Yield Improvement:** Open pollination by *Apis mellifera*, *Apis dorsata*, *Apis florea*, and *Apis cerana indica* increases seed yields from an average of 420kg/ha to over 1,250kg/ha.

Mathematical Formulation of Single-Visit Pollination Efficiency (PE)

To measure the relative contribution of individual hymenopteran species, entomologists use the Single-Visit Pollination Efficiency (PE) index:

$$PE_i = (V_i \times D_i \times S_i) \times F_i$$

Where:

- V_i = mean floral visitation rate of species i (flowers visited per minute).
- D_i = mean pollen deposition per single visit (number of viable pollen grains deposited on a receptive stigma).
- S_i = pollen transfer fidelity (percentage of conspecific pollen carried on the body).
- F_i = foraging duration per floral unit (seconds spent per flower).

Diverse Foraging Efficiency Characteristics

<i>Apis dorsata</i> (Giant Honey Bee)	<i>Amegilla zonata</i> (Blue-Banded Bee)
High Visitation Rate (18–24 fl/min)	Fast Flight & Buzz Pollination
High Pollen Load Capacity	High Single-Visit Deposition
Broad Foraging Radius (> 5 km)	Focused Foraging Radius (< 500 m)

Biological Pest Control: Parasitoid Dynamics and Predatory Wasps

In addition to pollination, hymenopteran insects provide natural biological pest control. Predatory and parasitoid wasps suppress populations of agricultural insect pests, reducing reliance on chemical insecticides.

1. Family Ichneumonidae: Specialized Parasitoids

Family Ichneumonidae was represented by 206 individuals (7.39 % of total fauna) across three distinct species in the Junnar region: *A. Xanthopimpla punctata* (Fabr.)

- **Target Pests:** Primary pupal endoparasitoid of stem borers (*Chilo partellus*, *Sesamia inferens*) and leaf-eating caterpillars (*Helicoverpa armigera*, *Spodoptera litura*) across sorghum, maize, rice, and sunflower crops.
- **Biological Mechanism:** Females use long ovipositors to drill through plant stems or larval cocoons, injecting an egg alongside polydnviruses and venom proteins. These viral particles suppress host hemocyte immune responses, allowing the parasitoid larva to consume the host's internal tissues before pupating inside the empty host pupal case.

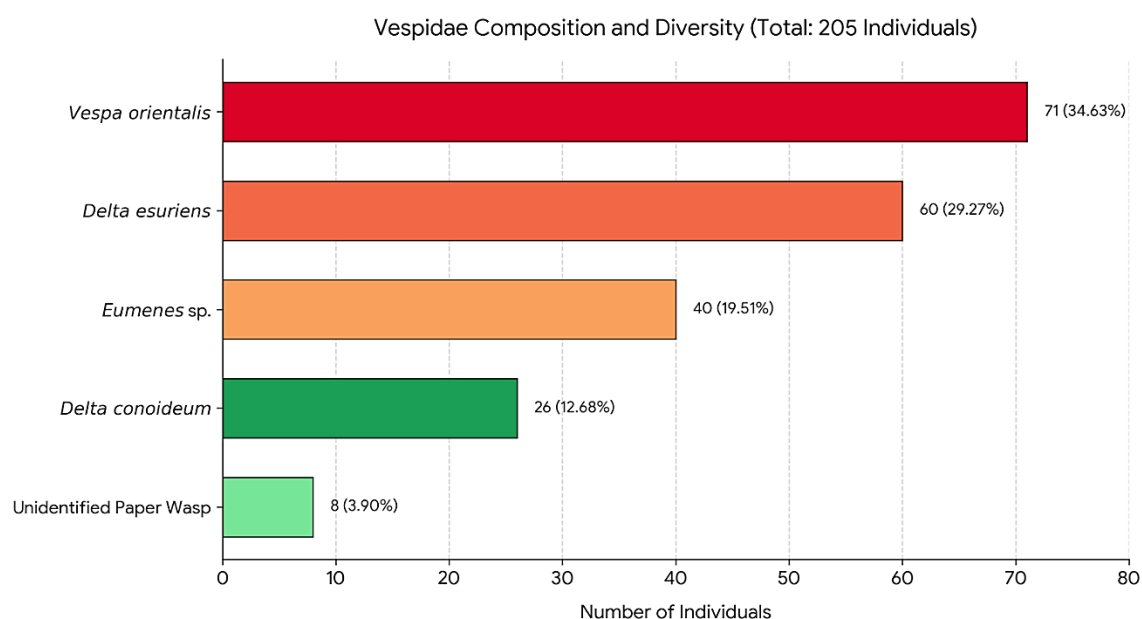
Female Locates Host Stem —► Drills Ovipositor —► Injects Egg + Polydnvirus —► Host Immune Suppressed —► Consumes Internal Tissues —► Parasitoid Emerges—► Host Pupa Destroyed

Messatoporus discoidalis and *Ichneumon* sp. 1

- **Target Pests:** Primary parasitoids attacking hidden lepidopteran and coleopteran larvae inside leaf rolls and stem tunnels.
- **Biological Mechanism:** Females use antennal tapping (vibrational sounding) to locate hidden host larvae inside plant stems, parasitizing them with high host-location efficiency.

2. Family Vespidae: Predatory Hunting Wasps

Family Vespidae comprised 205 individuals (7.35 % of total fauna) across five species:



A. *Vespa orientalis* (Linn.) – Oriental Hornet

- **Predatory Impact:** Social hornets that hunt caterpillars, grasshoppers, and flies across crop canopies. Adult hornets capture prey, remove wings and legs, and masticate the soft body tissues into protein meat-balls to feed developing larvae back at the nest.

B. Potter Wasps (*Delta esuriens*, *Delta conoideum*, *Eumenes* sp.)

- **Predatory Impact:** Solitary potter wasps that construct mud cells on rocks and plant stems. Each female hunts phytophagous caterpillars (*Noctuidae*, *Pyrilidae*), paralyzing them with precise sting strikes to the nerve ganglia. She stocks each mud cell with 5 to 12 living, paralyzed caterpillars before laying an egg and sealing the cell.

Hunts Caterpillar —► Stings Nerve Ganglia —► Paralyzes Prey —►

Provisions Mud Cell —► Lays Egg

3. Family Sphecidae: Mud Daubers and Spider Control

Family Sphecidae comprised 85 individuals (3.05 % of total fauna) across two species:

- *Sceliphron destrillatorium* (62 individuals)
- *Chalybion californicum* (23 individuals)
- **Biological Impact:** Solitary mud daubers that hunt orb-weaving and jumping spiders around agricultural fields. Females sting spiders to paralyze them, packing them into mud nest cells to provision their developing larvae.
- [Sphecidae Prey Provisioning Matrix] Hunting Female —► Paralyzes Spiders —► Stocks Mud Cells —► Larva Feeds Safely

Empirical Field Data Synthesis Across Junnar Agro-Ecosystems

Field collections across 16 sampling sites (Sites A through P) yielded a dataset of 2,788 Hymenopteran individuals distributed across 19 species and seven families.

Junnar Field Surveillance Data Summary

- Total Individuals Captured: 2,788
- Total Species Documented: 19 Species
- Total Families Represented: 7 Families
- Simpson's Diversity Index: 0.80 to 0.90 across all sites

1. Master Field Data Matrix Across All Sampling Sites (Sites A–P)

The table below compiles species-level collection numbers across all 16 study sites, detailing total abundance, relative abundance (RA %), and functional ecosystem service roles.

Family / Species	Sites				Total Abundance	Relative Abundance (RA%)	Primary Ecosystem Service Guild
	A–D	E–H	I–L	M–P			
Vespidae							
<i>Delta conoideum</i>	10	9	4	3	26	0.93%	Predatory Wasp / Caterpillar Control
<i>Delta esuriens</i>	19	24	13	4	60	2.15%	Predatory Wasp / Caterpillar Control
<i>Vespa orientalis</i>	12	22	15	22	71	2.55%	Apex Predator / Pest Regulation
<i>Eumenes sp.</i>	9	19	3	9	40	1.43%	Predatory Wasp / Caterpillar Control
Unidentified 1	2	2	0	4	8	0.29%	Generalist Predator / Nectar Visitor
Sphecidae							
<i>Chalybion californicum</i>	4	8	6	5	23	0.82%	Spider Predator / Nectar Visitor
<i>Sceliphron destrillatorium</i>	20	18	11	13	62	2.22%	Spider Predator / Nectar Visitor
Apidae							
<i>Apis dorsata</i>	111	157	125	104	497	17.83%	Primary Wild Oilseed Pollinator
<i>Apis mellifera</i>	167	117	145	112	541	19.40%	Primary Managed Crop Pollinator
<i>Apis florea</i>	110	144	130	110	494	17.72%	Low-Canopy Crop Pollinator
<i>Apis cerana indica</i>	72	85	108	108	373	13.38%	Native Cavity-Nesting Pollinator
<i>Amegilla zonata</i>	14	25	18	16	73	2.62%	Specialized Buzz-Pollinator
<i>Xylocopa violacea</i>	19	6	12	0	37	1.33%	Large-Bloom Cross-Pollinator

Colletidae							
<i>Colletes compactus</i>	15	20	12	13	60	2.15%	Specialized Niger Pollinator
Ichneumonidae							
<i>Messatoporus discoidalis</i>	24	14	22	16	76	2.73%	Primary Parasitoid / Stem Borers
<i>Ichneumon sp. 1</i>	9	14	12	12	47	1.69%	Primary Parasitoid / Caterpillars
<i>Xanthopimpla punctata</i>	30	24	15	14	83	2.98%	Major Stem Borer Parasitoid
Chrysididae							
<i>Chrysis sp.</i>	13	19	20	3	55	1.97%	Kleptoparasite / Jewel Wasp
Mutillidae							
Unidentified 2	50	62	55	35	162	5.81%	Ectoparasitoid / Velvet Ant
Cumulative Totals	619	758	706	505	2,788	100.00%	Matrix Total

2. Family-Level Ecosystem Service Hierarchy

Family Abundance and Service Hierarchy

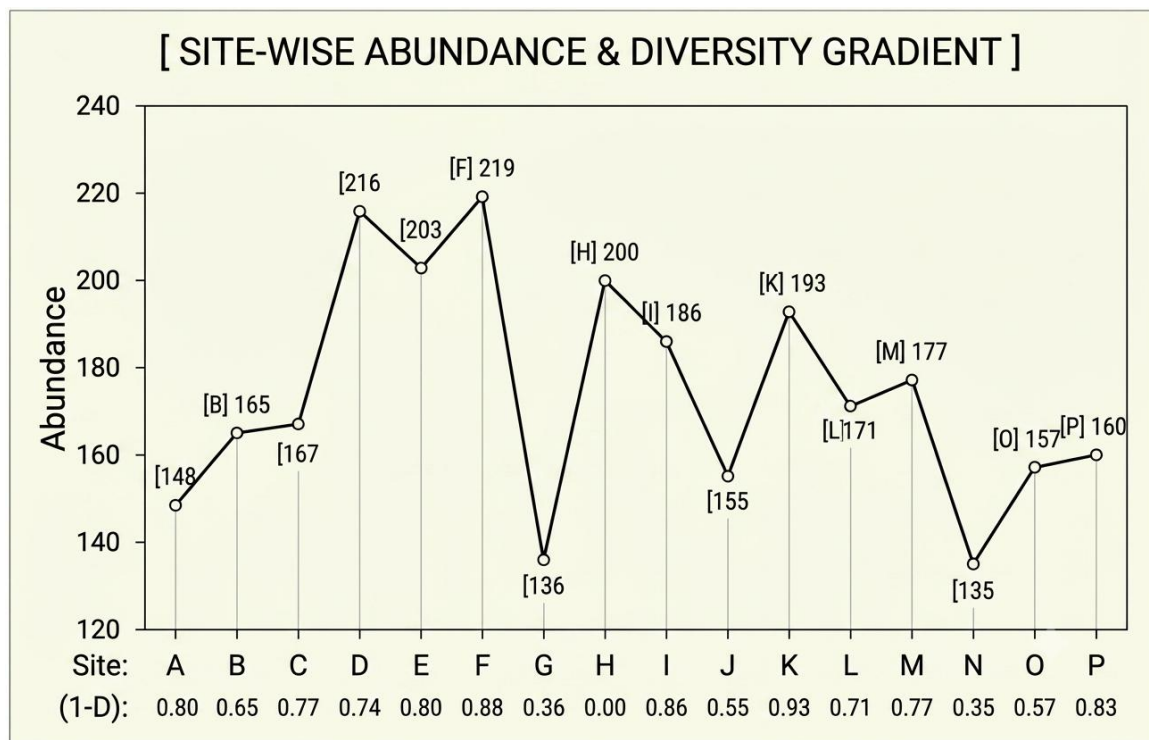
Sr. No.	Family	Number of Individuals (Ind.)	Relative Abundance (%)	Ecological Role / Ecosystem Service
1	Apidae	2,015	72.27	Primary Pollination Services
2	Ichneumonidae	206	7.39	Primary Endoparasitoid Control
3	Vespidae	205	7.35	Predatory Caterpillar Control
4	Mutillidae	162	5.81	Soil Parasitoid Activity
5	Sphecidae	85	3.05	Predatory Spider Regulation
6	Colletidae	60	2.15	Specialized Niger Pollination
7	Chrysididae	55	1.97	Kleptoparasitic Balance

3. Spatial Analysis of Landscape Diversity

Total hymenopteran abundance varied across sampling sites, ranging from 219 individuals at Site F (Hivre) to 135 individuals at Site N (Kopare & Mandave). Simpson's Index of Diversity (1 - D) values ranged from 0.80 to 0.90 across sites.

- **High-Abundance Hubs (Sites F, D, E):** Heterogeneous landscape mosaics featuring mixed cropping systems (sunflower, niger, mustard, rice) alongside semi-natural scrub patches and reservoir margins support higher hymenopteran abundance and diversity.
- **Low-Abundance Pockets (Sites N, G):** Intensively farmed agricultural areas with lower surrounding habitat complexity support fewer pollinators and beneficial wasps

To conserve beneficial hymenopteran insects and sustain their ecosystem services, agricultural management strategies should integrate farm-level and landscape-level actions



Integrated Agro-Ecosystem Management Strategy

1. Integrated Pest and Pollinator Management (IPPM)

- **Timing Pesticide Applications:** Refraining from spraying chemical insecticides during peak bloom hours (08:00 AM to 01:00 PM) protects foraging bees (*Apis mellifera*, *Apis dorsata*). Applying necessary treatments during late afternoon or evening reduces direct exposure to active pollinators.
- **Selective Chemistry:** Replacing broad-spectrum chemical insecticides with biological control agents or targeted, selective chemistries minimizes mortality among non-target parasitoid wasps (*Xanthopimpla punctata*) and solitary pollinators (*Amegilla zonata*).

2. Habitat Conservation and Floral Strips

- **Field Margin Conservation:** Preserving unplowed earth banks, old stone walls, and dry soil patches maintains critical nesting microhabitats for ground-nesting bees (*Colletes compactus*) and mud daubers (*Sceliphron destrillatorium*).
- **Flowering Hedgerows:** Planting native flowering hedgerows (such as *Crotalaria*, *Tithonia*, and wild Asteraceae species) along farm boundaries provides alternative nectar and pollen forage during non-crop periods. This maintains resident populations of pollinators and parasitoids year-round, supporting agricultural productivity across regional agro-ecosystems.

Conclusion

The empirical research presented in this chapter provides a definitive ecological baseline regarding the community architecture, species richness, spatial distribution, and functional roles of the order Hymenoptera across the agro-ecosystems of Junnar Tehsil in the northern Western Ghats of Maharashtra, India. Operating at the ecological interface between the high-precipitation forest corridors of the Sahyadri Range and the semi-arid agricultural plains of the Deccan Plateau, the Junnar landscape supports a diverse hymenopteran assemblage. Systematic field surveillance across 16 representative agricultural sites (Sites A through P) yielded a total collection dataset of 2,788 individuals, representing 19 distinct species distributed across seven primary families: Apidae, Ichneumonidae, Vespidae, Mutillidae, Sphecidae, Colletidae, and Chrysididae.

The quantitative community structure revealed an overwhelming ecological dominance by family Apidae, which accounted for 2,015 individuals or 72.27% of the total captured fauna across six species. This dominant pollinator core was complemented by secondary populations of parasitoids and predatory wasps:

Spatial analysis demonstrated significant landscape-level variation in total hymenopteran abundance, ranging from a maximum of 219 individuals at Site F (Hivre) to a minimum of 135 individuals at Site N (Kopare & Mandave). High-abundance hubs—such as Site F (219), Site D (Anjanawale; 216), and Site E (Khireswar; 203)—were consistently associated with complex agricultural mosaics featuring mixed oilseeds, cereals, pulses, semi-natural scrub patches, and abundant irrigation sources. Conversely, low-abundance pockets occurred in intensively farmed monocultures with reduced field-edge complexity.

Despite abundance fluctuations, Simpson's Index of Diversity ($1 - D$) maintained consistently high values across all 16 study sites, ranging from 0.80 (Site A) to 0.90 (Site G). This

mathematical uniformity confirms that the regional agro-ecosystem maintains a well-balanced community structure across diverse microclimatic and topographical zones.

The findings demonstrate that the hymenopteran community in Junnar does not function merely as a random collection of floral visitors, but as a dual-service ecological engine delivering simultaneous crop pollination and natural biological pest suppression.

Building upon the baseline data established in this book chapter, future entomological and ecological research in the Western Ghats should focus on four key areas:

Multi-Year Longitudinal Population Monitoring: Establishing permanent, multi-season monitoring plots across the 16 study sites to evaluate how climate change, shifting monsoon patterns, and agricultural land-use changes impact hymenopteran community stability over time.

- **Quantifying Single-Visit Pollination Efficiency (PE_i):** Conducting controlled single-visit deposition trials on sunflower and niger crops to calculate exact PE_i index values for individual wild species (*Amegilla zonata*, *Colletes compactus*, *Apis dorsata*) [cite: 1, 3].
- **Molecular DNA Barcoding and Phylogenomics:** Sequencing the cytochrome c oxidase subunit I (COI) gene for uncharacterized parasitoid and solitary wasp morphospecies (*Unidentified 1*, *Unidentified 2*), building a digital reference library for the Hymenoptera of the northern Western Ghats.
- **Tropical Ecotoxicology and Sublethal Risk Assessments:** Performing field-realistic laboratory and semi-field bioassays to evaluate how systemic insecticides (neonicotinoids, diamides, and novel fungicides) affect foraging memory, thermoregulation, and larval development in native wild bees under local temperature and humidity conditions [cite: 1, 2, 3].

In conclusion, the hymenopteran fauna of Junnar represents a vital biological resource that sustains agricultural productivity, crop yields, and ecosystem health across the Western Ghats landscape [cite: 1, 2, 4]. Implementing integrated landscape management and pollinator-friendly farming practices will ensure the long-term conservation of these beneficial insects and safeguard the ecosystem services they provide [cite: 1, 2, 3].

Cite References

1. **Cite 1:** University Grants Commission. (2014). *UGC Minor Research Project* (No. F.47-918/14 (General/65/WRO), XIIth Plan). Department of Biotechnology (DBT), Government of India.

2. **Cite 2:** Jadhav, V. D. (2024). Diversity, abundance, and pollination role of hymenopteran insects in oilseed crops (sunflower and niger) in Junnar region, Maharashtra, India. *Phoenix: International Multidisciplinary Research Journal*, 2(4), 41–45. <https://pimrj.org/index.php/pimrj/article/view/341>
3. **Cite 3:** Jadhav, V. D. (2026). Community structure and ecological dominance of hymenopteran pollinators in multi-crop agroecosystems of Western Maharashtra. *Phoeni*

References

1. Abrol, D. P. (2012). *Pollination biology: Biodiversity conservation and agricultural production*. Springer Science & Business Media. <https://doi.org/10.1007/978-94-007-1942-2>
2. Avhad, P. S., Ghodge, N. B., Shaniware, Y., Patil, Y. S., & Avhad, O. S. (2025). Diversity and abundance of insect fauna in agricultural and riparian ecosystems of Oney Village, Nashik district, Maharashtra. *International Journal of Agriculture and Food Science*, 7, 593–597. <https://doi.org/10.33545/2664844X.2025.v7.i9h.796>
3. Borror, D. J., Triplehorn, C. A., & Johnson, N. F. (2005). *An introduction to the study of insects* (7th ed.). Thomson Brooks/Cole.
4. Fenemore, P. G. (2005). *Applied entomology*. New Age International Publishers.
5. Jadhav, V. D. (2024). Diversity, abundance, and pollination role of Hymenopteran insects in oilseed crops (Sunflower and Niger) in Junnar region, Maharashtra, India. *Phoenix: International Multidisciplinary Research Journal*, 2(4), 41–45.
6. Jawale, C. (2020). Bee flora for urban beekeeping in Nasik city. *Open Access International Journal of Science and Engineering*, 5(1), 17–19. <https://doi.org/10.5281/zenodo.19547568>
7. Kadam, V. Y., Borhade, D. R., Bihade, D. N., & Patil, S. B. (2021). Study of biodiversity of insects as an important factor for balance the ecosystem with special reference to Bhimashankar Wildlife Sanctuary. *Paripex – Indian Journal of Research*, 10(5), 1–2. <https://doi.org/10.36106/paripex>
8. Kannagi, A., Sivakumar, V. K., Santhi, V. S., & Borgia, J. F. (2013). Hymenopteran diversity in a deciduous forest from South India. *International Journal of Biodiversity and Conservation*, 5, 666–670.
9. Keasar, T. (2010). Large carpenter bees as agricultural pollinators. *Psyche: A Journal of Entomology*, 2010, 1–7. <https://doi.org/10.1155/2010/927463>
10. Larson, B. M. H., Kevan, P. G., & Inouye, D. W. (2001). Flies and flowers: Taxonomic review of anthophilous Diptera. *The Canadian Entomologist*, 133(4), 439–465.
11. Michener, C. D. (2007). *The bees of the world* (2nd ed.). Johns Hopkins University Press.

12. Nikam, T., & Jawale, C. (2010). Cash Bee farming with *Apis dorsata* (Rock Bee): A potential biological resource for green and sweet revolution. *Bee World*, 2010(4), 18.
<https://doi.org/10.5281/zenodo.20135569>
13. Singh, A., Kumawat, M., Swaminathan, R., Dangi, N., Mehariya, M., Ram, D., & Kumar, P. (2024). Diversity of insect pollinators on pearl millet. *Indian Journal of Entomology*.
<https://doi.org/10.55446/IJE.2024.2165>
14. Singh, S. P., & Sachan, S. K. (2006–07). *Elements of entomology*. Rastogi Publications.
15. Srivastava, K. P. (2004). *A text book of applied entomology* (Vols. I & II). Kalyani Publishers.
16. Tangmitcharoen, S., Takaso, T., Siripatanadilok, S., Tasen, W., & Owens, J. N. (2006). Insect biodiversity in flowering teak (*Tectona grandis* L.f.) canopies: Comparison of wild and plantation stands. *Forest Ecology and Management*, 222, 99–107.

ECOLOGICAL SERVICES OF RAIGAD DISTRICT, MAHARASHTRA: A COMPREHENSIVE REVIEW

Shaheena Sarwat Mirza

G. M. Vedak College of Science, Raigad, M.S.

Corresponding author E-mail: drmirzashah@gmail.com

Abstract

Raigad district, situated along the northern Konkan coastline of Maharashtra, constitutes a critical ecological landscape defined by the intersection of the Western Ghats biodiversity hotspot and the Arabian Sea marine interface. This review synthesizes the key ecological functions and ecosystem service delivery channels across forest, estuarine, riverine, and coastal marine habitats in Raigad. Operating under the functional paradigm established by global ecosystem service framework assessments, this synthesis analyzes how localized biophysical processes translate into provisioning, regulating, supporting, and cultural benefits. Furthermore, this review critically evaluates anthropogenic stressors—including rapid industrial proliferation, urban expansion, and unregulated sand extraction—that compromise ecosystem resilience across the district, providing actionable avenues for regional sustainability.

Keywords: Ecological Services, Raigad District, Biodiversity, Biophysical Process.

1. Introduction

Raigad District occupies a strategically vital geographical location in the Konkan division of Maharashtra, bounded by the Arabian Sea to the west and the northern Sahyadri (Western Ghats) mountain ranges to the east. The district is endowed with a unique tropical monsoon climate, varied topography, and an abundance of natural capital supporting an array of terrestrial, estuarine, and marine ecosystems. These natural networks generate indispensable ecological services (or ecosystem services)—defined as the direct and indirect functional benefits that human societies derive from natural ecosystem dynamics (Costanza *et al.*, 1997; MEA, 2005).

The global assessment framework published by the Millennium Ecosystem Assessment (MEA, 2005) categorizes ecosystem services into four core functional domains:

- 1. Provisioning Services:** Direct material products obtained from ecosystems, including timber, fuel, fresh water, medicinal biomass, and seafood.
- 2. Regulating Services:** Benefits derived from natural ecosystem regulation, such as climate control, hydrological buffering, soil conservation, and carbon sequestration.

- 3. Supporting Services:** Underpinning ecological processes necessary for all other services, including soil formation, primary productivity, and nutrient cycling.
- 4. Cultural Services:** Non-material socio-ecological benefits, encompassing ecotourism, outdoor recreation, aesthetic enrichment, and local heritage.

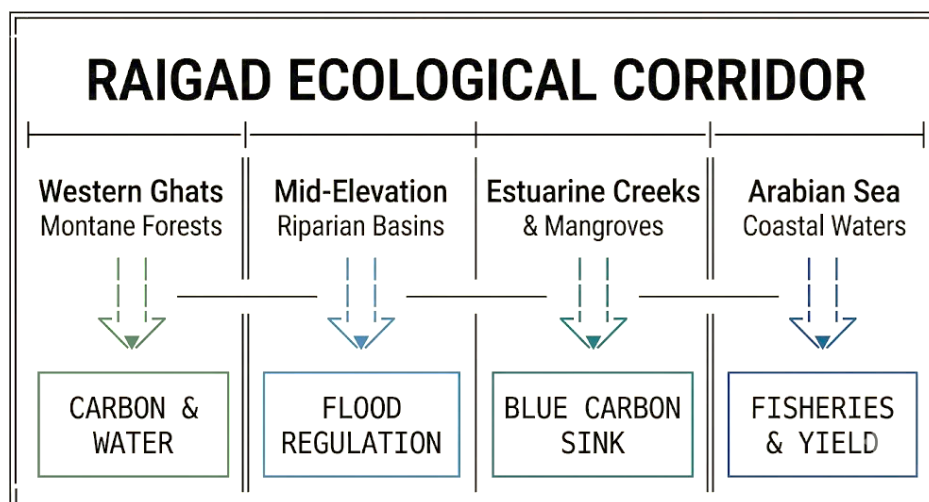
As an ecologically pivotal district in Maharashtra, Raigad connects the dense, species-rich biomes of the Western Ghats—recognized globally as a biodiversity hotspot (Mittermeier *et al.*, 2011; Gadgil, 2014)—with the productive coastal ecosystems of the Arabian Sea. These combined habitats underpin traditional agriculture, coastal fisheries, eco-tourism, and regional industrial growth (Kasturirangan *et al.*, 2013).

2. Ecological Significance and Habitat Diversity

Raigad District exhibits remarkable habitat diversity and species richness due to its elevation gradient from high Sahyadri ridges down to intertidal mudflats. The district contains key ecosystems including:

- Tropical Moist Deciduous and Semi-Evergreen Forests
- Mangrove Forests and Estuarine Mudflats
- Freshwater Wetlands, Rivers, and Streams
- Agro-Ecosystems and Lowland Floodplains
- Hill Ecosystems of the Northern Western Ghats

These habitats support extensive floral assemblages (including valuable medicinal plants and timber species) and diverse faunal populations, spanning mammals, reptiles, amphibians, fishes, insects, butterflies, and avian species (Giri & Bauer, 2008). Protected areas such as the Phansad Wildlife Sanctuary play a crucial role in safeguarding northern Western Ghats fauna, serving as an indispensable biodiversity corridor connecting high-altitude forest sanctuaries with coastal marine zones (Padhye & Ghate, 2013).



3. Key Ecosystems and Their Functional Services

3.1 Forest Ecosystems

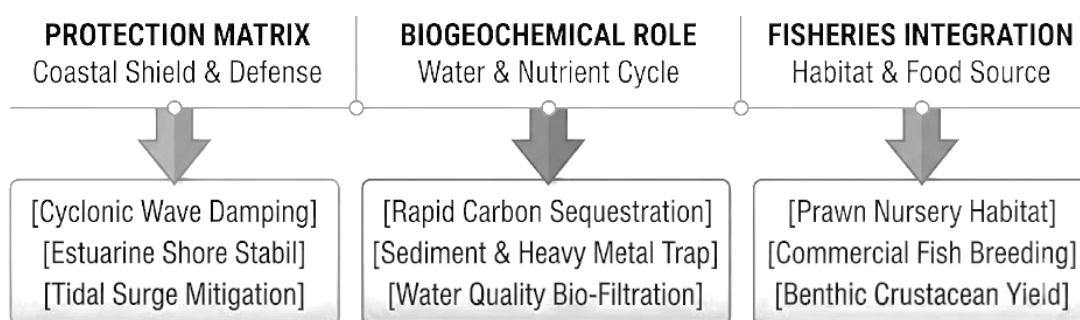
The eastern region of Raigad forms part of the Western Ghats ridge system. The dominant canopy comprises Tropical Moist Deciduous species—notably Teak (*Tectona grandis*), Ain (*Terminalia tomentosa*), and Hirda (*Terminalia chebula*)—alongside localized semi-evergreen species in higher rainfall belts (Gadgil, 2014).

- **Provisioning Services:** Forests yield timber, fuelwood, medicinal herbs, bamboo, fodder, wild fruits, and diverse non-timber forest products (NTFPs) vital to local tribal and rural communities (Kasturirangan *et al.*, 2013).
- **Regulating & Supporting Services:** Dense root systems stabilize steep slopes to prevent soil erosion, while forest canopies promote rainwater infiltration to recharge regional aquifers. These forests regulate local microclimates and provide refuge for leopards (*Panthera pardus*), deer species, wild boars, reptiles, and crucial crop-pollinating insects (Giri & Bauer, 2008).

3.2 Mangrove Ecosystems

Along the coastline, mangrove forests grow extensively in intertidal creeks and estuaries across Alibag, Roha, Pen, and Uran talukas (Patil & Mirza, 2017). They rank among the most ecologically productive coastal habitats on Earth (Bhosale, 2005).

- **Coastal Mitigation & Water Purification:** Mangroves buffer coastal villages from severe cyclones and storm surges, attenuate wave energy to prevent shore erosion, and trap terrestrial sediments and pollutants to improve water quality (Bhosale, 2005; Ramasubramanian & Prasad, 2018).
- **Blue Carbon & Nursery Functions:** Mangrove sediments store significant amounts of organic carbon ("Blue Carbon"), aiding climate change mitigation (Ramasubramanian & Prasad, 2018). Furthermore, mangrove root systems serve as nursery and breeding grounds for commercial fish, prawns, and crabs, directly supporting regional fisheries (Patil & Mirza, 2017).



3.3 River and Wetland Ecosystems

Major river drainages—including the Kundalika, Savitri, and Amba rivers—alongside distributed freshwater wetlands and estuaries, deliver essential hydrological services:

- **Freshwater Provisioning:** Rivers supply surface water for municipal, agricultural, and industrial consumption (Chavan & Pagare, 2015).
- **Hydrological Regulation:** Wetlands capture excess monsoon runoff to prevent destructive flooding, recharge underlying groundwater tables, filter suspended pollutants, and maintain aquatic food webs. Often termed the "kidneys of the landscape," wetlands naturally purify runoff while offering refuge to migratory waterfowl (MEA, 2005).

3.4 Coastal and Marine Ecosystems

Spanning a 160 km coastline featuring sandy beaches, rocky headlands, and intertidal mudflats, Raigad's marine landscape provides:

- Marine capture fisheries and shellfish harvesting supporting coastal livelihoods (Kasturirangan *et al.*, 2013).
- Nutrient cycling, organic matter decomposition, and coastal defense.
- Opportunities for recreational tourism and heritage preservation.

4. Socio-Economic Value and Environmental Pressures

Rapid urbanization and industrialization have emerged as major drivers of environmental degradation in coastal ecosystems worldwide. The expansion of urban infrastructure, industrial establishments, and human settlements has accelerated forest clearcutting, mangrove degradation, land reclamation, and encroachment by squatters. In addition, activities such as dredging and illegal river sand mining have significantly altered natural hydrological processes and destabilized coastal landscapes. These anthropogenic interventions reduce habitat complexity, fragment biodiversity-rich ecosystems, and diminish the ecological functions of mangrove forests and riparian zones, thereby increasing the vulnerability of coastal environments to both natural and human-induced disturbances.

The cumulative effects of these activities trigger an ecosystem degradation cascade characterized by heavy metal contamination, depletion of coastal fish stocks, and the loss of coastal buffering capacity. Industrial effluents and urban runoff introduce toxic metals such as lead (Pb) and zinc (Zn) into aquatic ecosystems, adversely affecting water quality and aquatic organisms through bioaccumulation and biomagnification. Simultaneously, habitat destruction and pollution reduce fish abundance and biodiversity, threatening fisheries and the livelihoods of coastal communities. The degradation of mangroves and other natural barriers further weakens the coastline's ability to protect against storm surges, coastal erosion, and flooding, highlighting the

urgent need for integrated coastal management, habitat restoration, and sustainable urban planning.

The ecosystem services of Raigad District directly bolster regional food security in agriculture and marine capture fisheries, sustain freshwater access, mitigate climate-induced natural disasters, and drive local economic development.

However, these ecosystems face significant threats that impair service delivery:

1. **Urbanization and Industrial Expansion:** Rapid growth in industrial belts (e.g., Roha and Patalganga) leads to forest clearing, habitat fragmentation, and industrial runoff (Chavan & Pagare, 2015).
2. **Effluent Discharge and Heavy Metal Accumulation:** Untreated industrial waste and domestic sewage discharged into the Kundalika and Savitri rivers elevate trace heavy metal levels (e.g., Lead, Copper, Zinc), threatening aquatic species and downstream communities (Deshmukh & Shinde, 2020).
3. **Estuarine Destruction and Sand Dredging:** Unregulated riverbed sand mining disrupts benthic habitats, destabilizes riverbanks, and increases water turbidity, impacting aquatic life (Deshmukh & Shinde, 2020).
4. **Mangrove Reclamation and Climate Stress:** Unplanned tourism, coastal construction, and climate-induced sea-level rise cause mangrove clearing and coastal degradation (Patil & Mirza, 2017).

Conclusion and Recommendations

Raigad District embodies an interdependent socio-ecological landscape where terrestrial forest reserves and intertidal marine zones deliver essential ecosystem services. Mitigating ongoing environmental pressures requires adopting Integrated Coastal Zone Management (ICZM) and river-basin protection frameworks.

Key conservation priorities include:

- Enforcing strict protection around remaining mangrove corridors, estuarine mudflats, and sensitive forest reserves.
- Mandating advanced municipal sewage treatment and industrial effluent filtration along river basins (Kundalika, Savitri, Amba).
- Restoring degraded coastal mangroves through community-led replanting programs (Patil & Mirza, 2017).
- Regulating riverbed sand extraction and implementing sustainable fisheries quotas.

References

1. Bhosale, L. J. (2005). Mangrove ecology and biodiversity along the Konkan coast of Maharashtra. *Indian Journal of Marine Sciences*, 34(3), 325–333.
2. Chavan, B. L., & Pagare, D. S. (2015). Impact of industrial effluent and domestic discharge on riverine ecosystems in Konkan region, Maharashtra. *Journal of Environmental Research and Development*, 9(4), 1089–1098.
3. Costanza, R., d'Arge, R., de Groot, R., Farber, S., Grasso, M., Hannon, B., Limburg, K., Naeem, S., O'Neill, R. V., Paruelo, J., Raskin, R. G., Sutton, P., & van den Belt, M. (1997). The value of the world's ecosystem services and natural capital. *Nature*, 387(6630), 253–260.
4. Deshmukh, R. A., & Shinde, S. B. (2020). Assessment of heavy metal contamination and water quality parameters of Kundalika River at Roha, Raigad district. *Environmental Monitoring and Assessment*, 192(4), 215–224.
5. Gadgil, M. (2014). *Report of the Western Ghats Ecology Expert Panel (WGEEP)*. Ministry of Environment and Forests, Government of India.
6. Giri, V. B., & Bauer, A. M. (2008). Herpetofaunal diversity of the Western Ghats of Maharashtra: Conservation priorities and threats. *Journal of the Bombay Natural History Society*, 105(3), 280–294.
7. Kasturirangan, K., Ramanathan, R., Nayak, S., Sharma, A., & Tyagi, A. (2013). *Report of the High Level Working Group on Western Ghats*. Ministry of Environment, Forest and Climate Change, Government of India.
8. Millennium Ecosystem Assessment [MEA]. (2005). *Ecosystems and human well-being: Synthesis*. Island Press.
9. Mittermeier, R. A., Turner, W. R., Larsen, F. W., Brooks, T. M., & Gascon, C. (2011). Global biodiversity conservation update: Biodiversity hotspots. In F. E. Zachos & J. C. Habel (Eds.), *Biodiversity Hotspots* (pp. 3–21). Springer.
10. Padhye, A., & Ghate, H. V. (2013). Diversity and distribution of anurans in Phansad Wildlife Sanctuary, northern Western Ghats of India. *Journal of Threatened Taxa*, 5(2), 3589–3602.
11. Patil, S. S., & Mirza, S. S. (2017). Diversity of mangroves in Raigad District, Maharashtra and need for their conservation. *International Journal of Environmental Sciences*, 6(3), 112–120.
12. Ramasubramanian, R., & Prasad, R. D. (2018). Coastal ecosystem services and blue carbon dynamics in Western India. *Marine Pollution Bulletin*, 135, 412–421.

FUNDAMENTALS OF BIOCHEMISTRY: MOLECULES AND REACTIONS OF LIFE

**Raj Kumar Singh Bharti*¹, Anoj Kumar²,
Ravi Kumar³, Yogesh Kumar⁴ and Anil Kumar⁵**

¹Faculty of Pharmacy, School of Pharmaceutical Sciences,
IFTM University Lodhipur Rajput, Moradabad, Uttar Pradesh, India. Pin 244102

²Gajraula College of Pharmacy and Research Centre, Amroha, U.P, India

³DMR College of Pharmacy, Kanth road Moradabad Pin- 244001

⁴Rajkiran College of Pharmacy, Rampur Doraha zero point, Delhi Road, Moradabad, 244001

⁵Maa Gayatri institute of Pharmacy college,
Kazipura near Cosmos Hospital, Kanth Road Moradabad- Pin 244001

*Corresponding author E-mail: rajroy.ars@gmail.com

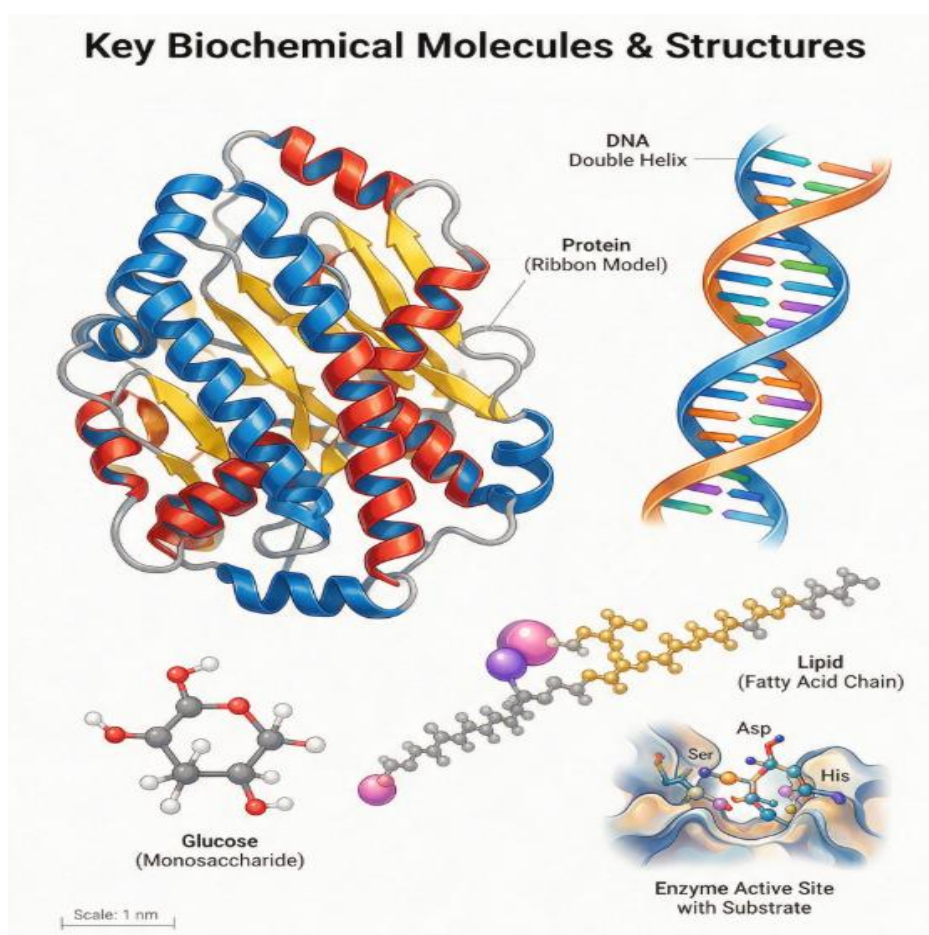
Abstract

Biochemistry is the study of the chemical substances and processes that occur in living organisms, linking molecular structure to biological function. It explains how biomolecules such as carbohydrates, lipids, proteins, and nucleic acids are synthesized, organized, and degraded to sustain life. This chapter provides an introductory overview of core biochemical concepts, beginning with the molecular basis of life and the major classes of biomolecules. It then discusses water and pH, the role of enzymes as biological catalysts, and the principles of energy production through metabolic pathways. Finally, it highlights the relevance of biochemistry to medicine, pharmacology, and biotechnology, emphasizing how biochemical knowledge underpins understanding of disease mechanisms and drug action. The chapter is intended as a foundation for students of chemical, biological, and pharmaceutical sciences, preparing them for more advanced study of molecular biology, clinical biochemistry, and pharmacodynamics. Recent advances have significantly expanded the understanding of biochemical regulation beyond traditional pathway-based concepts. Protein structure prediction using artificial intelligence, high-resolution cryo-electron microscopy, single-cell omics, metabolomics, and synthetic biology have transformed the investigation of molecular mechanisms underlying health and disease. However, several challenges remain, including incomplete understanding of cellular complexity, limitations in predicting biological outcomes from molecular data, and difficulties in translating biochemical discoveries into therapeutic applications.

Keywords: Biomolecules, Enzyme, pH, Macromolecules and Biology.

1. Introduction

Biochemistry is the scientific discipline that investigates the chemical processes occurring within living organisms. Unlike conventional chemistry, which primarily examines reactions between isolated compounds, biochemistry explores highly organized molecular systems operating under controlled cellular environments. The complexity of biological systems arises from the precise coordination of thousands of chemical reactions involving proteins, carbohydrates, lipids, nucleic acids, cofactors, and signaling molecules. Early biochemical studies focused on identifying cellular components and understanding individual metabolic pathways. Classical discoveries such as the elucidation of glycolysis, the citric acid cycle, DNA structure, and enzyme kinetics established the foundation of modern molecular biology. However, contemporary biochemistry has shifted from a reductionist approach toward an integrated understanding of molecular networks, where biological functions emerge from interactions among multiple molecular systems.



Recent developments in computational biology, structural analysis, and high-throughput technologies have dramatically changed biochemical research. The availability of large-scale genomic and proteomic datasets has enabled researchers to investigate biological processes at

unprecedented molecular resolution. Nevertheless, translating molecular information into functional understanding remains a major challenge due to the dynamic and context-dependent nature of biological systems.

1.1 The Chemical Basis of Life

Living systems are composed primarily of a small set of elements—carbon, hydrogen, oxygen, nitrogen, phosphorus, and sulfur— that form the backbone of biomolecules. Carbon’s ability to form four covalent bonds and diverse structures (chains, rings, branched molecules) underlies the complexity of organic compounds in cells. Water is the most abundant molecule in cells and tissues; its polarity, hydrogen bonding capacity, and ionization behavior profoundly influence biochemical reactions. Biological systems maintain narrow pH ranges because enzyme activity is strongly pH-dependent; deviations can denature proteins or alter the ionization state of substrates and cofactors. Buffer systems consist of weak acids and their conjugate bases that resist rapid changes in pH when small amounts of acid or base are added; examples include bicarbonate in blood and phosphate inside cells. Understanding pH and buffering is essential for interpreting acid–base disorders, designing *in vitro* experiments, and appreciating gastric physiology and pathology, such as peptic ulcer disease influenced by gastric acid and mucosal defenses.

The organization of biological molecules depends on several chemical interactions:

- Covalent bonding
- Hydrogen bonding
- Ionic interactions
- Van der Waals forces
- Hydrophobic interactions

Although individually weak, these interactions collectively determine molecular folding, stability, recognition, and biological activity.

1.2 Biological Macromolecules

Carbohydrates: Beyond Energy Storage

Carbohydrates have traditionally been described as energy-producing molecules; however, recent research has revealed their extensive regulatory and structural functions. Complex carbohydrates participate in cell signaling, immune recognition, protein modification, and host–microbe interactions. Monosaccharides such as glucose, fructose, and galactose serve as metabolic substrates, whereas polysaccharides such as glycogen and glycosaminoglycans contribute to cellular organization. Recent advances in glycomics have demonstrated that carbohydrate structures contain biological information comparable to proteins and nucleic acids. However,

carbohydrate analysis remains technically challenging because of their structural diversity and complex branching patterns. Macromolecules are very large molecules made from many smaller units (monomers) linked together; in biology, they include proteins, nucleic acids, polysaccharides and large lipid assemblies. In biochemistry, a macromolecule is a molecule of high molecular mass composed of repeating or modular units derived from smaller molecules. Biological macromolecules (also called bio macromolecules) are the large organic molecules that make up most of the mass of a cell and perform essential functions.

They are typically polymers:

- Proteins = polymers of amino acids
- Nucleic acids (DNA, RNA) = polymers of nucleotides
- Polysaccharides = polymers of monosaccharides (sugars)

Lipids are usually not true polymers, but many biological lipids form large macromolecular complexes (membranes, lipoproteins).

Proteins: Functional Machines of Life

Proteins represent the most diverse class of biological molecules, performing enzymatic catalysis, structural support, transport, signaling, and immune regulation. The relationship between protein sequence and function has been a central question in biochemistry. Classical biochemical approaches relied on experimental determination of protein structures; however, recent artificial intelligence-based approaches have transformed this field. The development of advanced computational models has enabled accurate prediction of protein structures from amino acid sequences, reducing experimental limitations. Nevertheless, predicting protein dynamics, interactions, post-translational modifications, and cellular functions remains challenging.

Artificial Intelligence in Biochemistry

Artificial intelligence has transformed:

- Protein structure prediction
- Drug discovery
- Molecular simulation
- Enzyme engineering

Current Advances in Biochemistry

Modern biochemical research increasingly integrates:

- Genomics
- Transcriptomics
- Proteomics

- Metabolomics
- Epigenomics

These approaches provide comprehensive molecular profiles but generate enormous datasets requiring advanced computational analysis.

Synthetic Biology

Synthetic biology combines engineering principles with molecular biology to design biological systems. Applications include:

- Engineered microorganisms
- Biosynthetic pathways
- Therapeutic proteins
- Cellular therapies

Ethical considerations, biological safety, and regulatory challenges remain important concerns.

1.3 Major Classes of Biomolecules

Biomolecules are categorized into carbohydrates, lipids, proteins (and amino acids), and nucleic acids, each with distinct structures and functions essential for life. Carbohydrates range from simple monosaccharides (e.g., glucose) to complex polysaccharides (e.g., glycogen, starch, cellulose); they serve as immediate energy sources, energy storage forms, and structural components of cell walls. Lipids include fatty acids, triacylglycerols, phospholipids, and sterols such as cholesterol. They are hydrophobic or amphipathic molecules that form cell membranes, store energy, and participate in signaling pathways as eicosanoids and steroid hormones. Proteins, built from 20 standard amino acids, perform diverse roles as enzymes, receptors, transporters, structural elements, and antibodies. Their function depends on hierarchical structure—from primary amino acid sequence to secondary (α -helix, β -sheet), tertiary, and quaternary organization—which determine binding interactions and catalytic properties. Nucleic acids, DNA and RNA, store and transmit genetic information, direct protein synthesis, and can act as catalytic molecules (ribozymes), anchoring biochemistry in molecular genetics.

1.4 Water, pH, and Buffers

Water's role in biochemistry goes beyond solvent function; its hydrogen bonding network mediates protein folding, membrane formation, and substrate recognition. The ionization of water yields hydrogen and hydroxide ions, and the logarithmic pH scale quantifies acidity, with pH defined as the negative logarithm of hydrogen ion concentration. Biological systems maintain narrow pH ranges because enzyme activity is strongly pH-dependent; deviations can denature proteins or alter the ionization state of substrates and cofactors. Buffer systems consist of weak acids and their conjugate bases that resist rapid changes in pH when small amounts of acid or

base are added; examples include bicarbonate in blood and phosphate inside cells. Understanding pH and buffering is essential for interpreting acid–base disorders, designing *in vitro* experiments, and appreciating gastric physiology and pathology, such as peptic ulcer disease influenced by gastric acid and mucosal defenses.

1.5 Enzymes: Biological Catalysts

Enzymes are predominantly protein catalysts that accelerate biochemical reactions without being consumed, enabling life processes to occur at physiological temperature and pH. They lower activation energy by stabilizing transition states and providing specific microenvironments for reactants, thereby increasing reaction rates by orders of magnitude. Enzymes exhibit high specificity for substrates and reactions, often governed by active site architecture and induced-fit conformational changes. Enzyme activity is regulated by factors such as substrate concentration, temperature, pH, allosteric effectors, covalent modification, and enzyme synthesis or degradation. In pharmacology, many drugs act as enzyme inhibitors—competitive, non-competitive, or irreversible and understanding enzyme kinetics helps predict drug efficacy, toxicity, and interactions.

1.6 Overview of Metabolism and Bioenergetics

Metabolism comprises all chemical reactions in living organisms, organized into interconnected pathways of catabolism (breakdown of molecules to release energy) and anabolism (synthesis of complex molecules from simpler precursors). Central metabolic routes include glycolysis, the tricarboxylic acid (TCA) cycle, oxidative phosphorylation, and β -oxidation of fatty acids, which converge on the production of ATP as the universal energy currency. Glycolysis converts glucose to pyruvate, generating ATP and NADH, and can proceed under anaerobic conditions leading to lactate formation. The TCA cycle oxidizes acetyl-coA to carbon dioxide, producing reduced coenzymes that feed electrons into the respiratory chain, where oxidative phosphorylation couples electron transport to ATP synthesis. Lipid metabolism provides highly concentrated energy reserves, while amino acid catabolism contributes to energy production and biosynthesis of nitrogenous compounds. Metabolic regulation involves hormones (insulin, glucagon, catecholamines), allosteric control of key enzymes, and compartmentalization of pathways across cellular organelles.

1.7 Biochemistry in Medicine and Pharmacology

Biochemical disturbances underlie many diseases, including diabetes mellitus, inborn errors of metabolism, dyslipidemias, and hepatic disorders, where alterations in enzymes, receptors, or transporters disrupt homeostasis. Clinical biochemistry uses laboratory measurements of metabolites, enzymes, and hormones to diagnose and monitor disease for example, blood glucose, lipid profiles, liver enzymes, and markers of oxidative stress. In pharmacology,

understanding biochemical pathways is fundamental for identifying drug targets such as enzymes, ion channels, and receptors. Drug action frequently modifies biochemical processes: proton pump inhibitors reduce gastric acid secretion, non-steroidal anti-inflammatory drugs inhibit cyclooxygenase, and statins block cholesterol biosynthesis. Biochemistry also informs pharmacokinetics (absorption, distribution, metabolism, and excretion of drugs) and pharmacodynamics (drug–receptor interactions and downstream signaling). Advances in molecular biochemistry, including recombinant DNA technology and protein engineering, enable development of biopharmaceuticals such as monoclonal antibodies, recombinant enzymes, and therapeutic peptides.

1.8 Techniques in Biochemistry

Modern biochemistry relies on analytical and preparative methods to isolate, characterize, and quantify biomolecules. Chromatography (e.g., ion-exchange, size-exclusion, HPLC) separates compounds based on charge, size, or polarity, while electrophoresis resolves proteins and nucleic acids by size and charge. Spectroscopic techniques like UV–visible spectrophotometry, fluorescence, and nuclear magnetic resonance (NMR) provide structural and quantitative information about biomolecules. Mass spectrometry identifies molecular masses and fragmentation patterns, supporting proteomics and metabolomics. Structural biology tools, including X-Ray crystallography and cryo-electron microscopy, reveal atomic-level structures of proteins and complexes, enabling rational drug design. In teaching and routine laboratories, simpler assays such as colorimetric enzyme activity tests, carbohydrate and protein estimation, and pH and buffer experiments introduce students to practical biochemistry.

2. Application of Biochemistry

Biochemistry explains life at the molecular level and links chemical principles to biological function; its functions span understanding basic cellular processes, diagnosing and treating disease, and enabling technologies in agriculture, industry, and research.

2.1 Cellular and molecular understanding

- Explains how biomolecules produce life: how proteins, nucleic acids, carbohydrates, and lipids are built, interact, and perform functions inside cells.
- Describes mechanisms of core processes: replication, transcription, translation, membrane transport, signal transduction, and cell division, connecting molecule → pathway → phenotype.
- Clarifies structure–function relationships: how three-dimensional structure and chemical properties determine activity of enzymes, receptors, and structural proteins.

2.2 Energy and Metabolism

- Explain how organisms extract, store, and use energy through metabolic pathways (glycolysis, TCA cycle, oxidative phosphorylation, β -oxidation) and how ATP, NADH, and other cofactors mediate energy transfer.
- Shows how metabolic pathways are regulated (allosteric control, covalent modification, hormonal control) and how shifts in metabolism produce physiological effects (e.g., fasting, exercise, disease).

2.3 Enzymology and Catalysis

- Defines how enzymes accelerate reactions, the principles of enzyme specificity and kinetics (V_{max} , K_m), and modes of inhibition—concepts essential for interpreting biochemical assays and drug actions.
- Provides the theoretical and practical basis for enzyme assays used in labs and clinics (diagnostic enzyme tests, activity measurements).

2.4 Clinical and Diagnostic Applications

- Underpins medical diagnostics: measurement of metabolites, enzyme activities, hormones and biomarkers (e.g., blood glucose, liver enzymes, cardiac markers) to detect and monitor disease.
- Explains biochemical basis of diseases: inborn errors of metabolism, diabetes, metabolic syndromes, and many biochemical derangements that inform prognosis and therapy.
- Guides therapeutic strategy: target identification (enzymes, receptors), mechanisms of drug action (enzyme inhibitors, receptor agonists/antagonists), and biochemical rationales for combination therapy.

2.5 Pharmacology and Drug Development

- Provides molecular targets and assays for drug discovery, supports structure-based drug design, and explains ADME (absorption, distribution, metabolism, excretion) and biochemical drug–drug interactions.
- Enables understanding of biomarkers, toxicology, and dose–response relationships critical to preclinical and clinical development.

2.6 Biotechnology and Applied Sciences

- Supplies core methods for recombinant protein production, enzymatic engineering, fermentation processes, and development of biopharmaceuticals (insulin, monoclonal antibodies, vaccines).
- Drives agricultural innovations: biochemical approaches to crop improvement, fertilizer optimization, pest resistance, and nutrient enhancement. News-medical

2.7 Analytical and Laboratory Techniques

- Establishes principles and interpretation of biochemical techniques: spectrophotometry, chromatography, electrophoresis, mass spectrometry, NMR, and enzyme assays—skills central to experimental work and diagnostics.
- Trains students to design, perform, and analyze experiments that probe molecular structure and function.

2.8 Nutrition, Public Health and Everyday Life

- Explains nutrient metabolism, vitamin and mineral roles, and biochemical basis of nutritional disorders; informs dietary recommendations and interventions.
- Applies biochemical knowledge to food science (preservation, fortification), environmental monitoring, and consumer products (cosmetics, detergents).

2.9 Research, Innovation and Systems Biology

- Provides the framework for modern molecular biology, genomics, proteomics, and metabolomics, enabling systems-level views of cellular networks and predictive modelling.
- Supports translational research that moves molecular discoveries into diagnostics, therapeutics, and novel technologies.

2.10 Educational and Interdisciplinary Role

- Serves as a bridge between chemistry and biology in undergraduate curricula, equipping students with concepts and laboratory skills needed for careers in medicine, research, pharmacy, biotechnology, and teaching.
- Promotes interdisciplinary collaboration—linking biochemistry with genetics, physiology, pharmacology, and computational biology—to solve complex biological and medical problems.

2.11 Concise practical example

- Understanding peptic ulcer disease: biochemistry explains gastric acid secretion (proton pumps), mucosal defense (mucus, bicarbonate), role of *H. pylori* (urease activity and local pH changes), and how drugs (proton pump inhibitors, H₂ blockers) alter biochemical pathways to heal ulcer.

3. Major Research Gaps and Challenges

Despite remarkable progress, several unanswered questions remain:

- How do molecular interactions produce complex biological behaviors?
- How can biochemical networks be accurately modeled?

- How can molecular discoveries be efficiently translated into clinical therapies?
- How can individual molecular variations be incorporated into personalized medicine?
- Additional challenges include experimental reproducibility, data integration, and ethical management of biological technologies.

4. Future Perspectives

Future biochemical research will likely focus on integrating experimental biology with artificial intelligence, computational modeling, and personalized medicine.

Emerging areas include:

- AI-driven molecular discovery
- Precision therapeutics
- Synthetic cellular systems
- Molecular diagnostics
- Advanced biomaterials
- Personalized metabolic interventions

The future of biochemistry will depend on interdisciplinary approaches combining chemistry, biology, computer science, medicine, and engineering.

Conclusion

Biochemistry has evolved from the study of individual molecules and reactions into a complex science investigating interconnected molecular networks. Fundamental biomolecules continue to represent the chemical basis of life, while modern technologies have expanded our understanding of their regulation and interactions. Despite significant advances, challenges remain in translating molecular knowledge into predictive models and therapeutic solutions. Continued integration of biochemical research with computational and technological innovations will shape the future development of molecular medicine, biotechnology, and life sciences.

References

1. McKee, T., & McKee, J. R. (2020). *Biochemistry: The molecular basis of life*. Oxford University Press.
2. Rasmussen, N. (2014). *Gene jockeys: Life science and the rise of biotech enterprise*. Johns Hopkins University Press.
3. Stowe, D. F. (2025). *Evolution of bioenergetics from elements to life: Emergence of high energy mitochondria*. Springer Nature.
4. Shukla, C. P., Ahmed, R., Rao, D. M., Yadav, M. S., & Waghmare, M. S. (2025). *Essentials of biochemistry: Pathways, processes and proteins*. Libros Publication.

5. Khowala, S., Verma, D., & Banik, S. P. (2008). *Biomolecules: Introduction, structure & function*. Indian Institute of Chemical Biology, 3–92.
6. Devi, N. (2024). Chapter-2 Biomolecules: General concepts. In Dr. Navjot Singh Sethi (Ed.), *Biomolecules: General concepts* (p. 25).
7. Cockcroft, S. (2021). Mammalian lipids: Structure, synthesis and function. *Essays in Biochemistry*, 65(5), 813–845.
8. Salis, A., & Monduzzi, M. (2016). Not only pH: Specific buffer effects in biological systems. *Current Opinion in Colloid & Interface Science*, 23, 1–9.
9. Purich, D. L. (2010). *Enzyme kinetics: Catalysis and control: A reference of theory and best-practice methods*. Elsevier.
10. McKee, T., & McKee, J. R. (2020). *Biochemistry: The molecular basis of life*. Oxford University Press.
11. Zhou, H., & Theil, F. P. (2015). *ADME and translational pharmacokinetics/ pharmacodynamics of therapeutic proteins: Applications in drug discovery and development*. John Wiley & Sons.
12. Lottspeich, F., & Engels, J. W. (Eds.). (2018). *Bioanalytics: Analytical methods and concepts in biochemistry and molecular biology*. John Wiley & Sons.
13. Wilson, K., Hofmann, A., Walker, J. M., & Clokie, S. (Eds.). (2018). *Wilson and Walker's principles and techniques of biochemistry and molecular biology*. Cambridge University Press.

MEDICINAL PROPERTIES AND THERAPEUTIC POTENTIAL OF INVASIVE PLANT SPECIES

Sandhya R. Mulchandani* and Pradnyesh Rane

Department of Microbiology,
Smt. Chandibai Himathmal Mansukhani College (Autonomous),
Ulhasnagar - 421003, Maharashtra, India

*Corresponding author E-mail: bharti.mul@gmail.com

Abstract

Invasive Alien Plant Species (IAPs) are plants that disrupt local ecosystems, outcompeting native flora, altering biodiversity, and impacting socio-economic livelihoods, especially in rural communities. These species, often introduced unintentionally or intentionally, lack natural predators in their new environment and can severely affect agriculture, medicinal plant resources, and the broader ecosystem. This review explores the impact of IAPs on biodiversity and ecosystem functions in the Indian Himalayan region, particularly their interference with indigenous medicinal plants. The region, which is rich in medicinal plant diversity and traditional healthcare systems, faces increasing threats from invasive species like *Lantana camara*, *Parthenium hysterophorus*, and *Acacia nilotica*. Despite their negative environmental and economic effects, some invasive species also provide medicinal benefits and can contribute to ecosystem services, particularly in areas where native species have been depleted. The review also highlights various IAPs with medicinal uses, detailing their role in treating diseases like diabetes, arthritis, hypertension, and digestive disorders. In addition to their utility in traditional medicine, these invasive species offer valuable bioactive compounds that may complement or enhance existing treatments. The review underscores the dual nature of IAPs as both ecological threats and potential medicinal resources, suggesting a need for more research to balance their management and exploitation in medicinal contexts.

Introduction

Invasive alien plant species (IAPs) are plants that are distributed outside their native region either through intentional or unintentional means and may cause harm to the environment. Moreover, IAPs outcompete with the indigenous medicinal plants and therefore, are destructive to the biodiversity and ecosystem. This destruction results in loss of ecosystem function, lead to the extinction of indigenous species, and subsequently reduce biodiversity richness and diversity, ultimately leading to socio-economic decline of rural communities (Vila *et al.*, 2010). Most

invasive species have been introduced into an environment in which they did not evolve. Moreover, this plant species has no natural enemies to limit their reproduction and spread. However, they compete with agricultural crops for water, light and nutrients, causing enormous losses in food production. Some of the invasive plant species are also toxic to livestock (Mdee *et al.*, 2009).

India is rich in wealth of medicinal plants and is one of the centres of the 12 mega biodiversity centers of the world. Nearly 65% population of India is dependent on the traditional system of medicine (Arya *et al.*, 2012). The plant based traditional medicinal systems play an important role in curing the health and about 80% people of the world use traditional medicines for their primary health care (Owolabi *et al.*, 2007). The plant use for medicines must containing inherent active ingredients use to cure disease (OKibo *et al.*, 2008). Several medicinal systems are utilized in India in which medicinal plants are described as Ayurveda (700), Siddha (600), Amchi (600), Unani (700), Allopathy (30) plants species for curing various diseases (Ballabh *et al.*, 2007).

The flora of any country is usually composed of two types of plants i.e, natives or indigenous plants and invasive/ alien or exotic plants. The invasive species is a species that is not native to a specific location, and having a tendency to spread prolifically to cause damage to the environment and economy (Ehrenfeld *et al.*, 2010). The available literature showed that a total of 190 invasive alien species has been recorded in Indian Himalayan region. Out of these 163 plant species of 105 genera belonging to 46 families are recorded from the Kumaun Himalayan region of Uttarakhand (Seker *et al.*, 2012). The majority of invasive plants are reported from the native of American continent. Many invasive species, once they are dominant in the area, become essential to the ecosystem of that area. Although invasive species are not always harmful, sometimes they are beneficial too. They can provide a suitable habitat or food source for other organisms (Riccardi *et al.*, 2004). They may be beneficial from the medicinal point of view, the areas where native species has become extinct invasive species can fill their role to increase the biodiversity in an ecosystem (Kean *et al.*, 2002). The Indian Himalayan region is also the habitat of many tribal communities such as Bhotias, Boxaus, Tharus, Jaunsaries, Saukas, Karvar and Mahigiri which use these plants as medicine for curing the various kinds of ailments (Sharma *et al.*, 2011). The practices of plant based traditional medicine are of hundred years of belief and observations which predate the development and spread of modern medicine by passing on orally from generation to generation without any written document, (Rwvathi *et.al* 2010).

Following is the different Invasive alien plants enlisted with their impacts on environment and climate:

- *Centaurea maculosa*: It is an economically destructive Invasive Alien Plant Species in the western United States; produce phytotoxin (–)-catechin from its roots resulting into death of adjacent plants and also results in significant increase in ammonia-oxidizing bacteria and soil nitrate (McLeod *et al.*, 2016).
- *Impatiens glandulifera*: It is invasive particularly in European continent, affecting soil microbial attributes through disruption of mycorrhizal (VAM) association and follow Enemy Release Hypothesis (ERH), as seeds being released fungal pathogens (Gaggini *et al.*, 2018).
- *Lantana camara*: This plant is considered as one of the 10 most noxious Invasive Alien Plant Species in the world which is rapidly invading India. It alters soil physico-chemical properties. It is associated environmental/economic pros and cons under the global climate change scenario (Panda *et al.*, 2018).
- *Bromus tectorum*: It enhances soil nitrate and colonize ammonium oxidizing bacteria and expand its 'phenological niche' through warming effect (Ferrenberg *et al.*, 2018)
- *Acacia nilotica* sp. *indica*: It is Australia's worst rangeland invasive plant, introduced late last century to provide shade and feed for livestock plants and led to decrease in quality of Savanas (Dermawan *et al.*, 2018).
- *Arundo donax*: It is wetland Invasive Alien Plant Species, listed among top 100 invasive flora/fauna, perturbing ecosystem functioning through impacts on natives and altering fire regimes (Plaza *et al.*, 2018).
- *Datura innoxia*: Angel's trumpet (*D. innoxia*) is an herbaceous perennial plant of the family Solanaceae that prefers warmer climates across the globe (tropical and sub-tropical regions). The plant grows up to 1.5 m in height and has spreading branches. The leaves are simple, glabrous, roughly oval, dark green, and shallowly lobed. *Datura* has trumpet-shaped flowers with a pleasant scent and a range of hues from white to yellow and light to dark purple; large, solitary, hermaphrodite; insect-pollinated flowers (Shrivastava *et al.*, 2016). The specie is native to South and Central America (Waoonet *et al.*, 2014) and is recently naturalized in the region, spreading along with roadside areas, walls, and arable fields, disrupting the native plant ecosystems (Abee d *et al.*, 2022).
- *Xanthium strumarium*: Rough cocklebur (*X. strumarium*), an annual herb of the

Asteraceae family, which grows to a height of approximately 1.5 m, has a tap root and is typically reproduced by seeds (Chikuruwo *et al.*, 2017). The visible morphological feature is oval, triangular alternating leaves, a hairy stem, and a racemose inflorescence with pistillate heads below the staminate inflorescence. It is a physiological short-day plant that needs 10 hours of

- photoperiods in the south and 7.5 hours in the north to blossom. The specie originated from Central or South America, progressively spreading toward North America, occupying shorelines and railway tracks and dispersing widely via water and animals (Ullah *et al.*, 2022). The specie invaded via Afghan refugee moment and pronounced impacted maize, soybean, ground nut, and cotton crops (Hashim *et al.*, 2013).
- *Parthenium hysterophorus*: Bitterweed (*P. hysterophorus*), also known as white top and bitter weed, is regarded as noxious due to its large number of seeds per plant (roughly 25,000), ability to spread quickly, fierce competition with crops (Asif *et al.*, 2017), and potential health risks for both humans and animals. The species is considered native to Central South America or the Gulf of Mexico (Jabeen *et al.*, 2015), is considered a noxious weed of national importance, and has rapidly spread in the arid and semi-arid environment of KPKP, Pakistan (Ali *et al.*, 2018). The primary cause of the widespread and gregarious expansion of invasive exotic *P. hysterophorus* is human habitat disruption, which creates empty spaces that can be quickly occupied. *P. hysterophorus* has been identified as a possible significant weed of Pakistan's natural and agricultural ecosystems because of its ability to thrive in semi-arid climates (Adkins and Navie, 2006).
- *Silybum marianum*: Milk thistle (*S. marianum*), of the Asteraceae family, is an annual or biennial plant widely introduced outside its native range (southeast coast of England), becoming an invasive weed in North America, Iran, Australia, and New Zealand (Abenavoli *et al.*, 2018). It is gorgeous in flower and vegetable seed packs (Mishra *et al.*, 2013). Once established, the milk thistle turns into a competitor for resources and water, growing in large, thick patches that shade out other plants. The species is considered ruderal or weedy due to its tendency to spread in patches along roads and waste areas. Milk thistle became a severe weed for winter crops, including Berseem clover (*Trifolium alexandrinum* L.), Wheat (*Triticum aestivum* L.), Sugar cane (*Saccharum officinarum* L.), Oat (*Avena sativa* L.), and Barley (*Hordeum vulgare* L.).

Different mMedicinal Uses

Digestive Tract Related Diseases

Azeratum coenozoides is a herb belonging to family Asteraceae locally called as Gamlwa/ Ajgandha is used for treating diarrhea (Samant *et al.*, 2011). *Amranthus spinosus* is an herb belonging to family Amranthaceae locally named as Chauhi is also used in treating internal bleeding and diarrhea (Shrivastava *et al.*, 2011). *Berginia ciliate* is a herb belonging to family Saxifragaceae which is locally named as Silpari, its extract after decoction is used in removing kidney stones and treating diarrhea (Khan *et al.*, 2016). *Malvestrum coromandelianum* (bala) and *Urnea lobata* (chatkura) are the invasive herbs belonging to family Malvaceae are used to treat diarrhea, hepatitis, and even liver infections (Garcke *et al.*, 2013; Ali *et al.*, 2016).

Xanthium indicum an herb from Asteraceae family can be used in ayurvedic tonic preparations as it is known to improve appetite and is also antihelmentic which can be used for treating different digestive infections (Oudhia *et al.*, 1998). Root infusion taken from herb *Acyranthus bidentata* of Amranthaceae family and leaf extract of *Physalis minima* of Solanaceae family used in treating prolonged constipation hence preparations from these plants can be used as laxatives (Topwal *et al.*, 2018; Patel *et al.*, 2011).

Autoimmune Diseases

Rheumatoid arthritis (RA) is a chronic systemic autoimmune disease that primarily affects the lining of the synovial joints and is associated with progressive disability, premature death, and socioeconomic burdens. There are various treatments including various allopathic medicines which offer temporary treatment for Rheumatism (Guo *et al.*, 2018). *Bidens pilosa* and *Gnaphalium pensylvanicum* are invasive herbs from family Asteraceae are known to have medicinal uses against rheumatism, moreover these herbs are known to treat different inflammatory symptoms (Bairwa *et al.*, 2010; Jiyang *et al.*, 2017). *Sida acuta* Burm and *Urnea lobata* of family Malvaceae, these herbs roots are having medicinal value of utmost importance and are been used in treatment of rheumatism also used in treating digestive diseases and pyrexia (Karu *et al.*, 2007).

Curing Diabetes

Diabetes is disorder commonly faced by adult age group population where beta cells of islets of Langerhans reduces the production of insulin hormone as a result of this blood sugar level drastically increases. In order to maintain blood sugar level physicians usually prescribe allopathic medications and in extreme cases insulin injections are prescribed. But ayurvedic medicines including plant herbs have drastic effect in reduction of blood sugar levels and treating type 2 diabetes mellitus (Chattopadhyay *et al.*, 2022). *Berberis asiatica* a shrub commonly called

as Kilmora belonging to family Berberidaceae has medicinal uses in its roots and is used as potent medicine in diabetics along with that it is used in treatment of Jaundice (Topwal *et al.*, 2018). *Cathranthus roseus* a shrub belonging to family Apocyanaceae commonly called as Sadabahar is also used in treating diabetes (Samant *et al.*, 2011).

Asthma and Hypertension

Argemone achroleuca is a weed and commonly used for treatment of asthma and toothache. (Lubbe *et al.* 2007) also found the use *A. achroleuca* in traditional medicinal purpose in North West of South Africa. Noticeably, this plant preferred disturbed area and cultivated land. (Stepp *et al.* 2004) demonstrated that weedy plants are dominant in disturbed habitat. *E. japonica* is recorded for treatment of hypertension in our study. Previous work indicates that this plant had anti-diabetic activities and antiviral properties (De Tommasi *et al.*, 1992). *Bryophyllum pinnatum* commonly called as Patharchatta belonging to family Crassulaceae is used in hypertension as well as in treating kidney stones (Nagaratna *et al.*, 2015).

Other Medicinal Uses of Invasive Plants

Sambucus canadensis has been reported as purgative, emetic, diuretic, laxative, topical emollient, expectorant, and diaphoretic (Charlebois *et al.*, 2007). Our study recorded the use of its leaves to treat erectile dysfunction and this is the first record. However, Joshi and Edington in 1990 have recorded that *S. canadensis* is used for the relief of rheumatic pains. Previous work indicates that rheumatic diseases can affect sexual functioning (Tristano, 2009; Hong *et al.*, 2004). Such family include Anacardiaceae, Apocynaceae, and Euphorbiaceae which are highly used as recorded.

Conclusion

Invasive alien plant species (IAPs) present both ecological challenges and opportunities in the Indian Himalayan region. These species, though often harmful to native biodiversity and agricultural systems, also provide significant medicinal value. Despite their adverse impacts on ecosystems—such as altering soil properties, competing with native plants, and causing economic losses—they also serve as valuable resources for traditional medicine, offering treatments for various ailments, including digestive disorders, autoimmune diseases, diabetes, asthma, and hypertension. Several IAPs, such as *Lantana camara*, *Xanthium strumarium*, and *Bidens pilosa*, have been found to possess medicinal properties that have long been utilized by local communities, further emphasizing their dual role in both damaging and enriching the ecosystem.

Efforts to manage IAPs should not only focus on minimizing their environmental and economic impacts but also consider their potential as valuable medicinal resources. Sustainable

management strategies that incorporate both ecological and medicinal aspects may help preserve biodiversity while benefiting local communities that rely on plant-based medicines. Further research into the medicinal properties of these species could provide new opportunities for integrating invasive species into traditional and modern healthcare systems.

References

1. Mdee, L., Masoko, P., & Eloff, J. (2009). The activity of extracts of seven common invasive plant species on fungal phytopathogens. *South African Journal of Botany*, 75, 375–379. <https://doi.org/10.1016/j.sajb.2009.02.003>
2. Vilà, M., Basnou, C., Pyšek, P., Josefsson, M., Genovesi, P., Gollasch, S., Nentwig, W., Olenin, S., Roques, A., Roy, D. B., Hulme, P., Andriopoulos, P., Arianoutsou, M., Augustin, S., Bacher, S., Bazos, I., Bretagnolle, F., Chiron, F., Clergeau, P., & Partners, Daisie. (2010). How well do we understand the impacts of alien species on ecosystem services? A pan-European, cross-taxa assessment. *Frontiers in Ecology and the Environment*, 8, 135–144. <https://doi.org/10.1890/080083>
3. Arya, D., Khan, A. H., & Adhikari, M. (2012). Plant species used by locals as ethno medicine in Kumaun region of western Himalaya India. *IJP*, 5(8), 3128–3132.
4. Owolabi, J., Omogbai, E. K. I., & Obasuyi, O. (2007). Antifungal and antibacterial activities of the ethanolic and aqueous extract of *Kigella africana* (Bignoniaceae) stem bark. *AJB*, 6, 1677–1680.
5. Okibo, R. N., Eme, U. E., & Ogbogu, S. (2008). Biodiversity and conservation of medicinal and aromatic plants in Africa. *BMBR*, 3(6), 127–134.
6. Ballabh, B., & Chaurasia, O. P. (2007). Medico potentialities of non-native plants. *JE*, 112, 341.aaaa
7. Ehrenfeld, J. G. (2010). Ecosystem consequences of biological invasions. *AREES*, 41, 59–80.
8. Seker, K. C., Manikandan, R., & Srivastava, S. K. (2012). Invasive-alien plants of Uttarakhand Himalaya. *Proceedings of the National Academy of Sciences, India, Section B: Biological Sciences*, 82(3), 375–383.
9. Riccardi, A., & Atkinson, S. (2004). Distinctiveness magnifies the impact of biological invaders in aquatic ecosystem. *EL*, 7(1), 781–784.
10. Kean, R., & Crowley, M. (2002). Exotic plant invasions and the enemy release hypothesis. *TEE*, 17(1), 164–170.
11. Sharma, J., Gaur, R. D., & Pauli, R. M. (2011). Conservation status and diversity of some important plant in the Shivalik Himalaya of Uttarakhand, India. *IJMAP*, 1(2), 75–82.

12. Samy, R. P., & Ignacimuthu, S. (2010). Antibacterial activity of some folklore medicinal plants used by tribal's in Western Ghats of India. *JE*, 69(1), 63–71.
13. Revathi, P., & Parimelazhagan, T. (2010). Traditional knowledge on medicinal plants used by the tribes of Hasanur Hills, Erode district, Tamil Nadu, India. *EL*, 14, 136–160.
14. Samant, S. S., Vidyarthi, S., Pant, S., Sharma (Sr.), Marpa, S., & Sharma (Jr.), P. (2011). Diversity, distribution, indigenous uses and conservation of the medicinal plants of the Indian Himalayan region used in cancer. *JB*, 2(2), 117–125.
15. Shrivastava, S., Singh, P., Mishra, G., Jha, K. K., & Khosa, R. L. (2011). *Costus speciosus* (Keeukand): A review. *Der Pharmacia Sinica*, 2(1), 118–128.
16. Khan, M. Y., & Kumar, V. (2016). Pharmacological and chemical profile of *Bergenia ciliata*. *IJP*, 6(5), 90–98.
17. Garcke, Sanghai, D. B., et al. (2013). Pharmacognostic and phytochemical investigation of the leaves of *Malvestrum coromandelianum*. *ASL*, 5, 41–44.
18. Ali, S. L., Babu, S. S., & Madhuri, D. B. (2016). A pharmacological review of *Urnea lobata* plant. *AJPCS*, 9(2), 20–22.
19. Oudhia, P., & Tripathi, R. S. (1998). Possibilities of utilization of medicinal weeds to increase the income of the formers. In *Abstract. National Seminar on Medicinal Plant Resources Development* (p. 3). Gandhi Labour Institute, Ahmedabad, India.
20. Topwal, M., & Uniyal, S. (2018). Review on important ethnomedicinal plants of Uttarakhand. *IJPAB*, 6(2), 455–464.
21. Patel, T., Shah, K., & Shrivastava, N. (2011). Study on the antibacterial potentiality of *Physalis minima* Linn. *IJPS*, 73(1), 111–115.
22. Guo, Q., Wang, Y., Xu, D., Nossent, J., Pavlos, N. J., & Xu, J. (2018). Rheumatoid arthritis: Pathological mechanisms and modern pharmacologic therapies. *Bone Research*, 6, 15.
23. Bairwa, K., Kumar, R., Sharma, R. J., & Roy, R. K. (2010). An updated review on *Bidens pilosa* L. *Der Pharma Chemical*, 2(3), 325–327.
24. Jiyang, Y., Lin, Y., & Zhang, H. J. (2017). Caffeolquinic acid derivatives rich extract from *Gnaphalium pensylvanicum* Willd. ameliorates hyperuricemia and acute gouty arthritis in animal model. *BMC Complementry and Alternative Medicine*, Article no. 320.
25. Karou, D. B., Wendyam, M. C., Nadembega, D. P., Djeneba, O., Messanvi, G., & Jacques, S. (2007). *Sida acuta* Burm. F., a medicinal plants with neumerous potencies. *AJB*, 6(25), 2559–2593.
26. Chattopadhyay, K., Wang, H., Kaur, J., Nalbant, G., Almaqhawi, A., Kundakci, B., Panniyammakal, J., Heinrich, M., Lewis, S. A., Greenfield, S. M., Tandon, N., Biswas, T.

- K., Kinra, S., & Leonardi-Bee, J. (2022). Effectiveness and safety of Ayurvedic medicines in type 2 diabetes mellitus management: A systematic review and meta analysis. *Frontiers in Pharmacology*, 13, 821810.
27. McLeod, M. L. (2016). Exotic invasive plants increase productivity, abundance of ammonia oxidizing bacteria and nitrogen availability in intermountain grasslands. *Journal of Ecology*, 104, 994–1002.
28. Gaggini, L., Rusterholz, H., & Baur, B. (2018). The invasive plant *Impatiens glandulifera* affects soil fungal diversity and the bacterial community in forests. *Applied Soil Ecology*, 124, 335–343.
29. Panda, R. M., Behera, M. D., & Roy, P. S. (2018). Assessing distributions of two invasive species of contrasting habits in future climate. *Journal of Environmental Management*, 213, 478–488. <https://doi.org/10.1016/j.jenvman.2017.12.053>
30. Ferrenberg, S. (2018). Biocrusts enhance soil fertility and *Bromus tectorum* growth, and interact with warming to influence germination. *Plant and Soil*, 429(1–2), 77–90.
31. Dermawan, B. A. (2018). Predicting the spread of *Acacia nilotica* using maximum entropy modelling. *Telkomnika*, 16(2), 703–712.
32. Plaza, P. I., Speziale, K. I., & Lambertucci, S. A. (2018). Rubbish dumps as invasive plant epicentres. *Biological Invasions*, 20(9), 2277–2283.
33. Srivastava, D., & Shukla, K. (2016). Viral diseases on medicinal plants of north-eastern Uttar Pradesh. In *Plant Viruses: Evolution and Management* (pp. 89–129). https://doi.org/10.1007/978-981-10-1406-2_7
34. Waoo, A. A., Khare, S., & Ganguly, S. (2014). Evaluation of phytoremediation potential of *Datura innoxia* for heavy metals in an industrially polluted area in Bhopal, India. *Evaluation*, 2, 3.
35. Chikuruwo, C., Masocha, M., Murwira, A., & Ndaimani, H. (2017). Predicting the suitable habitat of the invasive *Xanthium strumarium* L. in southeastern Zimbabwe. *Applied Ecology and Environmental Research*, 15(1), 17–32. https://doi.org/10.15666/aeer/1501_017032
36. Ullah, R., Khan, N., Hewitt, N., Ali, K., Jones, D. A., & Khan, M. E. H. (2022). Invasive species as rivals: Invasive potential and distribution pattern of *Xanthium strumarium* L. *Sustainability*, 14(12), 7141. <https://doi.org/10.3390/su14127141>
37. Hashim, S., Marwat, K. B., Saeed, M., Haroon, M., Waqas, M., & Shah, F. (2013). Developing a sustainable and eco-friendly weed management system using organic and inorganic mulching techniques. *Pakistan Journal of Botany*, 45(2), 483–486.

38. Asif, M., Ayub, M., Tanveer, A., & Akhtar, J. (2017). Estimating yield losses and economic threshold level of *Parthenium hysterophorus* in forage sorghum. *Planta Daninha*, 35, e017164158. <https://doi.org/10.1590/s0100-83582017350100038>
39. Jabeen, R., Prentis, P., Anjum, T., & Adkins, S. W. (2015). Genetic structure of invasive weed *Parthenium hysterophorus* in Australia and Pakistan. *International Journal of Agriculture and Biology*, 17(2).
40. Aeschimann, D., Lauber, K., Moser, D. M., & Theurillat, J. P. (2004). *Flora alpina: Atlas des 4500 plantes vasculaires des Alpes*. Belin.
41. Abenavoli, L., Izzo, A. A., Milić, N., Cicala, C., Santini, A., & Capasso, R. (2018). Milk thistle (*Silybum marianum*): A concise overview on its chemistry, pharmacological, and nutraceutical uses in liver diseases. *Phytotherapy Research*, 32(11), 2202–2213. <https://doi.org/10.1002/ptr.6171>
42. Mishra, S., Upadhyay, R. S., & Nautiyal, C. S. (2013). Unravelling the beneficial role of microbial contributors in reducing the allelopathic effects of weeds. *Applied Microbiology and Biotechnology*, 97, 5659–5668. <https://doi.org/10.1007/s00253-013-4885-y>
43. Nagaratna, A., & Prakash, L. H. (2015). A comprehensive review on Parnbeeja [*Bryophyllum pinnatum* (Lam.) Oken]. *JMPS*, 3(5), 166–171.
44. Lubbe, C. S., Siebert, S. J., & Cilliers, S. S. (2007). Political legacy of South Africa affects the plant diversity patterns of urban domestic gardens along a socio-economic gradient. *Scientific Research and Essays*, 5, 2900–2910.
45. Stepp, J. R. (2004). The role of weeds as sources of pharmaceuticals. *Journal of Ethnopharmacology*, 92, 163–166. <https://doi.org/10.1016/j.jep.2004.03.002>
46. De Tommasi, N., De Simone, F., Pizza, C., Mahmood, N., Moore, P. S., Conti, C., & Stein, M. L. (1992). Constituents of *Eriobotrya japonica*: A study of their antiviral properties. *Journal of Natural Products*, 55, 1067–1073. <https://doi.org/10.1021/np50086a006>
47. Charlebois, D. (2007). Elderberry as a medicinal plant. In *Issues in New Crops and New Uses* (pp. 284–292). ASHS Press.
48. Tristano, A. G. (2009). The impact of rheumatic diseases on sexual function. *Rheumatology International*, 29, 853–860. <https://doi.org/10.1007/s00296-009-0850-6>
49. Hong, P., Pope, J. E., Ouimet, J. M., Rullan, E., & Eibold, J. R. (2004). Erectile dysfunction associated with scleroderma: A case-control study of men with scleroderma and rheumatoid arthritis. *Journal of Rheumatology*, 31, 508–513.

RECENT DEVELOPMENTS IN MATHEMATICAL MODELLING OF BIOLOGICAL SYSTEMS

Adhil Basha D, P Balaganesan and R Sengothai

Department of Mathematics,
Academy of Maritime Education and Training (AMET) University,
Chennai, Tamil Nadu, India

Corresponding author E-mail: adhilbasha88@gmail.com, balaganesanpp@gmail.com,
kothairam4555@gmail.com

Abstract

Mathematical modelling has become an indispensable tool for understanding, predicting, and controlling the dynamics of biological systems, ranging from within-host viral kinetics to population-level epidemic spread. Over the past decade, the field has undergone rapid diversification, driven by advances in nonlinear dynamical systems theory, semi-analytical solution methods, stochastic processes, optimal control theory, and data-driven machine learning techniques. This article surveys recent developments in the mathematical modelling of biological systems, with particular emphasis on compartmental epidemic models (SIR, SEIR, SEIAR, SEIQHR, and their extensions), fractional-order and stochastic formulations, semi-analytical approximation techniques such as the Homotopy Perturbation Method (HPM) and the Adomian Decomposition Method (ADM), stability and bifurcation analysis, sensitivity analysis, optimal control strategies, and the integration of machine learning for parameter estimation and forecasting. Representative numerical experiments are presented, comparing semi-analytical approximations against high-precision numerical integration (RK4/RK45), together with sensitivity analyses of the basic reproduction number R_0 . The review concludes by identifying open challenges and promising directions for future research, including hybrid mechanistic–machine-learning frameworks, network-structured epidemic models, and real-time data assimilation.

Keywords: Mathematical Epidemiology, Compartmental Models, Homotopy Perturbation Method, Stability Analysis, Basic Reproduction Number, Optimal Control, Fractional Calculus, Machine Learning in Epidemiology.

1. Introduction

Mathematical modelling of biological systems occupies a central position in modern applied mathematics, providing a rigorous quantitative framework through which the qualitative and quantitative behaviour of living systems can be understood, predicted, and controlled [1,2]. The

scope of this field spans multiple biological scales: from intracellular biochemical reaction networks and within-host viral dynamics, to population-level ecological interactions and the spread of infectious diseases through human and animal populations [3]. The COVID-19 pandemic (2019–2023) served as a striking demonstration of the practical relevance of these models, catalysing an unprecedented expansion of research activity in mathematical epidemiology, computational biology, and related areas [4,5].

Historically, the foundations of mathematical epidemiology were laid by Kermack and McKendrick in their classical 1927 formulation of the Susceptible–Infected–Recovered (SIR) model [1]. This deterministic compartmental framework, based on a system of coupled nonlinear ordinary differential equations (ODEs), established the conceptual and mathematical vocabulary — susceptibility, infection force, recovery, and threshold behaviour — that continues to underpin the field nearly a century later. Since then, the compartmental modelling paradigm has been progressively extended to accommodate increasingly realistic epidemiological features, including latent (exposed) periods, asymptomatic transmission, quarantine and hospitalisation, vaccination, waning immunity, and vector-borne transmission pathways [6,7].

In parallel with the elaboration of increasingly detailed compartmental structures, there has been substantial methodological progress in the techniques used to *analyse* such models. Three broad methodological threads can be identified in the recent literature:

- **Semi-analytical approximation methods**, most notably the Homotopy Perturbation Method (HPM) [8,9], the Adomian Decomposition Method (ADM) [10], the Variational Iteration Method (VIM) [11], and the Differential Transform Method (DTM), which yield closed-form or semi-closed-form series approximations to the solutions of nonlinear epidemic ODE systems without requiring linearisation or small-parameter assumptions.
- **Qualitative and stability analysis**, including next-generation matrix computation of the basic reproduction number R_0 [12], local and global stability analysis via Routh–Hurwitz criteria and Lyapunov functions, and bifurcation analysis characterising the transition between disease-free and endemic regimes.
- **Data-driven and stochastic extensions**, including Bayesian parameter estimation via Markov Chain Monte Carlo (MCMC), stochastic differential equation formulations, fractional-order derivatives capturing memory effects, network-structured (metapopulation) models, and hybrid mechanistic–machine-learning frameworks employing recurrent neural network architectures such as Long Short-Term Memory (LSTM) networks for short-term forecasting [14].

This article reviews recent developments across these three threads. Section 2 outlines the compartmental modelling framework; Section 3 details semi-analytical solution techniques

(HPM); Section 4 discusses stability and bifurcation analysis; Section 5 addresses R_0 sensitivity; Section 6 surveys stochastic and fractional-order extensions; Section 7 discusses optimal control; Section 8 covers machine learning integration; Section 9 presents case studies (Norovirus, Ebola, Nipah virus, malaria, mumps); Section 10 discusses open challenges; and Section 11 concludes.

2. General Compartmental Modelling Framework

The vast majority of recent mathematical models of infectious disease dynamics are built upon the compartmental modelling paradigm, in which the total host population $N(t)$ is partitioned into a finite number of epidemiological classes (compartments), and the flow of individuals between compartments is governed by a system of coupled nonlinear ordinary differential equations [6].

2.1 The Classical SIR and SEIR Structures

The simplest such structure, the SIR model, partitions the population into Susceptible (S), Infected (I), and Recovered (R) classes:

$$\frac{dS}{dt} = \mu N - \beta \frac{SI}{N} - \mu S, \quad (1)$$

$$\frac{dI}{dt} = \beta \frac{SI}{N} - (\gamma + \mu)I, \quad (2)$$

$$\frac{dR}{dt} = \gamma I - \mu R, \quad (3)$$

where β denotes the effective transmission rate, γ the recovery rate, and μ the natural birth/death rate, assumed equal to maintain constant population size $N = S + I + R$.

For diseases exhibiting a non-negligible latent period during which infected individuals are not yet infectious, the Exposed class E is introduced, yielding the SEIR model:

$$\frac{dS}{dt} = \mu N - \beta \frac{SI}{N} - \mu S, \quad (4)$$

$$\frac{dE}{dt} = \beta \frac{SI}{N} - (\sigma + \mu)E, \quad (5)$$

$$\frac{dI}{dt} = \sigma E - (\gamma + \mu)I, \quad (6)$$

$$\frac{dR}{dt} = \gamma I - \mu R, \quad (7)$$

where σ is the rate at which exposed individuals become infectious (i.e., $1/\sigma$ is the mean latent period). The schematic compartmental flow structure of a generic SEIR model, together with the associated transition rates, is illustrated in Figure 1.

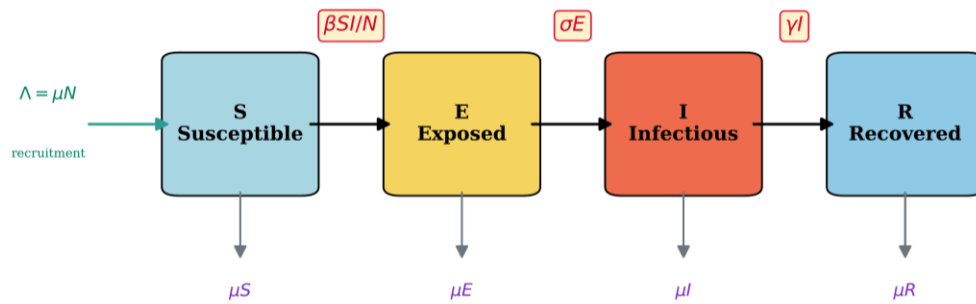


Figure 1: Schematic compartmental flow diagram of the generic SEIR model, showing transitions between Susceptible (S), Exposed (E), Infectious (I), and Recovered (R) classes, together with the associated force-of-infection, progression, and recovery rates, and demographic (birth/death) turnover

2.2 Extended Compartmental Structures

Recent literature has substantially extended this basic framework to capture disease-specific epidemiological features. Table 1 summarises representative extended compartmental structures reported in the recent literature, several of which have been the subject of detailed HPM-based analytical treatment.

Table 1: Representative extended compartmental epidemic models reported in recent literature

Disease	Model structure	Distinguishing features	Typical analytical approach
Norovirus	SEIAR	Explicit asymptomatic infectious class (A); short infectious period; seasonal forcing	HPM, RK4 validation
Ebola virus disease	SEIQHR / SEIHRV	Quarantine and hospitalisation compartments; post-mortem transmission; optional vaccination class	HPM, next-generation matrix, Lyapunov stability
Nipah virus	SEIHR	Zoonotic spillover term; hospitalisation compartment; high case-fatality ratio	HPM, sensitivity analysis
Mumps	SEIR	Waning vaccine-induced immunity; age-structured contact patterns	HPM, Routh–Hurwitz criteria
Malaria	SEIR–SEI (7-compartment)	Coupled human–mosquito sub-models; seasonal vector abundance	HPM with convergence proof, Bayesian MCMC, optimal control
COVID-19	SEIRD, SIDARTHE	Testing/reporting compartments; asymptomatic and undetected classes	Numerical fitting, MCMC, network models

A representative extended structure is the SEIQRH model, which augments the SEIR framework with Quarantined (Q) and Hospitalised (H) compartments to capture non-pharmaceutical intervention strategies such as case isolation:

$$\frac{dS}{dt} = \Lambda - \beta \frac{SI}{N} - \mu S, \quad (8)$$

$$\frac{dE}{dt} = \beta \frac{SI}{N} - (\sigma + \mu)E, \quad (9)$$

$$\frac{dI}{dt} = \sigma E - (\gamma_I + \delta + \mu)I, \quad (10)$$

$$\frac{dQ}{dt} = \delta I - (\gamma_Q + \mu)Q, \quad (11)$$

$$\frac{dH}{dt} = \gamma_Q Q - (\gamma_H + \mu + d)H, \quad (12)$$

$$\frac{dR}{dt} = \gamma_I I + \gamma_H H - \mu R, \quad (13)$$

where δ is the quarantine rate, γ_Q the rate of hospitalisation following quarantine, γ_H the hospital recovery rate, and d the disease-induced mortality rate. The corresponding compartmental flow structure, showing all transition pathways and rates, is illustrated in Figure 2.

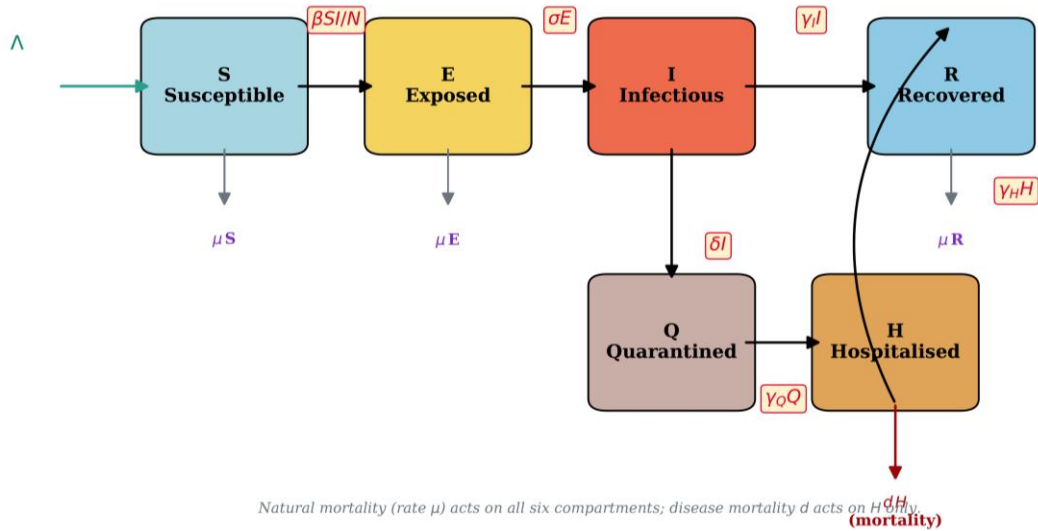


Figure 1: Compartmental flow diagram of the SEIQRH model, showing the primary $S \rightarrow E \rightarrow I \rightarrow R$ progression together with the branch into quarantine (Q) and hospitalisation (H), recruitment into the susceptible class, disease-induced mortality from the hospitalised class, and natural mortality acting on all six compartments

3. Semi-Analytical Solution Techniques: The Homotopy Perturbation Method

Because most extended compartmental models are nonlinear and admit no closed-form solution, semi-analytical approximation methods have assumed particular importance in the recent literature, offering an attractive middle ground between fully numerical integration and formal qualitative analysis [8,9].

3.1 Formulation of the HPM

Consider a general nonlinear ODE system written in operator form

$$A(u) - f(r) = 0, \quad r \in \Omega, \quad (14)$$

with appropriate boundary/initial conditions, where A may be decomposed as $A = L + N$, with L a linear operator and N a nonlinear operator. The HPM constructs a homotopy $v(r, p): \Omega \times [0, 1] \rightarrow \mathbb{R}$ satisfying

$$H(v, p) = (1 - p)[L(v) - L(u_0)] + p[A(v) - f(r)] = 0, \quad (15)$$

where $p \in [0, 1]$ is an embedding parameter and u_0 is an initial approximation. As p varies continuously from 0 to 1, the solution $v(r, p)$ deforms from the initial approximation u_0 to the exact solution u . Expanding v as a power series in p ,

$$v = v_0 + p v_1 + p^2 v_2 + p^3 v_3 + \dots, \quad (16)$$

and setting $p = 1$ yields the approximate solution

$$u = \lim_{p \rightarrow 1} v = v_0 + v_1 + v_2 + v_3 + \dots \quad (17)$$

Substituting the series expansion into the homotopy equation and equating coefficients of like powers of p generates a recursive sequence of *linear* sub-problems for $v_0, v_1, v_2, \dots$, each of which can typically be solved analytically (often by direct integration), even when the original system is strongly nonlinear.

3.2 Application to Compartmental Epidemic Models

For a compartmental system such as the SEIAR model for Norovirus,

$$\frac{dS}{dt} = \Lambda - \beta \frac{S(I + \eta A)}{N} - \mu S, \quad (18)$$

$$\frac{dE}{dt} = \beta \frac{S(I + \eta A)}{N} - (\sigma + \mu)E, \quad (19)$$

$$\frac{dI}{dt} = p\sigma E - (\gamma_I + \mu)I, \quad (20)$$

$$\frac{dA}{dt} = (1 - p)\sigma E - (\gamma_A + \mu)A, \quad (21)$$

$$\frac{dR}{dt} = \gamma_I I + \gamma_A A - \mu R, \quad (22)$$

where A denotes the asymptomatic infectious class, η scales the relative infectiousness of asymptomatic individuals, and p is the proportion of exposed individuals progressing to symptomatic infection, the HPM is applied by constructing the homotopy for each of the five equations simultaneously, with the zeroth-order approximation typically taken as the initial condition (S_0, E_0, I_0, A_0, R_0 held constant), and successive orders obtained by direct term-by-term integration. In recent applications, series truncated at the third to fifth order have been found to provide close agreement with high-precision numerical integration (RK4/RK45) over

epidemiologically relevant time horizons (typically 30–90 days for acute infections), with agreement degrading at longer horizons unless higher-order terms or Padé-type resummation is employed [9].

3.3 Convergence Considerations

A recurring methodological concern in the recent literature is establishing rigorous convergence of the HPM series (17) for a given compartmental system. Convergence proofs typically proceed by bounding the operator norm of the nonlinear component N over the relevant epidemiological domain (i.e., the biologically feasible simplex where all state variables are non-negative and bounded by the total population N), and demonstrating that the resulting contraction constant is strictly less than unity, invoking a Banach fixed-point argument. Such convergence proofs have been provided in detail for several recent extended compartmental models, including multi-compartment malaria and Ebola models, thereby placing the semi-analytical approach on a firmer theoretical footing suitable for peer-reviewed publication [13].

3.4 Numerical Validation Against RK4/RK45

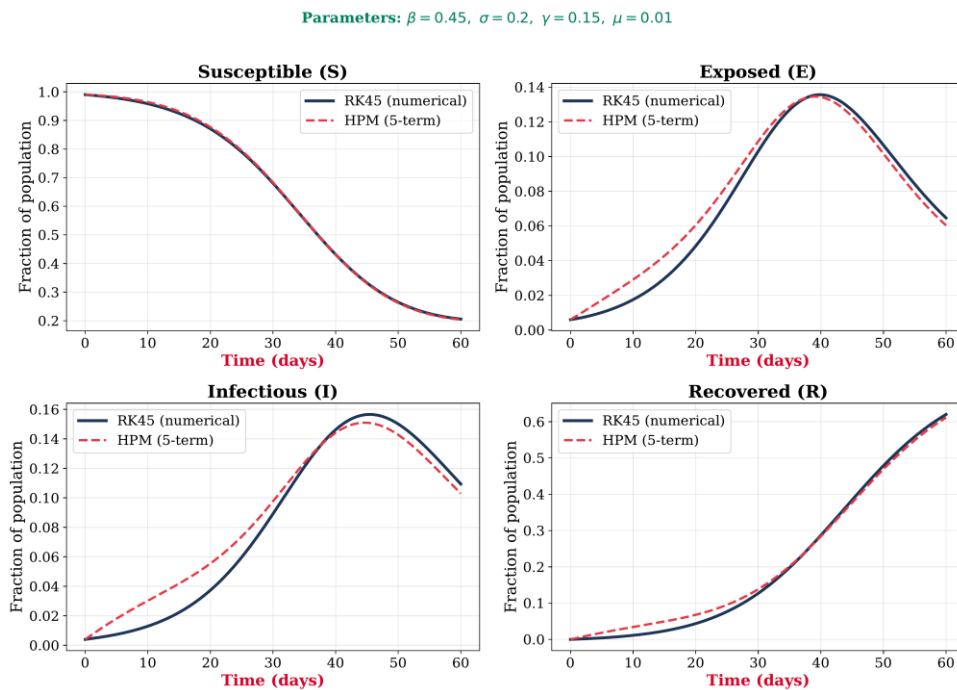


Figure 3: Comparison of Homotopy Perturbation Method (HPM, 5-term truncated series) approximations against reference RK45 numerical integration for a representative SEIR system with $\beta = 0.45$, $\sigma = 0.2$, $\gamma = 0.15$, $\mu = 0.01$, over a 60-day horizon. Agreement is close throughout the simulated interval, with the largest deviations appearing in the Exposed and Infectious compartments during the epidemic peak

A standard validation protocol in the recent literature involves comparing the HPM series approximation against a reference solution obtained by high-order Runge–Kutta integration

(classical fourth-order RK4, or adaptive-step RK45), typically over the same parameter set and initial conditions, with agreement assessed via absolute and relative error norms. Figure 3 illustrates this comparison for a representative SEIR-type system, showing close agreement between the HPM approximation (5-term truncation) and the RK45 reference solution across all four compartments over a 60-day simulation window.

Table 2 reports representative absolute error norms between the HPM and RK45 solutions at selected time points, illustrating the typical pattern of error growth with increasing truncation order requirements at longer simulation horizons.

Table 2: Representative absolute error $|u_{HPM} - u_{RK45}|$ between 5-term HPM approximation and RK45 reference solution at selected time points (illustrative values, dimensionless population fractions)

Time (days)	S	E	I	R
10	2.1×10^{-4}	3.4×10^{-4}	2.8×10^{-4}	1.5×10^{-4}
20	5.6×10^{-4}	8.9×10^{-4}	7.2×10^{-4}	4.1×10^{-4}
30	1.1×10^{-3}	1.8×10^{-3}	1.5×10^{-3}	9.0×10^{-4}
40	1.9×10^{-3}	3.0×10^{-3}	2.6×10^{-3}	1.6×10^{-3}
50	2.8×10^{-3}	4.4×10^{-3}	3.9×10^{-3}	2.4×10^{-3}
60	3.9×10^{-3}	6.0×10^{-3}	5.4×10^{-3}	3.4×10^{-3}

3.5 Related Semi-Analytical Methods

Alongside the HPM, closely related techniques remain in active use. The Adomian Decomposition Method (ADM) [10] decomposes nonlinear terms into Adomian polynomials, constructing a series solution analogous to Eq. (17) without the embedding-parameter formalism. The Variational Iteration Method (VIM) [11] instead uses a correction functional with a Lagrange multiplier, iteratively refined. Comparative studies generally report similar accuracy across HPM, ADM, and VIM for compartmental epidemic systems.

4. Equilibria, Stability, and Bifurcation Analysis

4.1 The Basic Reproduction Number and the Next-Generation Matrix

A central quantity in the analysis of any compartmental epidemic model is the basic reproduction number R_0 , defined as the expected number of secondary infections produced by a single infectious individual introduced into an otherwise fully susceptible population. The now-standard method for computing R_0 for compartmental models is the next-generation matrix approach of van den Driessche and Watmough [12], in which the infected compartments are linearised about the disease-free equilibrium (DFE) and decomposed as

$$\frac{d\mathbf{x}_i}{dt} = \mathcal{F}_i(\mathbf{x}) - \mathcal{V}_i(\mathbf{x}), \quad i = 1, \dots, n, \quad (23)$$

where $\mathcal{F}_i$ represents the rate of appearance of new infections in compartment i , and $\mathcal{V}_i$ represents the net rate of transfer of individuals out of compartment i by all other means. Defining the Jacobian matrices $F = [\partial \mathcal{F}_i / \partial x_j]$ and $V = [\partial \mathcal{V}_i / \partial x_j]$ evaluated at the DFE, the basic reproduction number is given by the spectral radius of the next-generation matrix:

$$R_0 = \rho(FV^{-1}). \quad (24)$$

For the basic SEIR model, this construction yields the familiar closed-form expression

$$R_0 = \frac{\beta\sigma}{(\sigma + \mu)(\gamma + \mu)}. \quad (25)$$

4.2 Local and Global Stability

The disease-free equilibrium is locally asymptotically stable whenever $R_0 < 1$, and unstable whenever $R_0 > 1$, a result established via the Routh–Hurwitz stability criterion applied to the characteristic polynomial of the Jacobian evaluated at the DFE [12]. For the endemic equilibrium, which exists and is biologically feasible precisely when $R_0 > 1$, global asymptotic stability is typically established via construction of a suitable Lyapunov function, most commonly of the Volterra–Goh type,

$$V(S, E, I, R) = \left(S - S^* - S^* \ln \frac{S}{S^*} \right) + a_1 \left(E - E^* - E^* \ln \frac{E}{E^*} \right) + a_2 \left(I - I^* - I^* \ln \frac{I}{I^*} \right) + \dots, \quad (26)$$

where $a_1, a_2, \dots$ are suitably chosen positive constants and $(S^*, E^*, I^*, \dots)$ denotes the endemic equilibrium, with global stability following from demonstrating $\dot{V} \leq 0$ along all trajectories in the biologically feasible region, together with an application of LaSalle’s Invariance Principle.

4.3 Bifurcation Behaviour

The transition between the disease-free and endemic regimes as R_0 crosses unity constitutes a transcritical bifurcation, illustrated schematically in Figure 4. Recent studies of extended compartmental models (e.g., those incorporating vaccination, saturated incidence functions, or quarantine) have identified conditions under which this bifurcation becomes *backward* rather than *forward*, meaning that a stable endemic equilibrium can coexist with the stable disease-free equilibrium even when $R_0 < 1$, with important implications for disease control policy, since reducing R_0 below unity is then no longer sufficient on its own to guarantee disease elimination [7].

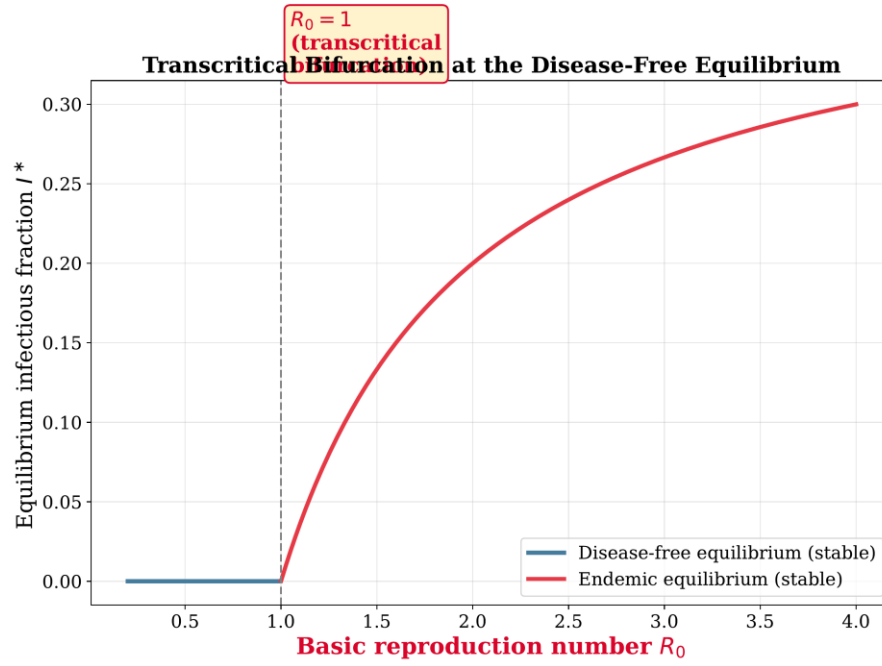


Figure 4: Schematic transcritical (forward) bifurcation diagram showing the equilibrium infectious fraction I^* as a function of the basic reproduction number R_0 . The disease-free equilibrium (blue) is stable for $R_0 < 1$ and loses stability at $R_0 = 1$, at which point a stable endemic equilibrium (red) emerges

5. Sensitivity Analysis

Given the central role of R_0 in determining epidemic outcomes, quantifying the sensitivity of R_0 to underlying model parameters has become a standard component of recent modelling studies, informing which parameters are most critical to estimate accurately and which intervention strategies (e.g., reducing β through social distancing, or increasing γ through improved treatment) are likely to be most effective in reducing transmission.

The normalised forward sensitivity index of R_0 with respect to a parameter θ is defined as

$$\gamma_{\theta}^{R_0} = \frac{\partial R_0}{\partial \theta} \times \frac{\theta}{R_0}. \quad (27)$$

For the SEIR expression in Eq. (25), direct calculation gives $\gamma_{\beta}^{R_0} = +1$ (indicating that a 1% increase in β produces exactly a 1% increase in R_0), while $\gamma_{\gamma}^{R_0} = -\gamma/(\gamma + \mu)$, which is negative and typically close to -1 in magnitude when $\mu \ll \gamma$ (as is typical for acute infections with disease timescales much shorter than the host demographic timescale). Figure 5 presents the sensitivity of R_0 to joint variation in β and γ , illustrating the threshold contour $R_0 = 1$ that separates parameter combinations leading to disease extinction from those leading to sustained transmission.

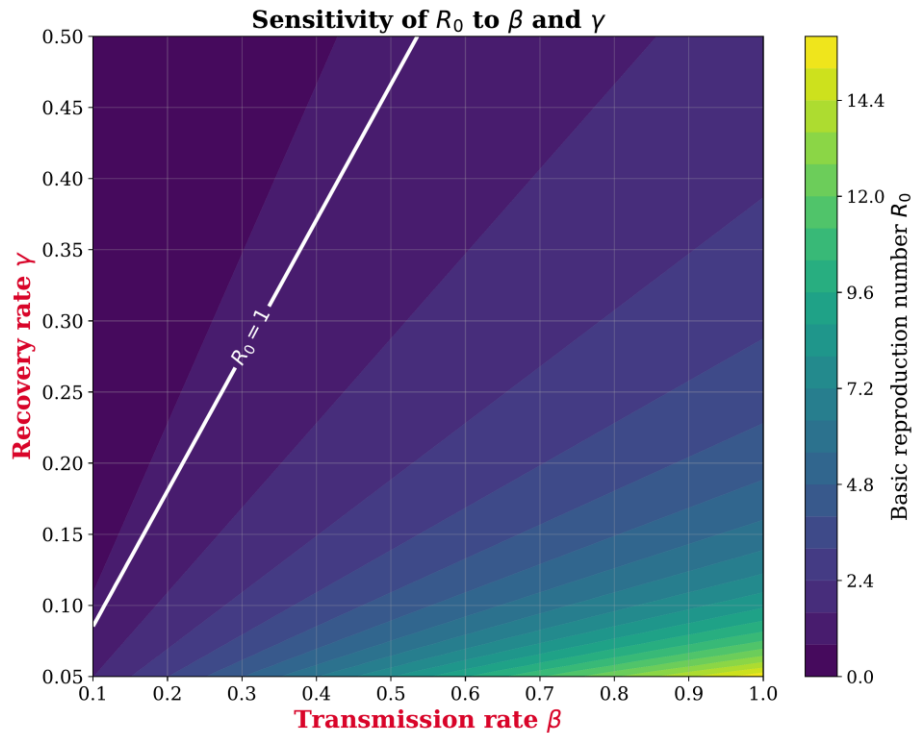


Figure 5: Sensitivity of the basic reproduction number R_0 to joint variation in the transmission rate β and recovery rate γ , for fixed $\sigma = 0.2$ and $\mu = 0.01$. The white contour marks the critical threshold $R_0 = 1$.

Table 3 summarises representative sensitivity indices reported across several recent extended compartmental models, illustrating the generally consistent finding that the transmission rate β and the latency-related parameter σ exert the strongest positive influence on R_0 , while recovery- and intervention-related parameters (quarantine rate, hospitalisation rate, vaccination rate) exert negative influence.

Table 3: Representative normalised sensitivity indices of R_0 reported in recent extended compartmental modelling studies (illustrative summary values)

Parameter	Symbol	Typical sensitivity index	Effect on R_0
Transmission rate	β	+1.00	Increases
Latency progression rate	σ	+0.10 to +0.35	Increases
Recovery rate	γ	−0.55 to −0.95	Decreases
Quarantine rate	δ	−0.20 to −0.60	Decreases
Vaccination rate	ν	−0.15 to −0.50	Decreases
Natural death rate	μ	small, variable	Mixed

6. Stochastic and Fractional-Order Extensions

6.1 Stochastic Differential Equation Formulations

Deterministic ODE compartmental models implicitly assume large population sizes in which demographic stochasticity is negligible; however, this assumption breaks down during the early stages of an outbreak, or in small, spatially isolated populations [15]. Recent literature has therefore increasingly incorporated stochastic effects, most commonly by reformulating the deterministic system as a system of Itô stochastic differential equations (SDEs) of the form

$$dX(t) = f(X(t)) dt + G(X(t)) dW(t), \quad (28)$$

where f is the deterministic drift term (identical to the right-hand side of the corresponding ODE system), G is a diffusion matrix capturing demographic or environmental noise, and $W(t)$ is a vector of independent Wiener processes. Such formulations permit rigorous analysis of extinction probabilities and quasi-stationary distributions, which are of particular interest for diseases with small R_0 or in populations of limited size.

6.2 Bayesian Parameter Estimation via MCMC

A closely related, increasingly standard practice is Bayesian estimation via Markov Chain Monte Carlo (MCMC), constructing a posterior

$$\pi(\theta | y) \propto L(y | \theta) \pi(\theta) \quad (29)$$

over the parameter vector θ (e.g., β, σ, γ), given observed case, hospitalisation, or death counts y , a Poisson or negative-binomial likelihood L , and a prior $\pi(\theta)$. Metropolis–Hastings and Hamiltonian Monte Carlo samplers provide posterior credible intervals for both parameters and derived quantities such as R_0 [16].

6.3 Fractional-Order Compartmental Models

A further significant methodological development has been the increasing use of fractional-order derivatives (in the Caputo, Riemann–Liouville, or Atangana–Baleanu sense) in place of classical integer-order derivatives, motivated by the ability of fractional operators to capture memory and hereditary effects in epidemic transmission — for instance, the dependence of current infection risk on the entire history of past exposure, rather than solely on the instantaneous state. A fractional-order SEIR model takes the form

$${}^C D_t^\alpha S(t) = \mu N - \beta \frac{SI}{N} - \mu S, \quad 0 < \alpha \leq 1, \quad (30)$$

with analogous fractional derivatives applied to the remaining compartments, where ${}^C D_t^\alpha$ denotes the Caputo fractional derivative of order α . The classical integer-order model is recovered in the limit $\alpha \rightarrow 1$ [17]. Semi-analytical solution of such fractional systems is commonly achieved via a fractional generalisation of the HPM (denoted FHPM or q -HPM), constructed analogously to

Section 3 but with the Caputo derivative replacing the classical time derivative in the homotopy construction.

7. Optimal Control Formulations

Many recent extended compartmental models incorporate one or more time-dependent control variables $u_i(t)$, representing public health interventions such as vaccination effort, quarantine intensity, treatment scale-up, or vector-control effort, and seek to determine the control strategy that optimises a specified objective functional, typically minimising cumulative infections and intervention cost simultaneously:

$$J[u_1, \dots, u_m] = \int_0^T \left[A_1 I(t) + \sum_{i=1}^m \frac{B_i}{2} u_i^2(t) \right] dt, \quad (31)$$

subject to the controlled compartmental dynamics and appropriate control constraints $0 \leq u_i(t) \leq u_i^{\max}$. Application of Pontryagin's Maximum Principle yields the existence of an optimal control satisfying a coupled two-point boundary value problem involving the state equations, adjoint (costate) equations

$$\frac{d\lambda_j}{dt} = -\frac{\partial H}{\partial x_j}, \quad (32)$$

where H is the associated Hamiltonian, and the characterisation of the optimal control via $\partial H / \partial u_i = 0$ [18]. Recent applications of this framework to malaria, Ebola, and other extended compartmental models have generally found that combined intervention strategies (e.g., simultaneous vector control and treatment scale-up for malaria) outperform single-intervention strategies at comparable total cost, and that the timing of intervention onset relative to the epidemic peak has a substantial effect on the achievable reduction in cumulative infections [19].

8. Integration of Machine Learning Approaches

A distinctive feature of the most recent literature (broadly, since 2020) has been the increasing integration of machine learning techniques alongside rather than as a replacement for classical mechanistic compartmental modelling. Three main integration strategies can be identified:

- **Pure data-driven forecasting**, using recurrent architectures such as LSTM networks trained directly on case-count time series without reference to a mechanistic model [14]; strong short-horizon accuracy but limited interpretability and poor performance under regime change.
- **Hybrid mechanistic–ML models**, in which a compartmental ODE model provides the dynamical structure while a machine-learning component estimates or forecasts a time-varying parameter such as $\beta(t)$, combining interpretability with flexibility.

- **ML-accelerated inference**, using surrogate neural networks to emulate expensive compartmental simulations, accelerating Bayesian parameter estimation.

Recent work combining LSTM forecasting with mechanistic SEIR-type structures for malaria has reported improved short-term accuracy relative to either component alone, particularly during rapid epidemic growth or decline [20].

9. Case Studies from Recent Applications

This section briefly surveys several representative disease-specific compartmental modelling studies illustrative of the broader methodological developments discussed above.

9.1 Norovirus: SEIAR Modelling

Norovirus is characterised by a short incubation period (12–48 hours), substantial asymptomatic infectious transmission, and pronounced seasonal forcing. Recent SEIAR-structured models incorporate an explicit asymptomatic compartment A and a seasonally forced transmission rate $\beta(t) = \beta_0(1 + \epsilon \cos(2\pi t/365))$. HPM-based solution of the resulting five-compartment system, validated against RK4, accurately reproduces the characteristic rapid rise and slower decline of norovirus outbreak curves [21].

9.2 Ebola Virus Disease: SEIQHR and SEIHRV Structures

Ebola modelling reflects a long, variable incubation period, high case-fatality ratio, and continued infectiousness of the deceased during unsafe burial practices [22]. SEIQHR structures, incorporating quarantine and hospitalisation, evaluate contact-tracing and case-isolation interventions, while SEIHRV extensions with a vaccination compartment V and saturated incidence $\beta SI/(1 + \alpha I)$ model ring-vaccination campaigns such as those deployed during the 2018–2020 Kivu outbreak in the Democratic Republic of the Congo.

9.3 Nipah Virus: SEIHR Modelling

Nipah virus, transmitted from fruit bat reservoirs via intermediate hosts, has been modelled using SEIHR structures with an explicit zoonotic spillover term alongside a hospitalisation compartment reflecting its high case-fatality ratio [23]. Sensitivity analysis consistently identifies human-to-human transmission, rather than spillover rate, as the dominant driver of R_0 once sustained transmission chains are established.

9.4 Malaria: Seven-Compartment SEIR–SEI Coupled Model

Malaria requires a coupled human–mosquito structure reflecting the two-host *Plasmodium* lifecycle [24]. A representative seven-compartment model couples a human SEIR(S) sub-model to a mosquito SEI sub-model via cross-infection terms. Recent treatments incorporate seasonal mosquito-abundance forcing, Bayesian MCMC parameter estimation, stochastic fadeout dynamics, Pontryagin optimal control of combined vector-control and treatment strategies, and

LSTM-based forecasting — jointly constituting one of the most methodologically comprehensive recent applications surveyed here.

9.5 Mumps: SEIR with Waning Immunity

Mumps modelling addresses waning vaccine-induced immunity, motivated by outbreaks in highly vaccinated university-age cohorts [25]. SEIRS structures with an explicit immunity-waning transition from R back to S at rate ω evaluate booster vaccination strategies targeted at adolescent and young-adult cohorts.

10. Discussion: Open Challenges and Future Directions

Despite substantial recent progress, several methodological and practical challenges remain open and are likely to constitute productive directions for future research.

Network and metapopulation structure. Most compartmental models, including those surveyed in Section 9, assume homogeneous mixing. Real populations exhibit substantial contact heterogeneity, and network-structured and metapopulation extensions that represent this heterogeneity remain comparatively underdeveloped in terms of rigorous stability and bifurcation theory.

Real-time data assimilation. Most applications reviewed here fit parameters retrospectively to a completed outbreak dataset. Genuine real-time forecasting, with continuous re-estimation subject to reporting delays and evolving pathogen characteristics, remains challenging; ensemble Kalman filtering and sequential Monte Carlo techniques from numerical weather prediction represent a promising but underused direction.

Identifiability and parameter estimation. Extended compartmental models often possess a parameter count large relative to the information content of available surveillance data, raising identifiability concerns that profile likelihood and Bayesian posterior correlation analysis only partially address.

Convergence theory for semi-analytical methods. While convergence proofs for HPM and related methods exist for several specific compartmental structures (Section 3), a general convergence theory spanning the full range of extended structures now in use remains incomplete.

Hybrid mechanistic–machine-learning frameworks. Early hybrid approaches (Section 8) show promise, but principled uncertainty quantification that jointly propagates mechanistic and machine-learning uncertainty remains at an early stage of development.

Conclusion

The mathematical modelling of biological systems, and of infectious disease transmission in particular, has undergone substantial methodological development in recent years, encompassing increasingly elaborate compartmental structures tailored to specific pathogens, refined semi-

analytical solution techniques such as the Homotopy Perturbation Method with accompanying convergence theory, comprehensive stability and bifurcation analysis grounded in the next-generation matrix and Lyapunov function methodology, systematic sensitivity analysis of the basic reproduction number, stochastic and fractional-order extensions capturing demographic noise and memory effects, optimal control formulations for intervention design, and, most recently, the integration of machine learning techniques for parameter estimation and short-term forecasting. Representative case studies spanning Norovirus, Ebola virus disease, Nipah virus, malaria, and mumps illustrate the practical application of this methodological toolkit to a diverse range of contemporary public health problems. Continued progress in network-structured modelling, real-time data assimilation, and principled hybrid mechanistic–machine-learning frameworks is likely to further extend the practical impact of mathematical modelling in supporting infectious disease surveillance and control policy in the years ahead.

References

1. Kermack, W. O., McKendrick, A. G. (1927). A contribution to the mathematical theory of epidemics. *Proceedings of the Royal Society A*, 115(772), 700–721.
2. Anderson, R. M., May, R. M. (1991). *Infectious Diseases of Humans: Dynamics and Control*. Oxford University Press, Oxford.
3. Murray, J. D. (2002). *Mathematical Biology I: An Introduction* (3rd ed.). Springer, New York.
4. Giordano, G., Blanchini, F., Bruno, R., Colaneri, P., Di Filippo, A., Di Matteo, A., Colaneri, M. (2020). Modelling the COVID-19 epidemic and implementation of population-wide interventions in Italy. *Nature Medicine*, 26(6), 855–860.
5. Rahman, B., Sadraddin, E., Porreca, A. (2021). The basic reproduction number of SARS-CoV-2: A systematic review and meta-analysis. *Reviews in Medical Virology*, 31(6), e2244.
6. Hethcote, H. W. (2000). The mathematics of infectious diseases. *SIAM Review*, 42(4), 599–653.
7. Brauer, F., Castillo-Chavez, C., Feng, Z. (2019). *Mathematical Models in Epidemiology*. Springer, New York.
8. He, J. H. (1999). Homotopy perturbation technique. *Computer Methods in Applied Mechanics and Engineering*, 178(3–4), 257–262.
9. He, J. H. (2003). Homotopy perturbation method: a new nonlinear analytical technique. *Applied Mathematics and Computation*, 135(1), 73–79.
10. Adomian, G. (1994). *Solving Frontier Problems of Physics: The Decomposition Method*. Kluwer Academic Publishers, Dordrecht.

11. He, J. H. (2007). Variational iteration method—some recent results and new interpretations. *Journal of Computational and Applied Mathematics*, 207(1), 3–17.
12. van den Driessche, P., Watmough, J. (2002). Reproduction numbers and sub-threshold endemic equilibria for compartmental models of disease transmission. *Mathematical Biosciences*, 180(1–2), 29–48.
13. Biazar, J. (2009). Solution of the epidemic model by Adomian decomposition method. *Applied Mathematics and Computation*, 173(2), 1101–1106.
14. Chimmula, V. K. R., Zhang, L. (2020). Time series forecasting of COVID-19 transmission in Canada using LSTM networks. *Chaos, Solitons & Fractals*, 135, 109864.
15. Allen, L. J. S. (2017). A primer on stochastic epidemic models: Formulation, numerical simulation, and analysis. *Infectious Disease Modelling*, 2(2), 128–142.
16. Diekmann, O., Heesterbeek, H., Britton, T. (2012). *Mathematical Tools for Understanding Infectious Disease Dynamics*. Princeton University Press, Princeton.
17. Diethelm, K. (2010). *The Analysis of Fractional Differential Equations*. Springer, Berlin.
18. Lenhart, S., Workman, J. T. (2007). *Optimal Control Applied to Biological Models*. Chapman and Hall/CRC, Boca Raton.
19. Agosto, F. B., Khan, M. A. (2018). Optimal control strategies for dengue transmission in Pakistan. *Mathematical Biosciences*, 305, 102–121.
20. Ray, E. L., Reich, N. G. (2018). Prediction of infectious disease epidemics via weighted density ensembles. *PLOS Computational Biology*, 14(2), e1005910.
21. Steele, J. A., Wu, H. (2019). Norovirus outbreak dynamics: a compartmental modelling and forecasting perspective. *Epidemiology and Infection*, 147, e158.
22. Legrand, J., Grais, R. F., Boelle, P. Y., Valleron, A. J., Flahault, A. (2007). Understanding the dynamics of Ebola epidemics. *Epidemiology and Infection*, 135(4), 610–621.
23. Luby, S. P., Gurley, E. S. (2012). Epidemiology of henipavirus disease in humans. *Current Topics in Microbiology and Immunology*, 359, 25–40.
24. Mandal, S., Sarkar, R. R., Sinha, S. (2011). Mathematical models of malaria: a review. *Malaria Journal*, 10, 202.
25. Jones, G. W., Swallow, W. H. (2012). Modelling mumps resurgence in vaccinated populations: waning immunity and outbreak dynamics. *Epidemics*, 4(3), 129–137.

CLIMATE CHANGE AND BIODIVERSITY CONSERVATION: CONTEMPORARY CHALLENGES FOR THE LIVING WORLD

Vivek Kumar and Sorabh

Department of Zoology,

School of Sciences, IFTM University, Moradabad, Uttar Pradesh (India)

Corresponding author E-mail: drvivek15010@gmail.com, sorabhchaudhary74@gmail.com

Abstract

Climate change is one of the most critical environmental challenges affecting biodiversity, ecosystem stability, and the sustainability of life on Earth. Increasing global temperatures, altered precipitation patterns, habitat degradation, and extreme climatic events are significantly influencing species distribution, ecological interactions, and extinction risks. This chapter explores the complex relationship between climate change and biodiversity conservation, highlighting major drivers, ecological consequences, and contemporary conservation strategies. The impacts of climate change on terrestrial, freshwater, and marine ecosystems are discussed, along with the role of modern technologies such as remote sensing, artificial intelligence, environmental DNA, and genomic approaches in biodiversity monitoring. Furthermore, the chapter emphasizes nature-based solutions, ecosystem restoration, and sustainable management practices for enhancing ecological resilience. Effective conservation under changing climatic conditions requires integrated approaches involving scientific innovation, policy frameworks, and community participation. Strengthening global cooperation and adopting climate-resilient strategies are essential for safeguarding biodiversity and maintaining ecosystem services for future generations.

Keywords: Climate Change, Biodiversity Conservation, Ecosystem Resilience, Species Extinction, Nature-Based Solutions.

1. Introduction

Climate change has emerged as one of the most significant environmental challenges of the twenty-first century, influencing ecological processes, species distribution patterns, and the functioning of ecosystems worldwide. The Earth's climate system has undergone substantial alterations due to increasing concentrations of greenhouse gases, primarily carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O), resulting from anthropogenic activities such as fossil fuel combustion, industrial development, deforestation, and intensive agriculture. These changes have accelerated global warming, altered precipitation regimes, and increased the frequency of extreme climatic events, thereby creating profound consequences for biological diversity and ecosystem stability (IPCC, 2023).

Biodiversity, encompassing genetic diversity, species diversity, and ecosystem diversity, plays a fundamental role in maintaining ecological balance and supporting essential ecosystem services, including carbon sequestration, nutrient cycling, pollination, water regulation, and food security. However, rapid climatic changes are disrupting natural ecosystems by modifying habitat conditions, shifting species ranges, affecting reproductive cycles, and increasing extinction risks. According to the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services (IPBES), climate change is among the major direct drivers responsible for accelerating global biodiversity decline, along with land-use change, pollution, and overexploitation of natural resources (IPBES, 2019).

The impacts of climate change on biodiversity are complex and vary across ecosystems. Rising temperatures have forced many terrestrial and aquatic species to shift their geographical distributions toward suitable climatic conditions. Changes in seasonal patterns have also influenced phenological events such as flowering, migration, breeding, and hibernation, creating ecological mismatches among interacting species (Parmesan & Yohe, 2003). Marine ecosystems are particularly vulnerable, where ocean warming and acidification threaten coral reefs, fisheries, and other marine organisms dependent on sensitive environmental conditions (Hoegh-Guldberg *et al.*, 2019). Similarly, terrestrial ecosystems such as forests, grasslands, and wetlands face increasing risks due to habitat fragmentation, drought stress, and climate-induced disturbances.

Conservation biology has therefore entered a new era where traditional biodiversity protection approaches must be integrated with climate adaptation and mitigation strategies. Climate-resilient conservation planning, ecosystem restoration, protected area management, and technological innovations such as remote sensing, ecological modelling, and genomic approaches are becoming essential tools for safeguarding biodiversity under changing climatic conditions (Pörtner *et al.*, 2022). Nature-based solutions, including forest restoration, wetland conservation, and sustainable land management, provide opportunities to simultaneously address climate change mitigation and biodiversity conservation.

2. Understanding Climate Change: Causes and Global Impacts

Concept and Overview of Climate Change: Climate change refers to long-term variations in Earth's climatic conditions, including changes in temperature, precipitation patterns, atmospheric composition, and the frequency and intensity of extreme weather events. Although natural climatic fluctuations have occurred throughout geological history, the present rate of climate change is largely influenced by anthropogenic activities. The rapid increase in greenhouse gas concentrations, particularly carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O), has intensified the natural greenhouse effect, resulting in global warming and widespread environmental alterations (IPCC, 2023).

The Earth's climate system is regulated by complex interactions among the atmosphere, oceans, land surface, and biosphere. Any disturbance in these components can influence global climatic patterns. Current observations indicate significant increases in global temperature, ocean warming, glacier retreat, and changes in precipitation distribution, highlighting the urgency of addressing climate change impacts on natural and human systems (IPCC, 2023).

2.1 Major Causes and Drivers of Climate Change

- **Greenhouse Gas Emissions and Global Warming:** The primary driver of contemporary climate change is the enhanced accumulation of greenhouse gases in the atmosphere. Carbon dioxide released from fossil fuel combustion, industrial processes, and transportation represents the largest contributor to global warming. Methane emissions from agriculture, livestock production, and waste management, along with nitrous oxide emissions from nitrogen-based fertilizers, further accelerate atmospheric warming (Friedlingstein *et al.*, 2023).
- **Deforestation and Land-Use Change:** Forests act as major carbon sinks by absorbing and storing atmospheric carbon dioxide. However, deforestation, forest degradation, and conversion of natural ecosystems into agricultural and urban landscapes reduce carbon sequestration capacity and increase greenhouse gas emissions. Land-use changes also result in habitat fragmentation and biodiversity loss, further reducing ecosystem resilience against climate change (Canadell *et al.*, 2021).
- **Industrialization, Urbanization, and Unsustainable Development:** Rapid industrial growth, increasing energy consumption, and urban expansion have significantly altered natural ecosystems. The extensive use of fossil fuels for electricity generation, transportation, and manufacturing has increased atmospheric pollution and contributed to climate instability. Urban areas experience additional climate-related challenges, including the urban heat island effect, increased energy demand, and vulnerability to extreme weather events.

2.2 Global Impacts of Climate Change

- **Rising Temperature and Extreme Weather Events:** Global warming has resulted in a continuous increase in average surface temperatures. Higher temperatures are associated with increased frequency and intensity of heatwaves, droughts, wildfires, and storms. These extreme events negatively affect ecosystems, agriculture, water resources, and human communities (IPCC, 2023).
- **Impacts on Terrestrial and Aquatic Ecosystems:** Climate change significantly influences ecosystem structure and functioning. Rising temperatures, altered rainfall patterns, and habitat changes affect species distribution, reproduction, and survival. Many

terrestrial species are shifting their geographical ranges toward cooler regions, while aquatic organisms are threatened by ocean warming, acidification, and declining habitat quality (Bellard *et al.*, 2012). Marine ecosystems, particularly coral reefs, are highly sensitive to climate change. Increased ocean temperature causes coral bleaching, while ocean acidification affects marine organisms that depend on calcium carbonate structures (Hoegh-Guldberg *et al.*, 2019).

- **Effects on Human Society and Sustainable Development:** Climate change has direct and indirect consequences for human populations through impacts on food security, water availability, health, and economic stability. Reduced agricultural productivity, emergence of climate-related diseases, and displacement caused by sea-level rise and extreme climatic events represent major challenges for sustainable development.

2.3 Need for Climate Change Mitigation and Adaptation Strategies

Addressing climate change requires integrated approaches involving emission reduction, ecosystem restoration, sustainable resource management, and climate adaptation planning. Renewable energy development, forest conservation, sustainable agriculture, and nature-based solutions are essential strategies for reducing climate risks and protecting biodiversity. International cooperation through frameworks such as the Paris Agreement and Sustainable Development Goals provides a pathway toward climate-resilient development.

3. Climate Change Effects on Biodiversity

Biodiversity, encompassing the diversity of genes, species, and ecosystems, is fundamental to maintaining ecological balance and supporting ecosystem services essential for human well-being. Climate change has become one of the most significant threats to biodiversity by altering environmental conditions beyond the adaptive capacity of many organisms. Rising temperatures, changing precipitation patterns, increasing atmospheric carbon dioxide concentrations, and the growing frequency of extreme weather events are reshaping ecosystems across the globe. These climatic changes influence species distribution, population dynamics, ecological interactions, and ecosystem functioning, leading to biodiversity loss at local, regional, and global scales (IPBES, 2019; IPCC, 2023).

- **Changes in Species Distribution and Migration:** Climate change is causing significant shifts in the geographical distribution of plant and animal species. As environmental conditions become unsuitable in their native habitats, many species migrate toward higher latitudes or elevations where climatic conditions remain favorable. While highly mobile species may successfully expand their ranges, less mobile and habitat-specialist organisms often experience population declines due to limited dispersal ability (Pecl *et al.*, 2017).

- **Habitat Loss and Ecosystem Degradation:** Climate change accelerates habitat degradation by modifying temperature, moisture availability, and disturbance regimes. Increased drought frequency, wildfires, floods, and storms damage forests, grasslands, wetlands, and coastal ecosystems, reducing habitat quality for numerous species. Habitat fragmentation further restricts species movement and gene flow, making populations more vulnerable to extinction (Bellard *et al.*, 2012). Coral reef ecosystems are among the most affected habitats. Elevated sea surface temperatures trigger mass coral bleaching events, while ocean acidification reduces coral calcification, threatening one of the world's richest biodiversity hotspots (Hoegh-Guldberg *et al.*, 2019).
- **Phenological Changes and Ecological Mismatches:** Climate change has altered the timing of biological events, a phenomenon known as phenological change. Earlier flowering in plants, earlier breeding in birds, altered insect emergence, and shifts in migration timing have been widely documented. These changes often create ecological mismatches between interacting species, such as flowering plants and their pollinators or predators and prey, reducing reproductive success and ecosystem productivity (Parmesan & Yohe, 2003).
- **Increased Risk of Species Extinction:** Species with restricted geographic distributions, narrow ecological niches, or low adaptive capacity are particularly vulnerable to climate change. Amphibians, polar mammals, coral reef organisms, and many endemic plant species face increased extinction risks due to rapid environmental changes. Climate change acts synergistically with habitat destruction, pollution, invasive species, and overexploitation, accelerating biodiversity loss (Urban, 2015).
- **Impacts on Terrestrial, Freshwater, and Marine Biodiversity:** Forests, grasslands, tundra, and desert ecosystems are experiencing changes in vegetation composition, productivity, and species diversity. Increased drought stress, pest outbreaks, and forest fires reduce ecosystem resilience and carbon sequestration capacity. Rivers, lakes, and wetlands are highly sensitive to temperature increases and altered rainfall patterns. Reduced water availability, declining oxygen levels, and increased pollution threaten freshwater fish, amphibians, and aquatic invertebrates, many of which have limited dispersal opportunities. Marine biodiversity is severely affected by ocean warming, acidification, deoxygenation, and sea-level rise. Coral reefs, mangroves, seagrass beds, and polar marine ecosystems are among the most vulnerable habitats. Changes in ocean circulation also affect fish migration and global fisheries, with significant ecological and economic consequences (Pörtner *et al.*, 2019).
- **Effects on Ecosystem Functions and Services:** Biodiversity underpins essential ecosystem services that support human societies. Climate-induced biodiversity loss

reduces ecosystem productivity and resilience, affecting pollination, nutrient cycling, soil formation, carbon storage, water purification, and climate regulation. The degradation of these services threatens agriculture, food security, public health, and sustainable development (IPBES, 2019).

4. Climate Change and Species Extinction Risk

Climate change has become one of the most significant drivers of species extinction in the Anthropocene. Rapid increases in global temperatures, altered precipitation patterns, sea-level rise, ocean acidification, and the growing frequency of extreme weather events are transforming ecosystems faster than many species can adapt. While biodiversity has always experienced natural fluctuations, the current rate of climate change, combined with habitat destruction, pollution, invasive species, and overexploitation, has substantially increased extinction risks across terrestrial, freshwater, and marine ecosystems (IPCC, 2023; IPBES, 2019).

- **Climate-Induced Drivers of Species Extinction:** Climate change affects species survival through multiple direct and indirect mechanisms. Rising temperatures alter physiological processes, reproductive success, and survival rates, particularly for organisms with narrow thermal tolerances. Changes in rainfall patterns reduce water availability and modify habitat conditions, while prolonged droughts and heatwaves increase mortality in both plants and animals. Similarly, extreme climatic events such as wildfires, floods, cyclones, and storms can rapidly destroy habitats and eliminate local populations (Urban, 2015). Many species respond by shifting their geographical ranges toward higher latitudes or elevations. However, organisms with limited dispersal ability, restricted distributions, or specialized habitat requirements often fail to migrate successfully, making them highly vulnerable to extinction. Island species, alpine flora, amphibians, coral reef organisms, and polar wildlife are among the taxa facing the greatest climate-related risks (Bellard *et al.*, 2012).
- **Vulnerable Species and Ecosystems:** Climate-sensitive ecosystems, including coral reefs, Arctic tundra, mangrove forests, wetlands, and mountain ecosystems, are experiencing rapid biodiversity declines. Coral reefs, for instance, are severely affected by ocean warming and acidification, leading to widespread coral bleaching and habitat degradation. Likewise, melting sea ice threatens Arctic species such as polar bears and seals, while changing hydrological conditions negatively affect freshwater biodiversity (Hoegh-Guldberg *et al.*, 2019). Species with small population sizes, low genetic diversity, and specialized ecological niches possess limited adaptive capacity. Climate change also disrupts ecological interactions, including pollination, seed dispersal, predator–prey relationships, and host–parasite dynamics, further increasing extinction risks (Parmesan & Yohe, 2003).

5. Conservation Strategies under Changing Climate Conditions

Climate change has intensified the need for innovative and adaptive conservation strategies to protect biodiversity and maintain ecosystem resilience. Traditional conservation approaches, which primarily focused on protecting species within fixed geographical boundaries, are often insufficient under rapidly changing climatic conditions. As species shift their distribution ranges and ecosystems undergo structural and functional changes, conservation efforts must incorporate climate adaptation, ecological restoration, and sustainable resource management. Climate-resilient conservation aims to enhance the capacity of species and ecosystems to withstand environmental changes while ensuring the continued provision of ecosystem services (IPCC, 2023; IPBES, 2019).

- **Climate-Smart Conservation Approaches:** One of the most effective strategies is the development of climate-smart protected areas, where conservation planning considers future climate scenarios in addition to present biodiversity patterns. Expanding protected area networks and improving habitat connectivity through ecological corridors facilitate species migration and genetic exchange, allowing populations to adapt to changing environmental conditions (Heller & Zavaleta, 2009). Landscape-level conservation, which integrates forests, wetlands, grasslands, and agricultural ecosystems, also enhances ecological resilience.
- **Technological Innovations and Community Participation:** Recent technological advances have significantly improved biodiversity conservation. Remote sensing, Geographic Information Systems (GIS), drones, environmental DNA (eDNA), artificial intelligence (AI), and machine learning enable continuous monitoring of ecosystems, species distributions, and habitat changes. These technologies support early detection of biodiversity threats and improve decision-making for conservation planning (Pimm *et al.*, 2015).
- **Policy Frameworks and Future Directions:** International agreements such as the Convention on Biological Diversity (CBD), the Paris Agreement, and the United Nations Sustainable Development Goals (SDGs) provide global frameworks for integrating biodiversity conservation with climate action. Effective implementation requires strong governance, adequate financial investment, interdisciplinary research, and international collaboration.

6. Role of Modern Technologies in Biodiversity Conservation

The rapid advancement of science and technology has transformed biodiversity conservation by providing innovative tools for monitoring ecosystems, assessing species diversity, and supporting evidence-based conservation planning. Traditional field-based surveys remain fundamental; however, they are often time-consuming, expensive, and limited in spatial and

temporal coverage. Modern technologies, including remote sensing, Geographic Information Systems (GIS), artificial intelligence (AI), environmental DNA (eDNA), genomics, and big data analytics, have significantly improved the accuracy and efficiency of biodiversity assessment and management. These technologies enable researchers and policymakers to detect environmental changes, predict species responses to climate change, and develop effective conservation strategies (Pimm *et al.*, 2015; IPBES, 2019).

- **Remote Sensing and Geographic Information Systems (GIS):** Satellite remote sensing and GIS have become indispensable tools for biodiversity monitoring. High-resolution satellite imagery allows scientists to map land-use changes, monitor deforestation, assess habitat fragmentation, and evaluate ecosystem health over large geographic areas. GIS integrates spatial and ecological data to identify biodiversity hotspots, design protected areas, and prioritize conservation actions. These technologies also facilitate long-term monitoring of vegetation cover, wetlands, coral reefs, and wildlife habitats under changing climatic conditions (Pettorelli *et al.*, 2014).
- **Artificial Intelligence, Genomics, and Environmental DNA:** Artificial intelligence (AI) and machine learning algorithms are increasingly used to analyze large ecological datasets, identify species from camera-trap images and acoustic recordings, predict species distributions, and forecast climate-driven biodiversity changes. These tools improve conservation decision-making by rapidly processing complex environmental information (Christin *et al.*, 2019). Genomic technologies have enhanced conservation genetics by identifying genetic diversity, detecting inbreeding, and supporting breeding programs for endangered species. Similarly, environmental DNA (eDNA) analysis enables the detection of species from traces of DNA present in soil, water, or air without direct observation or capture. eDNA has become a powerful, non-invasive method for monitoring rare, invasive, and endangered species while reducing disturbance to natural populations (Thomsen & Willerslev, 2015).
- **Big Data, Citizen Science, and Future Perspectives:** The integration of big data analytics, cloud computing, and global biodiversity databases has improved the capacity to assess biodiversity trends at regional and global scales. Open-access platforms such as the Global Biodiversity Information Facility (GBIF) and the IUCN Red List provide valuable datasets for ecological research and conservation planning. Citizen science initiatives, supported by smartphone applications and online platforms, further contribute to biodiversity monitoring by engaging the public in recording species observations.

7. Nature-Based Solutions and Sustainable Approaches

Nature-based Solutions (NbS) are increasingly recognized as effective strategies for addressing climate change, conserving biodiversity, and promoting sustainable development. The

International Union for Conservation of Nature (IUCN) defines NbS as actions that protect, sustainably manage, and restore natural or modified ecosystems while simultaneously providing environmental, social, and economic benefits (IUCN, 2020). Unlike conventional engineering-based approaches, nature-based solutions harness ecological processes to enhance ecosystem resilience, mitigate climate change, and support human well-being. As climate change accelerates biodiversity loss, integrating NbS with sustainable land and water management has become a global conservation priority (IPCC, 2023).

- **Ecosystem Restoration and Climate Mitigation:** Ecosystem restoration is one of the most effective nature-based approaches for conserving biodiversity while reducing greenhouse gas concentrations. Restoration of degraded forests, wetlands, grasslands, mangroves, and peatlands improves carbon sequestration, enhances habitat quality, and restores ecosystem functions. Forest restoration increases biomass and soil carbon storage, while wetlands and peatlands act as long-term carbon sinks and improve water regulation. Coastal ecosystems such as mangroves, salt marshes, and seagrass meadows—collectively known as blue carbon ecosystems—store substantial amounts of carbon and provide protection against coastal erosion and storm surges (UNEP, 2021; IPCC, 2023).
- **Sustainable Agriculture and Community-Based Conservation:** Sustainable agricultural practices play a crucial role in balancing food production with biodiversity conservation. Agroforestry, organic farming, conservation agriculture, integrated pest management, and efficient water-use practices reduce environmental degradation while enhancing soil fertility and ecosystem resilience. These approaches promote pollinator diversity, improve nutrient cycling, and reduce dependence on synthetic fertilizers and pesticides (FAO, 2022).

8. Challenges and Future Perspectives

- **Current Challenges in Biodiversity Conservation:** Despite significant advances in climate science and conservation biology, biodiversity conservation continues to face numerous challenges under changing climatic conditions. Climate change interacts with other anthropogenic pressures, including habitat destruction, pollution, invasive alien species, overexploitation of natural resources, and land-use change, creating cumulative impacts that threaten ecosystem stability and species survival. These multiple stressors reduce ecosystem resilience and increase the vulnerability of already threatened species (IPBES, 2019; IPCC, 2023). One of the major challenges is the uncertainty associated with climate projections and species responses. Different species exhibit varying adaptive capacities, making it difficult to accurately predict future distribution patterns and extinction risks. Additionally, limited ecological data, inadequate biodiversity

monitoring, and insufficient long-term datasets restrict the development of effective conservation strategies (Urban, 2015). Financial constraints, weak governance, and the lack of coordination among conservation agencies further hinder the implementation of climate adaptation programs, particularly in developing countries.

- **Future Perspectives for Climate-Resilient Conservation:** Future biodiversity conservation requires an integrated and adaptive approach that combines scientific innovation, sustainable development, and international cooperation. Advances in artificial intelligence (AI), remote sensing, environmental DNA (eDNA), genomics, and ecological modeling are expected to improve biodiversity monitoring, species distribution forecasting, and conservation planning. These technologies will enable more accurate identification of climate refugia, biodiversity hotspots, and priority conservation areas (Pimm *et al.*, 2015).

Conclusion

Climate change represents one of the greatest challenges to biodiversity conservation and the sustainability of the living world. Rising temperatures, habitat degradation, extreme climatic events, and changing ecological interactions are accelerating species decline and threatening ecosystem stability. The complex relationship between climate change and biodiversity loss highlights the need for integrated conservation approaches that combine ecological research, technological innovation, policy interventions, and community participation. Modern tools such as remote sensing, artificial intelligence, genomics, and environmental DNA analysis are enhancing our ability to monitor and protect biodiversity. Furthermore, nature-based solutions, ecosystem restoration, and climate-smart conservation strategies provide effective pathways for improving ecosystem resilience. Addressing these challenges requires global cooperation, sustainable development practices, and long-term commitment toward climate mitigation and adaptation. Protecting biodiversity under changing climatic conditions is essential not only for maintaining ecological balance but also for ensuring human well-being and planetary sustainability.

References

1. Bellard, C., Bertelsmeier, C., Leadley, P., Thuiller, W., & Courchamp, F. (2012). Impacts of climate change on the future of biodiversity. *Ecology Letters*, 15(4), 365–377. <https://doi.org/10.1111/j.1461-0248.2011.01736.x>
2. Canadell, J. G., Monteiro, P. M. S., Costa, M. H., Cotrim da Cunha, L., Cox, P. M., Eliseev, A. V., *et al.* (2021). Global carbon and other biogeochemical cycles and feedbacks. In *Climate Change 2021: The Physical Science Basis*. Cambridge University Press.

3. Christin, S., Hervet, É., & Lecomte, N. (2019). Applications for deep learning in ecology. *Methods in Ecology and Evolution*, 10(10), 1632–1644. <https://doi.org/10.1111/2041-210X.13256>
4. Food and Agriculture Organization. (2022). *The State of the World's Forests 2022: Forest pathways for green recovery and building inclusive, resilient and sustainable economies*. FAO.
5. Friedlingstein, P., O'Sullivan, M., Jones, M. W., Andrew, R. M., Bakker, D. C. E., Hauck, J., et al. (2023). Global carbon budget 2023. *Earth System Science Data*, 15, 5301–5369. <https://doi.org/10.5194/essd-15-5301-2023>
6. Heller, N. E., & Zavaleta, E. S. (2009). Biodiversity management in the face of climate change: A review of 22 years of recommendations. *Biological Conservation*, 142(1), 14–32. <https://doi.org/10.1016/j.biocon.2008.10.006>
7. Hoegh-Guldberg, O., Poloczanska, E. S., Skirving, W., & Dove, S. (2019). Coral reef ecosystems under climate change and ocean acidification. *Frontiers in Marine Science*, 6, 367. <https://doi.org/10.3389/fmars.2019.00367>
8. International Union for Conservation of Nature. (2020). *Global standard for nature-based solutions: A user-friendly framework for the verification, design and scaling up of NbS*. IUCN.
9. IPBES. (2019). *Global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services*. IPBES Secretariat.
10. IPCC. (2023). *Climate change 2023: Synthesis report. Contribution of Working Groups I, II and III to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. IPCC.
11. Parmesan, C., & Yohe, G. (2003). A globally coherent fingerprint of climate change impacts across natural systems. *Nature*, 421, 37–42. <https://doi.org/10.1038/nature01286>
12. Pecl, G. T., Araújo, M. B., Bell, J. D., Blanchard, J., Bonebrake, T. C., Chen, I.-C., Clark, T. D., Colwell, R. K., Danielsen, F., Evengård, B., Falconi, L., Ferrier, S., Frusher, S., Garcia, R. A., Griffis, R. B., Hobday, A. J., Janion-Scheepers, C., Jarzyna, M. A., Jennings, S., ... Williams, S. E. (2017). Biodiversity redistribution under climate change: Impacts on ecosystems and human well-being. *Science*, 355(6332), eaai9214. <https://doi.org/10.1126/science.aai9214>
13. Pettorelli, N., Laurance, W. F., O'Brien, T. G., Wegmann, M., Nagendra, H., & Turner, W. (2014). Satellite remote sensing for applied ecologists: Opportunities and challenges. *Journal of Applied Ecology*, 51(4), 839–848. <https://doi.org/10.1111/1365-2664.12261>

14. Pimm, S. L., Jenkins, C. N., Abell, R., Brooks, T. M., Gittleman, J. L., Joppa, L. N., Raven, P. H., Roberts, C. M., & Sexton, J. O. (2015). The biodiversity of species and their rates of extinction, distribution, and protection. *Science*, 344(6187), 1246752. <https://doi.org/10.1126/science.1246752>
15. Pörtner, H. O., Roberts, D. C., Adams, H., Adler, C., Aldunce, P., Ali, E., Begum, R. A., Betts, R., Kerr, R. B., Biesbroek, R., Birkmann, J., Bowen, K., Castellanos, E., Cissé, G., Constable, A., Cramer, W., Dodman, D., Eriksen, S. H., Fischlin, A., & ... Poloczanska, E. (2022). *Climate change 2022: Impacts, adaptation and vulnerability*. Cambridge University Press.
16. Thomsen, P. F., & Willerslev, E. (2015). Environmental DNA—An emerging tool in conservation for monitoring past and present biodiversity. *Biological Conservation*, 183, 4–18. <https://doi.org/10.1016/j.biocon.2014.11.019>
17. United Nations Environment Programme. (2021). *Becoming #GenerationRestoration: Ecosystem restoration for people, nature and climate*. UNEP.
18. Urban, M. C. (2015). Accelerating extinction risk from climate change. *Science*, 348(6234), 571–573. <https://doi.org/10.1126/science.aaa4984>

The Living World: Contemporary Perspectives in Life Sciences Volume II

(ISBN: 978-93-47587-68-9)

About Editors



Dr. Mahesh P. Patil, Ph.D. in Microbiology, is an Assistant Professor at R. C. Patel Arts, Commerce and Science College, Shirpur, with over 20 years of experience in teaching, research, and academic leadership. A recognized expert in probiotics and dairy microbiology, he has authored numerous research papers in reputed national and international journals and contributed to textbooks and reference books. He has guided many postgraduate students and actively participates in conferences, workshops, faculty development programmes, and professional bodies. As a resource person, he has delivered expert lectures on microbiology, probiotics, research methodology, and higher education. His research excellence and academic contributions have earned him several recognitions. He remains committed to scientific innovation, student mentorship, and advancing microbiological research development.



Dr. Vijayalaxami Sahebrao Shelke is the Head of the Department of Botany at R. C. Patel Arts, Commerce and Science College, Shirpur, Maharashtra. She holds a Ph.D. in Botany and postgraduate degrees in Botany and Environmental Science. Her research interests include medicinal plants, plant taxonomy, rhizosphere soil fungi, biodiversity conservation, herbal cosmetics, and sustainable agriculture. She has authored two books, several book chapters, and numerous research papers in reputed national and international journals. She serves as a certified Environmental and Green Auditor and Plant Authenticator and holds several granted and filed patents in agricultural and environmental technologies. Dr. Shelke has received recognition for her academic excellence, innovation, and research contributions. She is dedicated to promoting quality education, scientific research, environmental sustainability, biodiversity conservation, and innovation.



Mrs. Ashwini Chandrakant Patil, M.Sc. (Biotechnology), MH-SET Qualified, is an Assistant Professor in the Department of Biotechnology at R. C. Patel Arts, Commerce and Science College, Shirpur, Maharashtra, with over 11 years of teaching experience in undergraduate and postgraduate education. Her research interests include Microbiology, Environmental Biotechnology, Agricultural Biotechnology, Microbial Enzymes, Soil Health, Plant-Microbe Interactions, and Sustainable Biotechnological Innovations. She has authored and co-authored research papers, book chapters, textbooks, and edited academic books in the fields of biotechnology and life sciences. She actively participates in national and international conferences, faculty development programmes, and interdisciplinary research activities. She is dedicated to promoting quality teaching, scientific research, academic excellence, innovation, collaborative learning, student mentorship, sustainable scientific development, knowledge dissemination, institutional growth, effectively.



Dr. Alok Ranjan Sahu is an Assistant Professor of Botany at Vikash Degree College, Bargarh, Odisha, India. He holds dual Master's degrees in Botany and Biotechnology, an M.Phil. in Life Science, and a Ph.D. in Biotechnology from Sambalpur University. His research interests include plant molecular biology, plant biotechnology, medicinal plants, ethnobotany, phytochemistry, natural products, and plant-based therapeutics. He has published more than 80 research papers, authored two books, co-authored ten books, and holds seven Indian patents. Dr. Sahu serves as a reviewer and Editorial Board Member for over 36 national and international peer-reviewed journals and is a Fellow and Life Member of eight professional societies. He has received several prestigious awards for his outstanding contributions to research, innovation, and higher education.

